

Potassium Rate and Application Effect on Flue-Cured Tobacco

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ABSTRACT

Research was conducted at two locations in 2009 and 2010 to evaluate the effect of various K rates and application methods on the yield and quality of flue-cured tobacco (*Nicotiana tabacum* L.). Treatments included five rates of K from sulfate of potash magnesia (0–0–22): 0, 84, 140, 196, and 252 kg K₂O ha⁻¹ that were applied: broadcast 1 mo before transplanting, broadcast 1 wk before transplanting, banded at transplanting, and a split application with one-half rate banded at transplanting followed by one-half rate banded at layby. Tissue samples were collected throughout the season at three separate growth stages: layby, topping, and after curing. Tissue samples were analyzed for total alkaloid and reducing sugars, N, P, K, and Mg content. Soil samples were collected the same day as K fertilizer application from plots not receiving supplemental K. Data were subjected to ANOVA using the PROC GLM procedure in SAS. Treatment means were separated using Fisher's Protected LSD test at $p \leq 0.05$. Application method and timing had no effect on any measured parameters; furthermore, crop yield and quality was not affected by K rates >0 kg K₂O ha⁻¹ at three of four locations. It is likely that early broadcast applications of K₂O with current rate recommendations would only be of concern with combinations of conditions that included coarse soil textures, low K indices, and/or excessive leaching rainfall.

POTASSIUM FERTILIZER IS essential to the production of high yielding, high quality flue-cured tobacco, with a healthy crop typically requiring roughly 100 kg K₂O ha⁻¹ from the soil for optimum growth (Raper and McCants, 1967). Tobacco, in general, is considered to be a luxury consumer of K⁺ (Raper and McCants, 1967), and K application is often two to three times that needed to obtain maximum yield (Sims, 1985). Furthermore, K deficiencies have long been observed across a variety of soil types and growing conditions, and the overapplication of K fertilizer has been justified on the belief that deficiencies would be avoided and leaf and burn quality would improve (Collins and Hawks, 1993). Overapplication has traditionally been possible because of low input costs for fertilizers and liming materials; however, in recent years certain aspects of production have altered how tobacco producers address crop nutrition.

The first major issue is the cost of fertilizer inputs, specifically K, which has increased by 292% since the year 2000 (Huang, 2012). As a result of increasing K prices, overapplication of fertilizer is no longer an option and growers must become more efficient in making applications. Second, nearly 85% of all traditional tobacco producing soils have a high (Mehlich-3 P 61–120 g P m⁻³) or

very-high (Mehlich-3 P > 120 g P m⁻³) P index (Hardy et al., 2012; Smith, 2011). Years of overapplication of P have led to high P-indices and additional P is not required on most tobacco soils (Smith, 2011). Producers are now able to decouple N, P, and K, and move away from complete fertilizers which have traditionally been used. With independent applications of N and K producers now have more alternatives for sources and application methods of both nutrients.

With changing fertilizer demand, tobacco producers have the option of applying K in broadcast applications before transplanting, banded applications at or just after transplanting, or in banded applications split at transplanting and layby (roughly 4–6 wk after transplanting). Each method has advantages and disadvantages that must be considered.

Banded, sidedress applications between transplanting and layby are common in most flue-cured tobacco operations (Tso, 1990c). Banded applications of K after transplanting are common because the nutrient is placed near the root zone, thus improving use efficiency. Research conducted by Collins and Hawks (1993) shows a slight yield increase in band applications of K over broadcast applications. Band applications can be made anytime between transplanting and layby, with common applications being one or two bands at transplanting, one band 10 d after transplanting, or a split application of equal rates at transplanting and layby. Split applications of K were found to be more efficient than one single band application on soil types that are highly leachable (San Valentin et al., 1978).

Broadcast application of complete fertilizers on flue-cured tobacco is no longer a common practice due to the risk of fertilizer salt damage from N placement (Tso, 1990c; Collins and Hawks, 1993). As a result of alternative fertilizer programs and to improve

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Abbreviations: OTRS, Oxford Tobacco Research Station; UAN, urea ammonium nitrate; UCPRS, Upper Coastal Plain Research Station.

Table 1. Soil series, soil taxonomic class, soil pH, P index, and K index at each location.

Location	Soil series	Soil taxonomic class	pH†	P Index‡	K Index‡
UCPRS-09‡	Goldsboro loamy sand	fine-loamy, siliceous, subactive, thermic Aquic Paleudults	6.2	45.6	0.25
OTRS-09§	Helena sandy loam	fine, mixed, semiactive, thermic Aquic Hapludults	5.8	49.2	0.19
UCPRS1-10¶	Norfolk loamy sand	fine-loamy, kaolinitic, thermic Typic Kandiodults	5.8	162	0.30
UCPRS2-10#	Goldsboro loamy sand	fine-loamy, siliceous, subactive, thermic Aquic Paleudults	6.0	112.8	0.45

† Soil chemistry represents the upper 20 cm of the soil profile.

‡ Upper Coastal Plain Research Station in 2009.

§ Oxford Tobacco Research Station in 2009.

¶ Upper Coastal Plain Research Station in 2010.

Upper Coastal Plain Research Station in 2010.

operating efficiency, growers have begun to broadcast K before forming plant beds and in some cases before fumigation. Soil fumigants are typically applied in the raised bed about 3 to 5 wk before transplanting. Allowing for uniform application before bedding, K application must take place a significant amount of time before transplanting as well. In the time between K application and transplanting, there is potential for K to leach out of the rooting zone. The practice of broadcast application of K fertilizer is also an option for producers who do not apply soil fumigants. Potassium fertilizer can be broadcast and land can be prepared by creating beds just before transplanting. In both situations, the producers' intentions are to complete the K application before transplanting, therefore reducing workloads as the season progresses. The broadcast application of K fits very well with new fertilizer programs that apply N and K independently to reduce risk of salt injury.

Past research provides inadequate details as to when and how to apply K in different management systems. With newer, alternative fertilizer plans now being implemented by growers, information is needed to provide accurate K recommendations. The objective of this study was to determine if alternative application methods of K are acceptable in tobacco production, and to determine their effect on application rate.

METHODS AND MATERIALS

Field Procedures

The experiments were conducted at the Upper Coastal Plain Research Station (UCPRS) near Rocky Mount, NC, and the Oxford Tobacco Research Station (OTRS) in Oxford, NC, in 2009 and at two separate locations at the UCPRS in 2010 (designated as UCPRS-1 and UCPRS-2). An additional test site at OTRS in 2010 was lost to the disease tobacco mosaic virus (TMV). Test sites were selected based on soil type, soil texture, and proximity to major flue-cured tobacco production regions (Table 1). Individual plots were four rows wide by 12.19 m long, with all four rows treated and the two center rows harvested for yield and quality. Individual plant spacing was 55 by 122 cm. Cultivars used in this study were NC 71 (Goldleaf Seed Company, Hartsville, SC) at the UCPRS and NC 297 (Goldleaf Seed Company, Hartsville, SC) at the OTRS. Different cultivars were selected because of their popularity among producers (Fisher et al., 2009) and because of the need for specific disease resistance at both locations (Mila and Radcliff, 2011). Tobacco was produced using recommendations from the North Carolina Cooperative Extension Service, except for treatments imposed. Cured tobacco was weighed for yield and assigned a USDA

government grade. Each government grade has an associated grade index value which describes the leaf maturity and ripeness (Bowman et al., 1988). Additionally, each USDA grade has an associated monetary value; this value was multiplied by crop yield to establish overall crop value.

Field Conditions

Field conditions (soil series, soil taxonomic class, pH, P-Index, and K-Index) and monthly rainfall are described by location in Tables 1 and 2. Treatments included 0, 84, 140, 196, and 252 kg K₂O ha⁻¹. Each rate was applied either broadcast 1 mo before transplanting, broadcast 1 wk before transplanting, banded at planting, or banded with one-half rate at transplanting and one-half rate at layby. Potassium application timings were chosen to represent different crop management programs for producers. Applications made 1 mo before transplanting represent production systems in which producers would be applying a soil fumigant. Soil fumigants are applied roughly 1 mo before transplanting to avoid crop injury. Alternatively, applications made 1 wk before transplanting represent production systems where producers do not have a need to apply soil fumigants. Broadcast applications of K fertilizer occur before the formation of raised plant beds. Full rate applications made at transplanting as well as half-rate applications made at transplanting and half-rate applications made at layby are standard production practices which are common throughout tobacco-producing regions in the United States.

Transplanting dates, fertilizer application dates, and tissue sampling dates at each location are described in Table 3. Broadcast applications were made by hand before transplanting and

Table 2. Monthly total precipitation from pretransplant to harvest at each location.

Month	UCPRS-2009†	OTRS-2009‡	UCPRS1-2010§	UCPRS2-2010¶
	cm			
March	18.1	12.7	20.0	20.0
April	2.0	2.1	2.8	2.8
May	8.7	18.4	13.2	13.2
June	9.1	11.3	3.2	3.2
July	8.9	6.4	5.4	5.4
August	7.4	8.1	11.3	11.3
September	12.1	1.0	26.2	26.2
October	3.5	2.8	3.9	3.9
Total	69.8	62.8	86.0	86.0

† Upper Coastal Plain Research Station in 2009.

‡ Oxford Tobacco Research Station in 2009.

§ Upper Coastal Plain Research Station in 2010.

¶ Upper Coastal Plain Research Station in 2010.

Table 3. Dates for transplanting, fertilizer application, and tissue sampling at each location.

Location	Transplanting	Fertilizer application				Tissue sampling		
		Broadcast 1 mo	Broadcast 1 wk	Banded at transplanting	Banded at layby	Layby	Topping	After curing
UCPRS-09†	29 Apr.	30 Mar.	22 Apr.	29 Apr.	18 June	18 June	16 July	15 Sept.
OTRS-09‡	21 May	21 Apr.	14 May	21 May	25 June	25 June	31 July	31 July
UCPRS1-10§	27 Apr.	25 Mar.	13 Apr.	27 Apr.	4 June	4 June	28 June	10 Nov.
UCPRS2-10¶	27 Apr.	25 Mar.	13 Apr.	27 Apr.	4 June	4 June	28 June	10 Nov.

† Upper Coastal Plain Research Station in 2009.

‡ Oxford Tobacco Research Station in 2009.

§ Upper Coastal Plain Research Station in 2010.

¶ Upper Coastal Plain Research Station in 2010.

Table 4. Composite soil analysis of plots not receiving supplemental potassium.

Test site and sampling interval	Sampling date	Depth cm	P mg dm ⁻³	K – cmol _c kg ⁻¹	Mg
UCPRS-09† 1 mo	30 Mar.	0–15	45.6	0.34	1.08
		15–30	67.9	0.29	0.81
UCPRS-09† 1 wk	22 Apr.	0–15	88.8	0.40	1.20
		15–30	42.7	0.22	0.98
UCPRS-09† planting	29 Apr.	0–15	84.3	0.30	0.85
		15–30	71.7	0.23	0.85
UCPRS-09† layby	18 June	0–15	94.0	0.31	0.93
		15–30	99.5	0.22	0.87
OTRS-09‡ 1 mo	21 Apr.	0–15	159.6	0.19	0.60
		15–30	121.5	0.17	0.69
OTRS-09‡ 1 wk	14 May	0–15	129.9	0.19	0.63
		15–30	114.3	0.17	0.61
OTRS-09‡ planting	21 May	0–15	155.8	0.17	0.54
		15–30	139.9	0.15	0.48
OTRS-09‡ layby	25 June	0–15	152.7	0.13	0.53
		15–30	149.1	0.15	0.51
UCPRS1-10§ 1 mo	25 Mar.	0–15	219.7	0.30	1.16
		15–30	192.4	0.26	0.64
UCPRS1-10§ 1 wk	13 Apr.	0–15	212.2	0.37	1.20
		15–30	197.1	0.28	0.72
UCPRS1-10§ planting	27 Apr.	0–15	200.1	0.44	1.51
		15–30	225.7	0.29	0.78
UCPRS1-10§ layby	4 June	0–15	186.2	0.28	1.06
		15–30	220.5	0.23	0.55
UCPRS2-10¶ 1 mo	25 Mar.	0–15	115.6	0.32	0.99
		15–30	50.0	0.28	0.69
UCPRS2-10¶ 1 wk	13 Apr.	0–15	122.8	0.30	0.97
		15–30	78.8	0.31	0.69
UCPRS2-10¶ planting	27 Apr.	0–15	151.3	0.41	1.27
		15–30	138.1	0.30	0.68
UCPRS2-10¶ layby	4 June	0–15	150.9	0.30	1.00
		15–30	93.4	0.29	0.61

† Upper Coastal Plain Research Station in 2009.

‡ Oxford Tobacco Research Station in 2009.

§ Upper Coastal Plain Research Station in 2010.

¶ Upper Coastal Plain Research Station in 2010.

incorporated into the soil using a field cultivator. Posttransplanting applications were made with a single band application which was 12 cm away from the plant and 12-cm deep. Band applications were placed into a furrow created with a 140 Farmall tractor (Case International Harvester, Racine, WI). Following K application treated furrows were closed with single row rolling cultivators.

Tissue samples were collected at three growth stages: layby (when plants were roughly 38 cm tall), topping (just after flower was removed), and after curing (weighted composite sample

of all four harvested stalk positions). Sampling intervals were chosen because of the critical times they represent during the growing season. Layby occurs 4 to 6 wk after transplanting and is the last opportunity tobacco producers have to make fertilizer applications. Topping occurs 8 to 10 wk after transplanting and is the time when K deficiencies are most often observed. Cured leaf samples were collected to determine final nutrient content. Soil samples were collected from two depths (15 and 30 cm) at four separate intervals: 1 mo before transplanting, 1 wk before transplanting, at planting, and at layby. Soil sampling dates for each interval are described in Table 4.

All K was supplied as sulfate of potash magnesia (0–0–22) (K-Mag, The Mosaic Company, Plymouth, MN). Sulfate of potash magnesia also supplied Mg (11%) and S (22%), and rates were chosen that ensured adequate amounts of both nutrients for plant use. Nitrogen was supplied in split applications from 30% urea ammonium nitrate (UAN) applied at a total rate of 78 kg N ha⁻¹ at the UCPRS and from calcium nitrate (15.5–0–0) applied at a total rate of 67 kg N ha⁻¹ at the OTRS.

Analytical Procedures

Total Alkaloids and Reducing Sugars

Total alkaloids and reducing sugars were determined by the North Carolina State University Tobacco Analytical Services Lab on campus. Fifty-gram cured leaf samples were prepared for each plot by compositing cured leaf from each stalk position on a weighted-mean basis. Oven-dried samples were ground to pass through a 1-mm sieve and analyzed for percent total alkaloids and percent reducing sugars using the method of Davis (1976).

Nitrogen, Phosphorus, Potassium, and Magnesium

Percent N in the leaf tissue was determined by dry combustion using the method of Plank (1992) using a PerkinElmer PE 2400 elemental analyzer (PerkinElmer Inc., Waltham, MA). Percent P, K, and Mg in the leaf tissue was determined using the method of Plank (1992) using a PerkinElmer Optima 2000 DV elemental analyzer (PerkinElmer Inc., Waltham, MA).

Soil Samples

Soil samples were analyzed at the North Carolina Department of Agriculture and Consumer Services Agronomic Lab in Raleigh, NC. Samples were analyzed for P, K, Ca, Mg, S, Na, Mn, Cu, and Zn by means of Mehlich-3 extractant using inductively coupled argon plasma spectroscopy on a volume basis (Mehlich, 1984a). Cation exchange capacity (CEC) was determined by summation of basic cations (excluding Na) and buffer acidity (Mehlich et al., 1976). Soil

Table 5. Probability > F values for yield, quality, value, total alkaloids, reducing sugars, and elemental leaf content.

Variable	Location × Rate × Application		Location × Rate			
	Application	Application	Application	Application	Rate	Application
Yield	0.8086	0.8236	0.0202*	0.9788	0.0361*	0.9900
Grade index	0.5649	0.5869	0.7780	0.6872	0.5591	0.5936
Crop value	0.9429	0.8971	0.2356	0.9067	0.4577	0.6332
Total alkaloids	0.4944	0.4184	0.0528	0.1010	0.9177	0.6730
Reducing sugars	0.3991	0.8189	0.2936	0.1002	0.0238*	0.4340
N-layby	0.8889	0.1214	0.4202	0.4851	0.1477	0.5720
N-topping	0.9713	0.9809	0.3896	0.3775	0.9573	0.0643
N-after curing	0.7740	0.5395	0.3187	0.5575	0.0964	0.9103
P-layby	0.5589	0.6626	0.2123	0.9247	0.1385	0.1125
P-topping	0.3110	0.1928	0.7479	0.0204*	0.5022	0.1264
P-topping-UCPRS-09	–	0.7555	–	–	0.4261	0.1200
P-topping-OTRS-09	–	0.1544	–	–	0.7869	0.0891
P-topping-UCPRS1-10	–	0.7926	–	–	0.4195	0.1995
P-topping-UCPRS2-10	–	0.8363	–	–	0.1546	0.0658
P-after curing	0.9443	0.5425	0.5239	0.5852	0.3129	0.0323*
K-layby	0.7917	0.1237	0.6114	0.0015*	0.0584	0.3047
K-topping	0.4049	0.5651	0.6321	0.0343*	0.4800	0.1848
K-after curing	0.4558	0.7495	0.0319*	0.3898	<0.0001*	0.1847
Mg-layby	0.3930	0.4213	0.2272	0.1936	0.0054*	0.0101*
Mg-topping	0.3574	0.4499	0.6098	0.1131	0.6051	0.2137
Mg-after curing	0.2069	0.5926	0.5374	0.2082	0.0113*	0.1435

* Significant at the 0.05 probability level.

pH was determined on a 1:1 soil/water volume ratio. Humic matter determinations were made using a NaOH digestion with colorimetric determination (Mehlich, 1984b).

Statistical Analysis

Treatments were replicated four times and arranged in a randomized complete block design. Data for crop yield, crop quality, crop value, percent total alkaloids, percent reducing sugars, and elemental leaf content were subjected to ANOVA using the PROC GLM procedure in SAS (SAS version 9.1, SAS Institute, Cary, NC). Treatment means were separated using Fisher's Protected LSD test at $p \leq 0.05$.

RESULTS

A significant location effect was noted across all parameters because of varying soil types and growing conditions; however, the interaction of location × rate × application method was not significant and data are pooled where appropriate (Table 5).

Physical Characteristics, Chemical Characteristics, and Crop Value

Yield

The response of leaf yield to increasing rates of K_2O was different at one location (Table 5). The only location that demonstrated a yield response was the 2009 OTRS site, a trend is difficult to establish based on application rate alone, but a response was observed as rates were increased to 252 kg K_2O ha⁻¹ at this site (Table 6). At all other locations, 0 kg K_2O ha⁻¹ was adequate. The lack of response to additional units of K_2O is possibly due to soils having medium to high residual K (Table 1), which is common on >60% of tobacco producing soils in the state (Smith, 2011), as well as adequate soil moisture throughout the growing season (Table 2).

Table 6. Yield response to increasing rates of K_2O across all treatments at individual locations.†

K_2O Rate	UCPRS-09‡	OTRS-09§	UCPRS1-10¶	UCPRS2-10#
	kg ha ⁻¹			
0	3364a	3428ab	3374a	3260a
84	3331a	3252b	3411a	3405a
140	3239a	3385ab	3387a	3404a
196	3183a	3234b	3514a	3206a
252	3126a	3644a	3627a	3422a

† Means followed by the same letter within the same category are not significantly different.

‡ Upper Coastal Plain Research Station in 2009.

§ Oxford Tobacco Research Station in 2009.

¶ Upper Coastal Plain Research Station in 2010.

Upper Coastal Plain Research Station in 2010.

Grade Index

The quality of cured tobacco was not significantly affected by increasing application rates of K_2O from 0 to 252 kg K_2O ha⁻¹ (Tables 5 and 7). Results are similar to those from Chaplin and Miner (1980) and Collins and Hawks (1993) that did not consistently find a correlation between K_2O rate and leaf quality when rates in excess of those needed for maximum yield were used.

Crop Value

The value of cured tobacco was not affected by increasing rates of K_2O (Tables 5 and 7). Results are similar to Collins and Hawks (1993) who could not establish a correlation between increased rates of K_2O and leaf quality, which would increase value.

Total Alkaloids

Total alkaloid accumulation in cured leaves was not affected by K_2O rate (Tables 5 and 7). Results are similar to those by Woltz et

Table 7. Physical and chemical response to increasing rates of K₂O. Data are pooled over all locations.†

K ₂ O rate kg ha ⁻¹	Grade index‡	Value \$ ha ⁻¹	Total alkaloids§	Reducing sugars§
			—	—
0	80a	11287a	4.06a	12.39ab
84	79a	10502a	4.04a	12.65a
140	79a	10500a	4.07a	11.97ab
196	79a	10588a	4.08a	11.78b
252	78a	10914a	4.06a	11.84b

† Means followed by the same letter within the same category are not significantly different.

‡ Grade index is a measure of tobacco quality on a ranking scale of 1 to 100, with 100 having the highest quality.

§ Total alkaloid and reducing sugar data were collected from a weighted composite sample of all four harvested stalk positions.

al. (1949), Elliot (1968), and Chaplin and Miner (1980) who could not establish a correlation between increased rates of K₂O and total alkaloid content. Total alkaloids are typically only influenced by K₂O rate when a corresponding yield affect is observed (Leggett et al., 1977; Chaplin and Miner, 1980).

Reducing Sugars

Reducing sugar content was affected by increasing rates of K₂O (Table 5). As rates of K₂O increased, the reducing sugar content of cured tobacco slightly decreased (Table 7); however, reducing sugar content is within the established range for flue-cured tobacco. These findings are in conflict with Woltz et al. (1949), Elliot (1968), Chaplin and Miner (1980), and Tso (1990a) who could not establish a correlation between increased rates of K₂O and reducing sugar content.

Elemental Leaf Content

Elemental leaf content for N, P, K, and Mg was measured at three separate intervals during the growing season: at layby, at topping, and after curing.

Nitrogen

Neither application method or application rate of K₂O had an effect on total N content in leaf tissue at any sampling

interval (Table 5). This is in conflict with Tso (1990a) who reported that as rates of K₂O increase, total N content is decreased. Overall, additional K did not decrease the uptake of N from the soil which would have decreased overall N content in plant tissue.

Phosphorus

Application method of K₂O had a significant effect on P content in leaf tissue at topping when all locations are pooled (Table 5); however, once each test site was analyzed individually effects are not significant (Table 5). Application method of K₂O also had a significant effect on P content in leaf tissue after curing (Table 5). The range of P content after curing was 0.21 to 0.24% (data not shown), which is not agronomically significant because P content at each interval is above established deficiency level of 0.12% for leaf tissue (Campbell, 2009) and therefore was not a limiting factor.

Potassium

An application method response was observed at all sampling intervals at UCPRS2-10 (Tables 5 and 8). Alternatively, a rate response in K leaf content was observed after curing at three locations (Tables 5 and 9). Establishing a trend for application methods is difficult where a response was observed; however, K content tends to increase when the nutrient is applied closer to transplanting (Table 8). Potassium rate had a significant effect on elemental leaf content after curing at UCPRS-09, OTRS-09, and UCPRS1-10 (Table 9). Elemental K content tends to increase as rates of K₂O increase to 196 kg K₂O ha⁻¹ (Table 9). Overall, K content was sufficient at all rates and application methods to maximize yield (Tables 8 and 9), and elemental leaf content at each interval was above the established deficiency level of 1.00% (Flower, 1999). No visual K deficiency symptoms were observed in this study, thus furthering the justification for lower rates of applied K. The results are in agreement with Chaplin and Miner (1980) who determined that K content in cured tobacco leaves increased as rates of K₂O increased.

Table 8. Leaf K content by application method at layby, topping, and after curing at individual locations.†

Application method	UCPRS-09‡			OTRS-09§			UCPRS1-10¶			UCPRS2-10#		
	Layby	Topping	After curing	Layby	Topping	After curing	Layby	Topping	After curing	Layby	Topping	After curing
None applied††	4.32a	2.17a	2.42a	3.11a	1.87a	1.48a	3.62a	2.51a	2.23a	3.86d	2.41c	2.15c
BC-1M‡‡	4.69a	2.41a	2.52a	3.65a	3.90a	2.35a	4.02a	2.67a	2.33a	4.08cd	2.77b	2.25bc
BC-1W§§	4.72a	2.60a	2.60a	3.79a	2.97a	2.30a	4.08a	2.54a	2.25a	4.15bc	2.87b	2.43ab
AP¶¶	4.65a	2.60a	2.52a	3.59a	3.25a	2.32a	4.14a	2.99a	2.48a	4.38ab	3.28a	2.47ab
Split###	4.67a	2.59a	2.66a	3.53a	3.15a	2.29a	4.03a	2.67a	2.41a	4.45a	3.22a	2.53a

† Means followed by the same letter within the same category are not significantly different.

‡ Upper Coastal Plain Research Station in 2009.

§ Oxford Tobacco Research Station 2009.

¶ Upper Coastal Plain Research Station 2010.

Upper Coastal Plain Research Station 2010.

†† None applied = control where 0 kg K₂O ha⁻¹ was applied.

‡‡ BC-1M = all potassium broadcast applied 1 mo before transplanting.

§§ BC-1W = all potassium broadcast applied 1 wk before transplanting.

¶¶ AP = all potassium band applied at transplanting.

Split = all potassium band applied at one-half rate at transplanting and one-half rate 4 to 6 wk later.

Table 9. Leaf K content response to increasing rates of K₂O after curing at individual locations.†

K ₂ O rate kg ha ⁻¹	UCPRS- 09‡	OTRS- 09§	UCPRS1- 10¶	UCPRS2- 10#
0	2.41b	1.48c	2.22b	2.15a
84	2.37b	2.13b	2.13b	2.40a
140	2.53ab	2.36ab	2.32ab	2.37a
196	2.80a	2.29ab	2.63a	2.43a
252	2.59ab	2.48a	2.39ab	2.47a

† Means followed by the same letter within the same category are not significantly different.

‡ Upper Coastal Plain Research Station in 2009.

§ Oxford Tobacco Research Station in 2009.

¶ Upper Coastal Plain Research Station in 2010.

Upper Coastal Plain Research Station in 2010.

Magnesium

At layby, both application rate of potassium magnesium sulfate and application method had a significant effect on elemental Mg leaf content (Table 5). As rates of applied potassium magnesium sulfate increased Mg leaf content increased; however, the range of Mg content was 0.52 to 0.57% (data not shown) which does not have an agronomic impact on the crop. Potassium magnesium sulfate applied at planting contained the highest amount of Mg in leaf tissue with 0.57% (data not shown); all other intervals contained 0.52 to 0.55% (data not shown). After curing, leaf Mg content was only affected by application rate of potassium magnesium sulfate (Table 5). Magnesium content in the cured leaf was not significantly improved above 140 kg K₂O ha⁻¹. The range of Mg content after curing was 0.60 to 0.63% (data not shown), which is not agronomically significant because the nutrient was not limiting and leaf tissue from plots with the lowest Mg content were above the established deficiency level of 0.15% (Tso, 1990b).

Soil Analysis

Soil Analysis

Soil samples were only taken from plots receiving 0 kg K₂O ha⁻¹ and sampling occurred each time K applications were made to other plots. Two samples were taken, one from 0 to 15 cm and another from 15 to 30 cm. Soil samples were not statistically analyzed because they were not taken across all treatments; however, it is worth noting the data for each test site. Variability in soil fertility was observed across all locations (Table 4). Although there are no specified critical nutrient levels for tobacco, it is possible that Piedmont soils with <30 g m⁻³ of P and Coastal Plain soils with <50 g m⁻³ of P may exhibit deficiency symptoms without supplemental fertilization (Hardy et al., 2012). Soils with <0.50 cmol_c kg⁻¹ of K and 0.25 cmol_c kg⁻¹ of Mg may also exhibit deficiencies when no supplemental fertilizer is applied (Hardy et al., 2012). Soil samples taken from control plots during research were used to make generalizations about individual test site characteristics.

For all test sites, soil P and Mg levels were sufficient for optimum tobacco growth for the entire season (Table 4). Before transplanting at UCPRS-09 soil P levels were below the critical level; however, at the time of transplanting soil P was adequate (Table 4). Visual P and Mg deficiency symptoms were not observed at any test site. Soil K levels were below the previously mentioned limit of 0.50 cmol_c kg⁻¹ at all test sites (Table 4); however, visual

deficiency symptoms were not observed. Soil fertility was adequate all season for the respective nutrients.

DISCUSSION

Application method and application timing had no significant agronomic effect on yield, quality, value, or any measured leaf constituents. In both years, crop quality was not affected by application rate and application method, thus demonstrating that under conditions present at test sites alternative application methods are acceptable. Leaf yield demonstrated a favorable response to 252 kg K₂O ha⁻¹ at OTRS-09; however, at all other locations 0 kg K₂O ha⁻¹ was enough to produce favorable yields with all other measured parameters remaining unaffected. The 0 kg K₂O ha⁻¹ rate is adequate, but according to the state soil testing lab a minimum rate of 84 kg K₂O ha⁻¹ would be recommended to overcome the possibility of unfavorable growing conditions as well as to prevent the mining of soil nutrients. Under the environmental conditions of these experiments, 84 kg K₂O ha⁻¹ applied broadcast 1 mo before planting, broadcast at planting, banded at planting, or applied in split applications provided adequate amounts of K to ensure sufficient yield and quality at all locations. It is likely that early broadcast applications of K₂O with current rate recommendations would only be of concern with combinations of conditions that included coarse soil textures, low residual soil K, and/or excessive leaching rainfall.

Soil texture and depth to clay have a major impact on K application rates, and both must be considered. Test sites for this study were selected based on soil type and texture, and these characteristics had a significant effect on crop response to K application. The loss of K⁺ is also of concern on coarse-textured soils similar to the Norfolk soil series at UCPRS-1. Coarse textured soils have lower cation exchange capacities and monovalent cations are often lost from the rooting zone in the event of leaching rainfall. However, if the depth to clay is <25 cm it is likely that K⁺ lost to leaching is held in the clay subsoil where it can be used once adequate root growth has occurred. It is advisable for producers to take soil samples down to 25 cm to provide a reasonable estimate of the nutrient content in the soil. Subsoil K that remains unaccounted for is the reason why responses to K application are not observed, even when a soil test indicates it should. Additionally, coarse-textured soils have difficulty maintaining adequate soil moisture during periods of drought or extended dry conditions. Soil moisture facilitates diffusion of the K⁺ ion from the point of application to the root zone, and in the absence of adequate moisture diffusion is greatly inhibited.

The considerations of soil texture and depth to clay are extremely important when making K recommendations and fit very well with reports from Denton et al. (1987). Denton et al. (1987) report findings similar to those from this study for flue-cured tobacco produced across a variety of soil textures. Optimum rates of K₂O were found to differ significantly among Fertility Capability Classification (FCC) soil groups; however, the highest optimum rates were on soils with a sand or loamy sand surface where the A and E horizons were >0.5 m thick (Denton et al., 1987). Alternatively, the lowest optimum K rates reported by Denton et al. (1987) were on soils with thin, loamy surfaces, which indicate a relatively shallow depth to clay. As previously mentioned, K⁺ can leach from the upper soil profile when cation exchange capacity is low, such as with a sandy topsoil; however, if

the depth to clay is <25 cm it is likely that K⁺ lost to leaching is held in the clay subsoil where it can be used once adequate root growth has occurred. The subsoil K is more accessible to tobacco plants on soils with thin, loamy surfaces than on soils with thick, sandy surfaces due to the increased depth to the B horizon (Denton et al., 1987). Ultimately, responses to increased rates of K₂O would be expected under these conditions as a result of compensation for the loss of K⁺ due to leaching.

Additionally, the independent application of K works extremely well for producers using alternative fertility programs. As mentioned previously, additional application of P is not necessary on 85% of the soils used for tobacco production in the state (Smith, 2011), and as a result producers can decouple complete N–P–K fertilizers and apply N and K independently of one another. This has allowed producers to explore alternate sources for both nutrients. Sulfate of potash magnesia fertilizer (0–0–22) has gained popularity as a result of it being a cheaper source of K than when supplied in a complete fertilizer and because of the additional Mg and S it supplies. Also, as a part of alternative fertility programs, liquid N (28% UAN, 30% UAN, and 32% UAN) sources are now being implemented as the only source of N for the entire growing season by many producers. Liquid N sources are often cheaper than other N sources used in tobacco production, and are easier to apply for producers with experience applying them to other crops.

CONCLUSION

Applying K₂O independent of other nutrients, as done in this study, fits extremely well with alternative fertilizer plans that flue-cured tobacco producers are now implementing. Furthermore, lower K recommendations and alternative fertilizer management plans are suitable for producers when both residual soil K as well as soil texture are considered. Early broadcast applications of K₂O with current rate recommendations would only be of concern with combinations of conditions that included coarse soil textures, low K indices, and/or excessive leaching rainfall. There is great potential for producers on fine- or medium-textured soils to reduce K₂O rates and apply K₂O through different methods without limiting yield and quality, thus reducing workloads and creating larger returns.

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