

ABSTRACT

LI, QI. Characterization of Atmospheric Reactive Nitrogen Emissions from Global Agricultural Sources. (Under the direction of Dr. Viney P. Aneja).

Reactive nitrogen (Nr) includes inorganic chemically reduced nitrogen compounds such as ammonia (NH₃) and ionic ammonium (NH₄⁺); inorganic chemically oxidized nitrogen compounds such as nitrogen oxides (NO_x), nitric acid (HNO₃), nitrate ion (NO₃⁻), and nitrous oxide (NO₂); and organic nitrogen compounds such as urea, amino acids, and proteins. They are biologically active, chemically reactive, and radiatively active in the Earth's atmosphere and biosphere, which are different from non-reactive nitrogen gas.

Here, three statistical model (N₂O_STAT, NH₃_STAT, and NO_STAT) are developed for characterizing atmospheric Nr emissions from agricultural sources. We obtained Nr emissions and physicochemical variables (i.e., air temperature, soil temperature, soil moisture, soil pH, and Ni input to the soil) from published journal articles. The statistical model was developed by expressing a multiple linear regression equation between Nr emission and the physicochemical variables. The model was evaluated for 2012 Nr emissions. Results of the model are compared with other global and regional models (e.g. EDGAR, etc.). The results indicate that, in comparison to other data sets, the model generates a lower global N₂O estimate by 9-20% (N₂O_STAT: 3.75 Tg N yr⁻¹; EDGAR: 4.49 Tg N yr⁻¹; FAO: 4.07 Tg N yr⁻¹), a lower global NH₃ estimates by 57%, (NH₃_STAT: 14.2 Tg N yr⁻¹; EDGAR: 33.0 Tg N yr⁻¹), and a lower global NO_x estimates by 87%, (NO_STAT: 0.2 Tg N yr⁻¹; EDGAR: 1.6 Tg N yr⁻¹). We also performed a region-based analysis (U.S., India, and China) using the N₂O_STAT and NH₃_STAT models. Based on results, the difference in the global estimates is attributed to the lower estimates in major agricultural countries like China and India. Statistical models can capture the spatial distribution of global Nr emissions by utilizing a more simplified approach

than those used by other readily available data sets. Moreover, three models provide an opportunity of predict future Nr emissions in a changing world.

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Characterization of Atmospheric Reactive Nitrogen Emissions from Global Agricultural Sources

by
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DEDICATION

For my Mom and Dad.

For my friends in U.S. and in China.

For Super Junior.

BIOGRAPHY

Qi Li was born in Anyang, Henan Province, China. She finished her elementary, middle, and high schools in her hometown and was admitted to Ocean University of China in 2011. As freshmen, she joined Environmental Protection Association in college and showed strong interests in environment issues, especially in air quality problems.

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CHAPTER 1 Introductions

Reactive nitrogen includes inorganic chemically reduced nitrogen compounds such as ammonia (NH_3) and ionic ammonium (NH_4^+); inorganic chemically oxidized nitrogen compounds such as nitrogen oxides (NO_x), nitric acid (HNO_3), nitrate ion (NO_3^-), and nitrous oxide (NO_2); and organic nitrogen compounds such as urea, amino acids, and proteins. They are biologically active, chemically reactive, and radiatively active in the Earth's atmosphere and biosphere, which are different from non-reactive nitrogen gas (N_2).

1.1. N_2O

The largest human influence on climate change since the end of the 20th century is the emission of greenhouse gases to the atmosphere. In addition to carbon dioxide (CO_2), nitrous oxide (N_2O) is one of the most important greenhouse gases, which has a global warming potential (GWP) of ~300 for a 100-year timescale (Myhre et al., 2013). N_2O also contributes to the destruction of ozone in the stratosphere (Ravinshankara et al., 2009). The lifetime of N_2O in the atmosphere is ~114 years, indicating its long-term influence on a planetary scale (Forster et al 2007).

Since 1978, globally-averaged N_2O levels in the atmosphere have been collected at Moana Loa, Hawaii. Prior to 1978, N_2O levels have been extracted from ice core data from Antarctica. N_2O concentrations in the atmosphere were relatively stable until the start of the Industrial Revolution. Since that time, N_2O concentration has increased by 18%, from ~270 ppb to the current value of ~331 ppb (Figure 1.1). This increase is primarily due to intensive human activities, particularly related to agriculture.

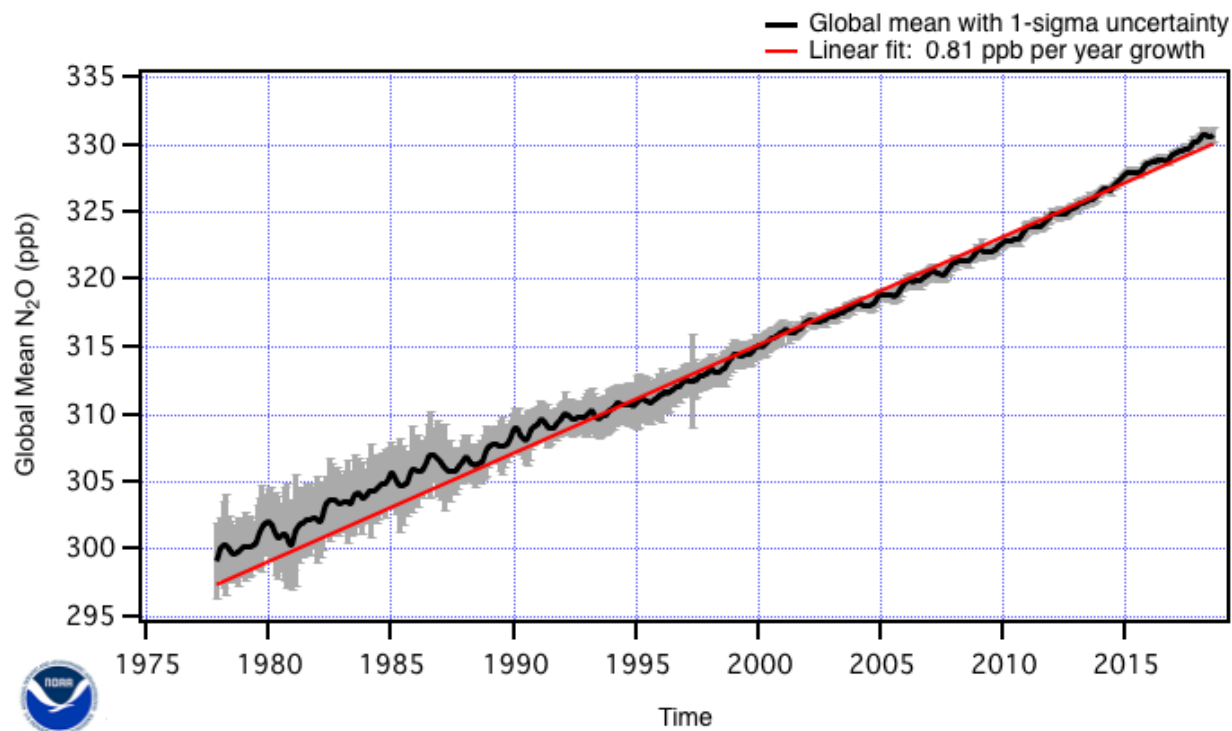


Figure 1.1 Atmospheric N₂O concentration trend from the combined Global Monitoring Division data set for the period 1977-2018 (NOAA, 2018).

Ciais et al. (2013) have estimated the yearly emission of N₂O to be 17.9 Tg N yr⁻¹ for the period 2006-2011. N₂O emissions from natural sources are 11.0 Tg N yr⁻¹, which includes sources from land and ocean. Agricultural activities are the primary anthropogenic source of N₂O, contributing to 5.6 Tg N yr⁻¹ (Schlesinger and Bernhardt, 2013). With increasing food, fiber and energy production, along with climate change and agricultural intensification, N₂O emissions from soils have increased since 1990, from 3.5 Tg N yr⁻¹ to 4.1 Tg N yr⁻¹ (IPCC, 2006).

During the period 1961-2010, global N₂O emissions from manure usage have grown from 1.17 to 2.03 Tg N yr⁻¹ at an average rate of 1.10% yr⁻¹ (Smith et al., 2014). Meanwhile during the same period, N₂O emissions from synthetic fertilizers have increased 0.14 to 1.40 Tg N yr⁻¹ at an average rate of 3.90% yr⁻¹ globally (Tubiello et al., 2013). This indicates that synthetic fertilizers will have a greater contribution to N₂O emission than manure within less

than 10 years (Smith et al., 2014). Besides agriculture, other sources such as fossil fuel combustion, industrial processes, and biomass burning also contribute to the total N₂O emissions, but these are all relatively small sources. Intensive fertilizer usage has increased the emissions of reactive nitrogen compounds, including N₂O from soils through nitrification and denitrification processes (Figure 1.2). Denitrification is a microbially facilitated process that leads to significant N losses from agricultural systems. During denitrification, nitrate and nitrite are reduced, and a portion converted into N₂O (Butterbach-Bahl et al., 2013). Nitrification is the aerobic microbial oxidation of ammonium or ammonia to nitrite followed by oxidation of nitrite to nitrate. It is an important process in the nitrogen cycle in which N₂O is generated as a by-product that leaks from microbial cells into soil and eventually into atmosphere (Schlesinger and Bernhardt, 2013).

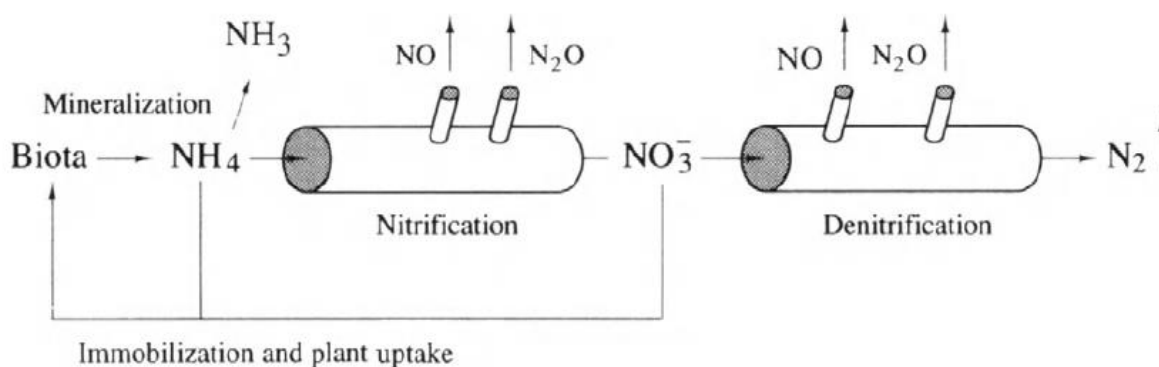


Figure 1.2 N₂O production through nitrification and denitrification in soil (Firestone and Davidson, 1989, as modified by Schlesinger and Bernhardt, 2013).

The ratio of N₂O/NO₃⁻ (nitrification) and N₂O/N₂ (denitrification) products are important parameters determining the change of global budget, and they are affected by various physical, chemical and biological factors and their interacting effects (Granli and Bockman, 1994). To develop a predictive model for N₂O emission under different agroecosystems, climate variables, soil characteristics, and cropping practices must be considered. Granli and Bockman (1994) provided a review on factors that control N₂O emissions from soil, including moisture

and aeration, temperature, soluble and decomposable carbon, soil and fertilizer nitrogen, soil pH and salinity. Table 1.1 summarizes the variables that affect product ratios during nitrification and denitrification in agricultural soils. In nitrification, N₂O emission will increase with decreasing O₂ concentration, increasing water above field capacity, low NH₄⁺ concentrations, increasing or decreasing pH, and increasing temperature. The production of N₂O by denitrification is enhanced in conditions characterized by high nitrate, low C availability, low pH, low temperature, low N₂O reductase activity, and 60 to 90% water filled pore space (WFPS) (Stehfest and Bouwman, 2006; Rashti et al., 2015; Wang et al., 2017).

Table 1.1 Variables that affect product ratios during nitrification and denitrification in agricultural soils (Granli and Bockman, 1994; Firestone and Davidson, 1989).

Process	Variable	Effect
Nitrification	[O ₂]	<i>Will increase N₂O/NO₃⁻ ratio</i> Decreasing O ₂ concentration
	[H ₂ O]	Increasing H ₂ O above field capacity
	[NH ₄ ⁺]	Low NH ₄ ⁺ concentrations
	pH	Increasing or decreasing pH
	Temperature	Increasing temperature
Denitrification	[NO ₃ ⁻] or [NO ₂ ⁻]	<i>Will increase N₂O/N₂ ratio</i> Increasing oxidant
	[O ₂]	Increasing O ₂
	Carbon	Decreasing C availability
	pH	Decreasing pH
	Temperature	Decreasing temperature
	Enzyme status	Low N ₂ O reductase activity
	[H ₂ O]	Decreasing between 60 and 90% WFPS

Uncertainty of estimates of sources and sinks still exist in global N₂O budget, especially from agricultural soils (Oertel et al., 2016). Inadequate understanding of the real mechanism in N₂O emission from agricultural soils may be responsible for limited attention to agriculture as a

major contributor of the increasing atmospheric N₂O. With increasing food and energy production, along with climate change and agricultural intensification, N₂O emissions from soils and groundwater will continue to increase, suggesting the necessity of an updated global budget for N₂O that accounts for these recent changes.

1.2. NH₃

Among all the reactive nitrogen species, NH₃ is an important component of the global nitrogen cycle (Fowler et al., 2013, 2015), as well as a large anthropogenic source of reactive nitrogen. Its emission is primarily from agricultural sources (Schlesinger and Hartley 1992, Aneja et al., 2009). A large loss of reactive nitrogen from loss as ammonia (NH₃), mainly from the use of animal manure and synthetic fertilizer, is a major problem in agricultural systems (Aneja et al., 2008; Bouwman et al., 2011). Nitrogen loss into the environment represents a big economic cost in lost nutrients. If the majority of the NH₃ emissions can be prevented/conserved and recovered, it could save a significant amount of money (assuming approximately 33% nitrogen use efficiency by crops, the savings annually maybe about \$15.9 billion (Raun and Johnson, 1999)). This problem has produced impacts not only in agricultural systems, but also in adjacent, and downwind ecosystems. NH₃ reacts with other atmospheric compounds and serves as a precursor of fine particulate matter (PM_{2.5}), which is listed as one of the six crucial atmospheric pollutant by U.S. Environmental Protection Agency (U.S. EPA). Exposure to high PM_{2.5} concentration can lead to adverse health effects and premature death (Kwok et al., 2013). It can also reduce visibility and lead to regional haze (Zhuang et al., 2014). In addition, NH₃ is deposited in terrestrial, aquatic and marine ecosystems, where it causes ecosystem degradation (Bobbink et al., 2010). Through indirect microbial transformations, NH₃ emission also contributes to the elevated levels of N₂O emission by increasing the rate of nitrogen cycling in the natural environment (Davidson, 2009). N₂O is one of the most important greenhouse gases, which contributes to the global warming.

Since last century, the use of synthetic nitrogen fertilizers and cultivation of nitrogen-fixing crops has expanded significantly (Battye et al, 2017). FAO predicts that the fertilizer

usage in south Asia will more than double between 2006 and 2060. Due to these increasing agricultural activities, nitrogen compounds emitted to the atmosphere have also increased dramatically, including significant increases in NH_3 concentrations, which are seen globally especially in agricultural regions. From 2002 to 2016, there have been substantial increases in atmospheric NH_3 emissions over several of the world's major agricultural regions, e.g., US (2.61% yr⁻¹), the European Union (EU) (1.83% yr⁻¹), and China (2.27% yr⁻¹) (Warner et al., 2017).

NH_3 emitted from agricultural activities has the largest contribution to the atmospheric budget of reactive N (Reis et al., 2009). There are other sources of NH_3 , including biomass burning, industrial processes, vehicular emissions, and volatilization from soils and oceans. Recent studies indicate that there has been an increase in total NH_3 emission over the past few decades. The estimated global NH_3 emission for 1990 is about 54 Tg N yr⁻¹ (Bouwman et al. 1997).. However, an assessment by Schlesinger and Hartley (1992) indicates the global annual sources of NH_3 in the atmosphere to be about 75 Tg of N as NH_3 . It also shows that the anthropogenic sources contribute about 80 % of global NH_3 . Therefore, a comprehensive review of emissions of NH_3 from various anthropogenic sources is needed, so that effective control strategies can be formulated to reduce such emissions (Behera et al, 2013).

NH_3 may be used in fertilizers either as its salts, in solution or anhydrously. Ammonium fertilizers play a crucial role in high-yield crop production and contribute to large NH_3 emission (Warner et al, 2017). Around 2% of the world energy is used to produce the annual 120Mt N_r for fertilizer and other uses, mainly as ammonia through the Haber-Bosch process (Abrol et al., 2017). The Haber-Bosch process combines atmospheric nitrogen with hydrogen under high pressures and temperatures to produce NH_3 . This process, on one hand, feeds more population

with more food; but enhanced N-fixation due to human activities also impacts the nitrogen cycle. Nitrification in agricultural soils has the effect of reducing the release of NH_3 to the atmosphere. Some of the NO_3 produced is later denitrified. However, the contribution and increases of reactive nitrogen by the Haber-Bosch process is not fully offset by denitrification. From the global perspective (Battye et al., 2017), the global denitrification flux is estimated to be 600 Tg N y^{-1} , which would be sufficient to balance the rate of nitrogen fixation, but there are imbalances on local and regional scales that lead to adverse environmental problems.

Accurate estimates of NH_3 emission are necessary for global inventories. Global estimates are also important for developing better models to assess the impact of NH_3 emission on the atmosphere and the deposition of ammonia in terrestrial ecosystems. However, the NH_3 emissions estimates used in current modeling efforts are subject to considerable uncertainties (Battye et al, 2003).

There are many factors that affect NH_3 emission from soil. For example, ammonia emission increases with increasing nitrogen content in fertilizer and pH in soils. It also increases exponentially with temperature (Riddick et al., 2016). NH_3 emission also requires a minimum level of soil moisture for microbial activities (Warner et al., 2017).

1.3. NO_x

Nitrogen oxides (NO_x = NO + NO₂) are important trace constituents in the troposphere that regulate the consumption and production of photochemical oxidants, ozone (O₃) and hydroxy radicals (Warneck 1988). Although these gases do not directly affect the radiative balance, the formation of ozone and aerosols in the troposphere can result in negative consequences on air quality, radiative balance, dynamics, and chemistry of the atmosphere. Most of air pollution problems are caused by NO_x through chemical reactions, e.g. smog, acid rain. NO₂, as the most prevalent form of NO_x in the atmosphere, has been listed in criteria air pollutants by U.S. Environmental Protection Agency (U.S. EPA). In the National Ambient Air Quality Standards (NAAQS), the primary and secondary standard for NO₂ is 0.053 parts per million (ppm).

Tropospheric ozone pollution is one of the most significant air pollution problems in the United States, and most developing countries. It is harmful to both human health and vegetable and crop growth. NO_x is one of the most important precursors of tropospheric ozone formation. NO₂ reacts in the presence of air and sunlight to produce NO. When NO concentration is below 3-8 ppt, NO reacts with O₃ to produce NO₂ and O₂, but when NO concentration is higher, NO catalyzes the oxidation of CH₄, CO and volatile organic compounds (VOC) to produce O₃ (Warneck 1988), and NO gets recycled to NO₂ by free radicals. In rural environments, the reaction of NO with biogenic VOC can be a predominant source of ozone. NO_x in the troposphere can be captured by moisture in the form of nitric acid (HNO₃) to form acid rain, which directly accelerates acidification and eutrophication processes in the regional ecosystems. Acid rain is extremely harmful to not only the ecosystem, but also some segments of our economy. Based on these facts, NO_x emission needs to be reduced and well regulated.

NO_x can come from both natural sources, such as lightning, soil microbial activities and biomass burning, as well as anthropogenic sources such as fossil fuel combustion. Fossil fuel combustion contributes more than half of the global NO₂ budget. For NO, there are great portion of sources attributable to NO emissions from soils. The estimate of NO emission from soils can be up to 40% (Hutchinson, 1995). Published field measurements and inventories show substantial NO emissions from tropical savannas (Poth et al 1995), successional pastures (Keller and Reiners, 1994), and intensively managed agriculture (Valente and Thornton, 1993).

Many factors could control the NO emission from soil, e.g. soil temperature, soil pH, soil moisture, as well as available N into the soil. Many researchers have designed experiments to test how these factors affect the rate of NO emission rate from soil. Pilegaard (2013) points out that the most important factor that affects NO emission is the availability of nitrogen in the soil. It can come from nitrogen fixation, nitrogen deposition, or input as fertilizer/manure. In agricultural soil, NO emission is most affected by the available nitrogen from fertilizer/manure. Soil water amount controls the rate of O₂ supply, which directly affects the nitrification and denitrification. Goldberg & Gebauer (2009) observed that NO emission decreased after precipitation, but increased during the drought. In addition, due to the positive effect of soil temperature on microbial processes, NO emission generally increases with soil temperature. Schindlbacher et al. (2003) indicated that there is a positive exponential relationship between NO emission and soil temperature. Although there is no direct relationship between the rate of NO emission and soil pH, nitrification process intensified with high soil pH.

With recent advanced technology, there are an increasing number of field studies and modeling w models that predict NO_x emissions. Therefore, understanding the comprehensive mechanism of NO emission from soil and the knowledge of controlling factors for NO emission

have been improved. However, uncertainties for present global and regional estimates for NO emission still exist, and the estimates show a great range between the lowest and highest estimates. Also, most measurements are short-term, which do not represent the spatial and temporal variation of NO emission. This indicates a necessity to use the existing field measurement results and statistical models, with few input parameters, to estimate NO emission from agricultural soil.

1.4. Objectives

The goal of this project is to develop a statistical model to predict Nr emissions from agricultural soils containing synthetic and organic fertilizers using physicochemical properties of ecosystems from different regions. We also analyze the spatial distribution of Nr emissions from agricultural soils, and compare the results with other modeled emission inventories (i.e. EDGAR v.4.3.2, EPA, and FAOSTAT). This project offers global and regional scale estimates for the sources of N₂O, NH₃, and NO_x from fertilizer and manure application on land used for agricultural activities for the base year 2012. Statistical models provide an innovative way to estimate Nr emissions as well as an opportunity of predict future Nr emissions in a changing world.

CHAPTER 2 Methodology

2.1. Flowchart

Figure 2.1 is a flowchart that demonstrates the methodology we have adopted to estimate NH₃ global emissions from agricultural soil. Based on an initial literature review, we found four variables of importance in controlling NH₃ emissions from soil: (1) soil moisture, (2) soil and air temperature, (3) synthetic and organic fertilizer usage, and (4) soil pH. We performed a series of statistical analyses utilizing the R studio statistical software (<https://github.com/rstudio/rstudio>) to examine the distribution of data and to fit an appropriate regression model with Nr as the response (dependent) variable and other variables as independent predictors. Lastly, we used Integrated Land and Water Information System (ILWIS) v.3.31 Academic (<https://www.itc.nl/ilwis/download/ilwis33/>), a GIS tool to prepare the global data sets and apply the statistical model for predicting NH₃ emissions.

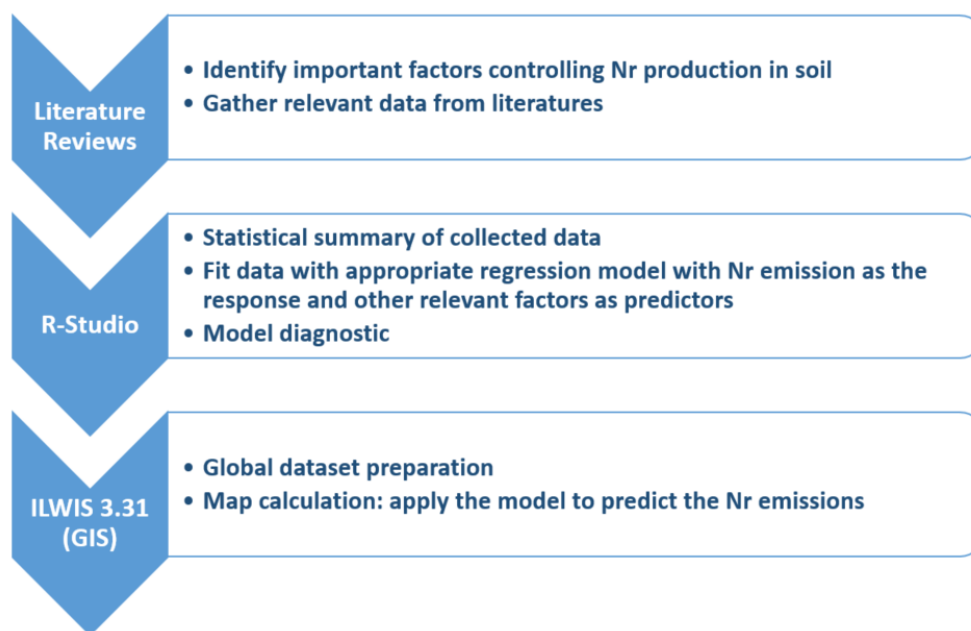


Figure 2.1 Flowchart of the methodology.

2.2. Data Collection

Data collection includes two parts, (1) data sets for developing the statistical model (Table 2.1a), and (2) global data sets for extrapolating the results to larger areas (Table 2.1b). For the statistical model, we conducted a literature review regarding agricultural soil N₂O and NH₃ emission (published after 2000), and NO emissions (published after 1990).

Table 2.1a. Data for statistical model development.

Parameter	Unit
N _r emissions	kg N ha ⁻¹ year ⁻¹
Air temperature	°C
Soil temperature	°C
Soil pH	-
Soil moisture	%
Fertilizer N content usage	kg N ha ⁻¹ year ⁻¹
Manure N content usage	kg N ha ⁻¹ year ⁻¹

Table 2.1b. Global data set used for prediction.

Parameter	Data sets
Soil temperature; Soil pH	Harmonized World Soil Database v 1.2 http://www.fao.org/soils-portal/soil-survey/soil-maps-and-databases/harmonized-world-soil-database-v12/en/
Soil moisture	The Global Soil Dataset for Earth System Modeling http://globalchange.bnu.edu.cn/research/soilw
Air temperature	ERA-Interim Global Atmospheric Reanalysis https://www.ecmwf.int/en/forecasts/datasets/archive-datasets/reanalysis-datasets/era-interim
Cropland cover, fertilizer usage, manure usage	Adjusted EarthStat's Cropland and Pasture Area http://www.earthstat.org/

2.3. Model Setup

2.3.1. N₂O

Based on the statistical analysis, we found that the data distribution, shown as the histogram of N₂O emissions (Figure 2.2a), was skewed to the right. To normalize the data, we transformed the data to its logarithmic value (Figure 2.2b) and determined that the data were then close to a normal distribution.

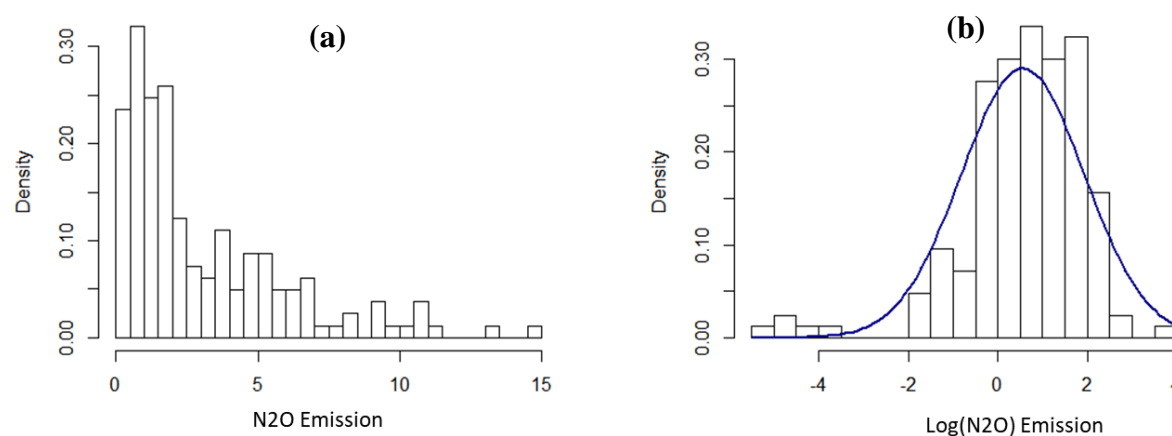


Figure 2.2. (a) Histogram of N₂O emission; and (b) log of N₂O emission.

Then, we used a multiple linear regression model to fit the response variable (the log of N₂O emissions), using the physicochemical variables as predictors. The statistically derived model (hereinafter: N₂O_STAT) to predict N₂O emissions from agricultural soils is mathematically expressed as the following:

$$N_2O \text{ emission} = (\exp [A + B \times T_{soil} + C \times SM + D \times pH_{soil} + E \times N \text{ input} + F \times \text{Fertilizer type}]) \times \frac{28}{44} \quad (1)$$

Where, T_{soil} refers to soil temperature (°C), SM soil moisture (%), and the coefficients A, B, C, D, E, and F are statistically derived parameters (Table 2.2). N input is differentiated by synthetic or organic fertilizer, and is expressed as kg N ha⁻¹ yr⁻¹. The units for predicted N₂O

emission are kg N ha⁻¹ yr⁻¹. Initially, air temperature was included as a variable in the model, but it was later omitted from the equation as it has insignificant correlation with N₂O emission.

Table 2.2 summarizes the coefficients and p-values of each variable. Based on the p-value, the soil moisture, soil pH, fertilizer usage and fertilizer type are statistically significant. The Residual standard error is 0.928, and R-squared is 0.2. The F-test shows that this multivariate linear regression model is statistically significant at 90% confidence level.

Table 2.2 Summary of the N₂O_STAT model.

Variable	Parameter	Coefficient	P-value
A	Intercept	1.3437	0.0295
B	Soil temperature	0.0291	0.0515
C	Soil moisture	0.0196	0.0003
D	Soil pH	-0.3454	0.0007
E	Nitrogen input	0.0003	0.5802
F	Fertilizer type	0.4567	0.0073

2.3.2. NH₃

Similar for NH₃, we found that the data distribution, shown as the histogram of NH₃ emissions (Figure 2.3a), was skewed to the right. To normalize the data, we transformed the data to its logarithmic value (Figure 2.3b) and determined that the data were then close to a normal distribution.

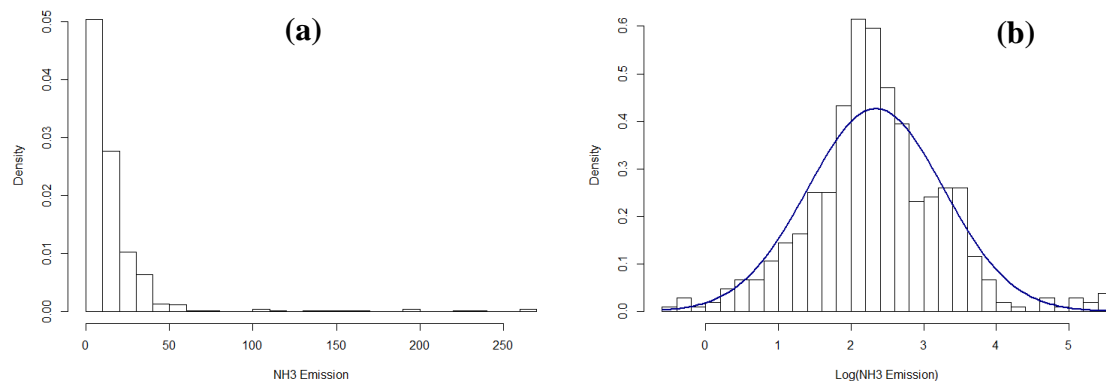


Figure 2.3. (a) Histogram of NH₃ emission; and (b) log of NH₃ emission.

Then, using the same methodology, we used a multiple linear regression model to fit the response variable (the log of NH₃ emissions), using the physicochemical variables as predictors. The statistically derived model (hereinafter: NH₃_STAT) to predict NH₃ emissions from agricultural soils is mathematically expressed as the following:

$$\text{NH}_3 \text{ emission} = (\exp [A + B \times T_{\text{soil}} + C \times \text{SM} + D \times \text{pH}_{\text{soil}} + E \times \log(\text{N input}) + F \times \text{Fertilizer type}]) \times \frac{14}{17} \quad (2)$$

Where, T_{soil} refers to soil temperature (°C), SM soil moisture (%), and the coefficients A, B, C, D, E, and F are statistically derived parameters (Table 2.3). Table 2.3 summarizes the coefficients and p-values of each variable. Based on the p-value, the soil moisture, soil pH, fertilizer usage and fertilizer type are statistically significant. The Residual standard error is 0.743, and R-squared is 0.4. F-test shows that this multivariate linear regression model is statistically significant (99% confidence level).

Table 2.3 Summary of the NH₃_STAT model.

Variable	Parameter	Coefficient	P-value
A	Intercept	-4.641412	< 2e-16
B	Soil temperature	0.015031	0.020691
C	Soil moisture	0.006613	0.008920
D	Soil pH	0.086288	0.04883
E	Log(Nitrogen input)	1.166524	< 2e-16
F	Fertilizer type	0.508622	0.000569

2.3.3. NO_x

Similar to previous Nr gases, we found that the data distribution, shown as the histogram of NO_x emissions (Figure 2.4a), was skewed to the right. To normalize the data, we transformed the data to its logarithmic value (Figure 2.4b). However, the distribution of log(NO) not appear as a normal distribution. This might due to the relatively small data set. Most data are located between -2 and 2. Therefore, we assume the data are normally distributed.

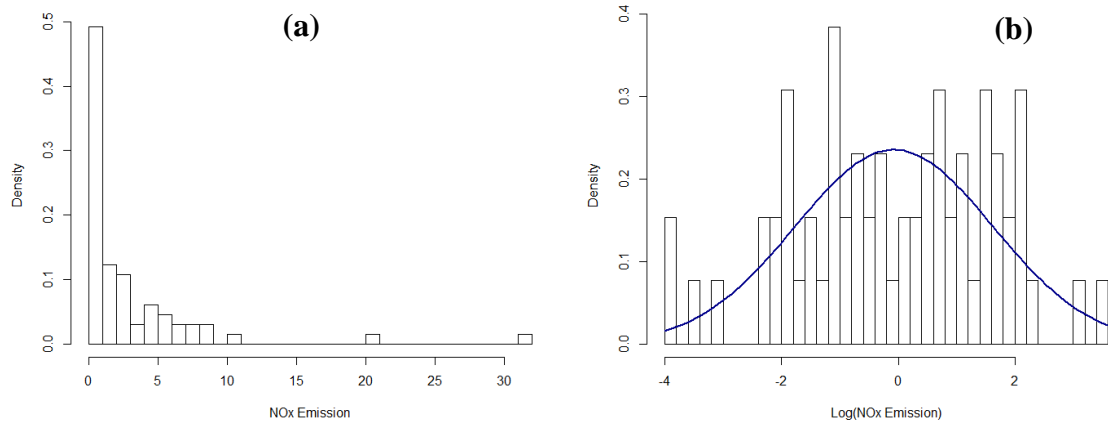


Figure 2.4. (a) Histogram of NO_x emission; and (b) log of NO_x emission.

Then, we used a multiple linear regression model to fit the response variable (the log of NO_x emissions), using the physicochemical variables as predictors. The statistically derived model (hereinafter: NO_STAT) to predict NO_x emissions from agricultural soils is mathematically expressed as the following:

$$NO \text{ emission} = (\exp [A + B \times T_{soil} + C \times SM + D \times pH_{soil} + E \times \log(N \text{ input}) + F \times Fertilizer \text{ type}]) \times \frac{14}{30} \quad (3)$$

Where, T_{soil} refers to soil temperature (°C), SM soil moisture (%), and the coefficients A, B, C, D, E, and F are statistically derived parameters (Table 2.4). Table 2.4 summarizes the coefficients and p-values of each variable. Based on the p-value, the soil moisture, soil pH, fertilizer usage and fertilizer type are statistically significant. The Residual standard error is

1.345, and R-squared is 0.4. F-test shows that this multivariate linear regression model is statistically significant (99% confidence level).

Table 2.4 Summary of the NO_STAT model.

Variable	Parameter	Coefficient	P-value
A	Intercept	-6.161950	0.0140
B	Soil temperature	0.022040	0.3375
C	Soil moisture	0.017233	0.0436
D	Soil pH	- 0.132707	0.5883
E	Log(Nitrogen input)	1.229934	1.61e-05
F	Fertilizer type	-0.077396	0.9241

2.4. Model Diagnostics

After running the regression analysis, we verified that the three models work well for fitting the data and comply with the assumptions of linear regression models. A two-step model diagnostic was performed by analyzing the variance and distribution of the residuals (Figure 2.5, Figure 2.6, and Figure 2.7). The equal distribution with no distinct patterns of residuals around the horizontal provides a good indication the likelihood of a linear relationship. Additionally, the residuals are well fitted on the straight line, indicating that the data are indeed normally distributed. The outliers indicated by residuals that are far from the line are excluded in further analysis.

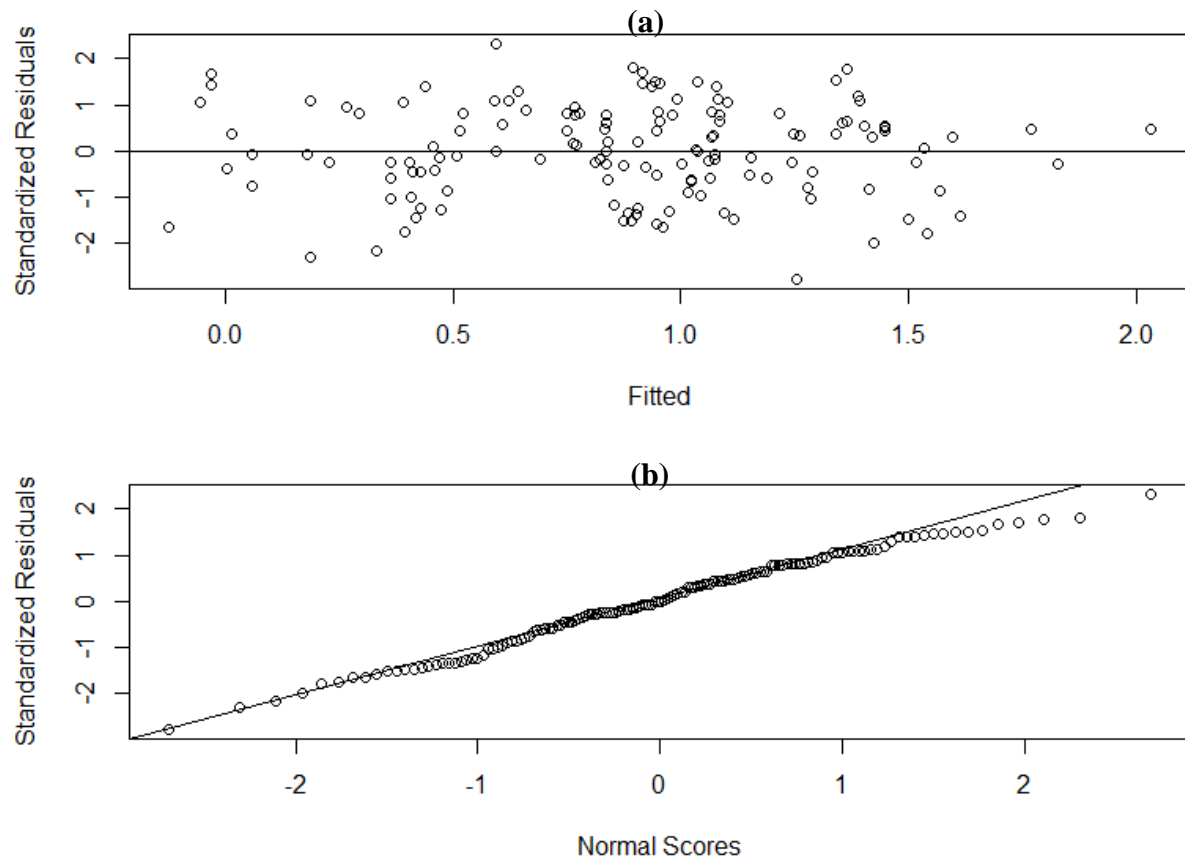


Figure 2.5. N₂O Model diagnostic: (a) Standardized residual; (b) Q-Q Plot.

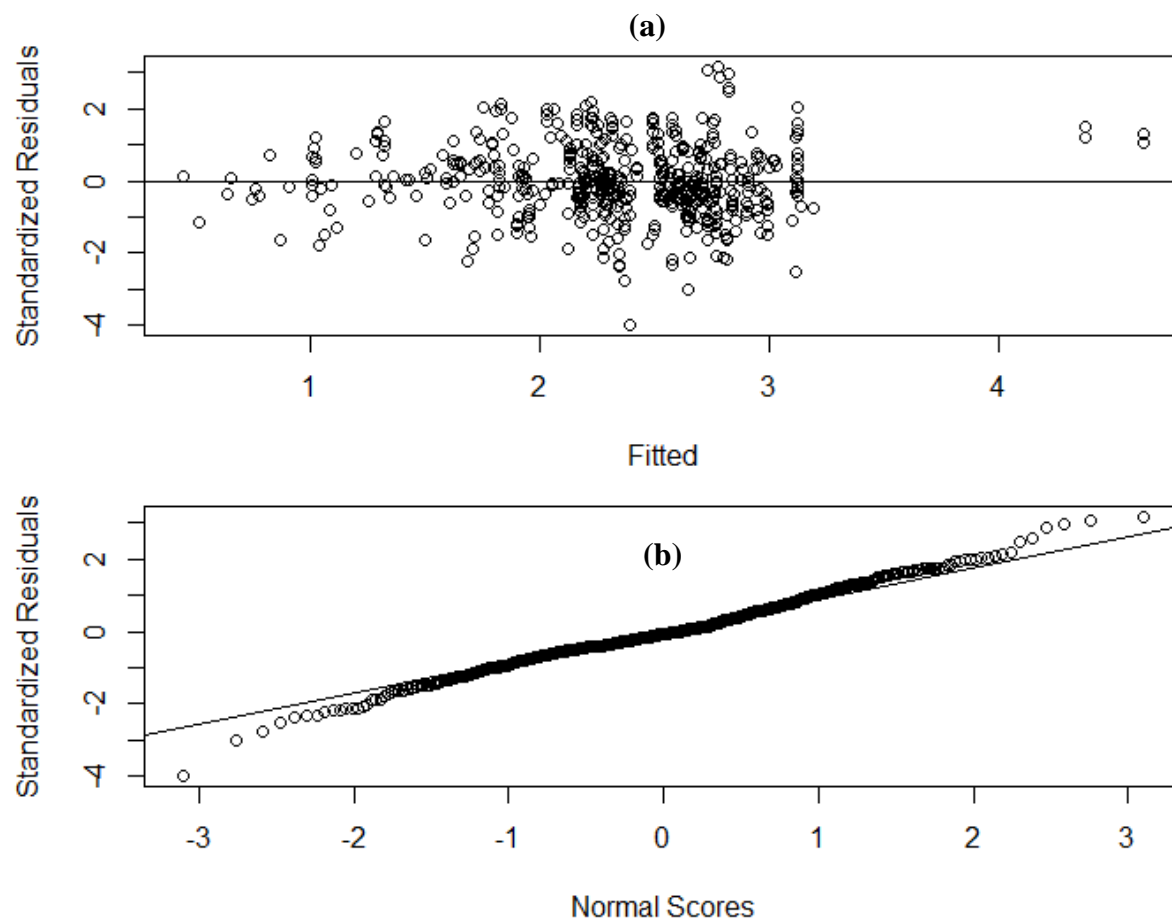


Figure 2.6. NH3 Model diagnostic: (a) Standardized residual; (b) Q-Q Plot.

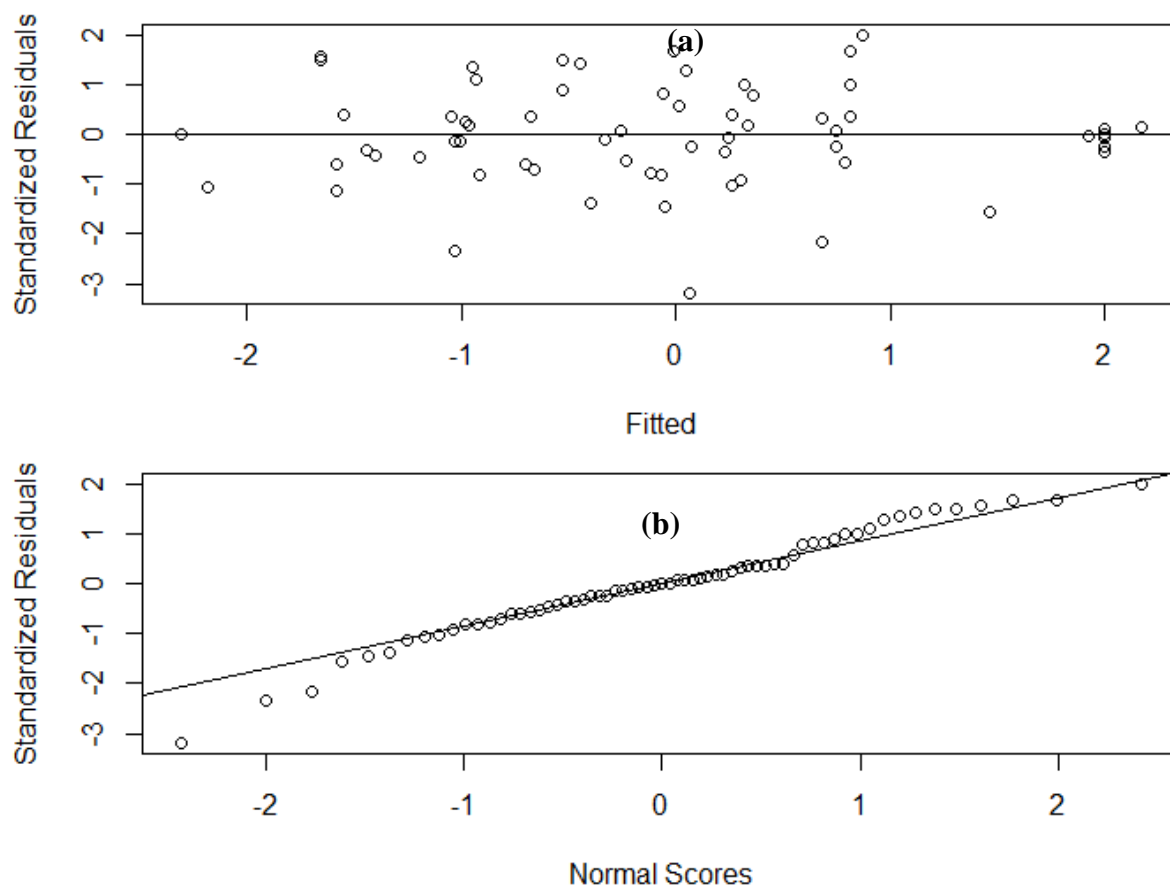


Figure 2.7. NOx Model diagnostic: (a) Standardized residual; (b) Q-Q Plot.

We tested different model forms, i.e., linear form (classical emission factor approach) and exponential form (this study). The exponential form ($r^2 = 0.2$, and residual standard error = 0.93) while the linear form ($r^2 = 0.14$, and residual standard error = 1.02), suggesting that the exponential model performs slightly better in modeling the reactive nitrogen emission.

2.5. Other Emission Inventories

EDGAR: The Emission Database for Global Atmospheric Research compiles anthropogenic global N₂O, NH₃, NO emissions and trends from 1970 to 2012 based on international statistics and emission factors (Janssens-Maenhout et al. 2017). The activity data for agricultural sectors originate mainly from International Fertilizer Industry Association (IFA)

and the Food and Agriculture Organization Statistics Division (FAOSTAT). The N₂O emission factor for direct soil emissions from the use of synthetic fertilizer and from manure and from crop residues is based on the 2006 IPCC Guidelines (IPCC, 2006). For the comparison purposes, we use the 2012 global N₂O data set for agricultural soils (subsector 4C+4D). However, for NH₃ and NO_x, we use the 2012 global data set for agriculture sectors (subsector 4B+4C+4D+4F), which includes Enteric fermentation, Manure management, Rice cultivation, and Agricultural soils.

FAOSTAT: Food and Agriculture organization of the United Nations develops methods and standards for food and agriculture statistics, provides technical assistance services and disseminates data for global monitoring. It provides the N₂O global emissions from synthetic fertilizers, manure applied to soils, manure left on pasture, and crop residues in 2012.

EPA greenhouse gas inventory report and USGS county-level data sets: We calculated N₂O emission from agriculture soils over the continental U.S. based on subsectors described in the EPA report (EPA, 2018). The subsector emissions are taken from the county-level information compiled by USGS from the 2012 census of agriculture (Gronberg and Spahr, 2012; Gronberg and Arnold, 2017). The county-level information consists of fertilizer usage, manure applications, and crop residue. Then we calculated the emissions which are interpolated to generate a spatial distribution of the emission for the United States.

EPA National Emission Inventory 2014 version 2: We calculated NH₃ emission from agricultural soils over the continental U.S. based on the county-level emission data (EPA, 2018). We selected the emission from selected source classification codes (SCCs) listed on Table 2.5. The information consists of NH₃ emissions from different types of fertilizer, manure from different types of animals, and fractions from unclassified sources.

Table 2.5. Selected NEI2014 v.2 source classification codes (SCCs) included in this study.

SCC	Item	Source
2801700001	Anhydrous Ammonia	Fertilizer Application
2801700002	Aqueous Ammonia	Fertilizer Application
2801700003	Nitrogen Solutions	Fertilizer Application
2801700004	Urea	Fertilizer Application
2801700005	Ammonium Nitrate	Fertilizer Application
2801700006	Ammonium Sulfate	Fertilizer Application
2801700007	Ammonium Thiosulfate	Fertilizer Application
2801700008	Other Straight Nitrogen	Fertilizer Application
2801700009	Ammonium Phosphates	Fertilizer Application
2801700010	N-P-K (multi-grade nutrient fertilizers)	Fertilizer Application
2801700011	Calcium Ammonium Nitrate	Fertilizer Application
2801700012	Potassium Nitrate	Fertilizer Application
2801700013	Diammonium Phosphate	Fertilizer Application
2801700014	Monoammonium Phosphate	Fertilizer Application
2801700015	Liquid Ammonium Polyphosphate	Fertilizer Application
2801700099	Miscellaneous Fertilizers	Fertilizer Application
2805001300	Land application of manure	Beef cattle - finishing operations on feedlots (drylots)
2805002000	Not elsewhere classified	Beef cattle production composite (93%)
2805007300	Land application of manure	Poultry production - layers with dry manure management systems
2805008300	Land application of manure	Poultry production - layers with wet manure management systems
2805009300	Land application of manure	Poultry production - broilers
2805010300	Land application of manure	Poultry production - turkeys
2805018000	Not elsewhere classified	Dairy cattle production composite (26%)
2805019300	Land application of manure	Dairy cattle - flush dairy
2805021300	Land application of manure	Dairy cattle - scrape dairy
2805022300	Land application of manure	Dairy cattle - deep pit dairy
2805023300	Land application of manure	Dairy cattle - drylot/pasture dairy
2805025000	Not elsewhere classified	Swine production composite (4%)
2805030000	Not elsewhere classified	Poultry production composite (7%)
2805035000	Not elsewhere classified	Horse production composite (26%)
2805039300	Land application of manure	Swine production - operations with lagoons (unspecified animal age)
2805045000	Not elsewhere classified	Goats production composite (26%)
2805047300	Land application of manure	Swine production - deep-pit house operations (unspecified animal age)

CHAPTER 3 Characteristic of N₂O Emission

3.1. Global

After calculating the coefficients of all variables, we applied the N₂O_STAT model for each grid cell and generated a global map for N₂O emissions. Figure 3.1(a) gives the spatial distribution of global N₂O emission from agricultural soils calculated using N₂O_STAT in kg N₂O yr⁻¹ grid cell⁻¹. The resolution of this map is 5 arc-minute by 5 arc-minute, which is equivalent to 8464 ha. The average N₂O emission from agricultural soils is ~9000 kg yr⁻¹ grid cell⁻¹, which is comparable to average values of 1 kg ha⁻¹ for agricultural soils, reported by Kim et al. (2013) and Shcherbak et al. (2014). Figure 3.1(b) is spatial distribution of global N₂O emission from agricultural soil based on EDGAR. Total annual global N₂O emissions from N₂O_STAT and EDGAR are 3.06 Tg N yr⁻¹ and 4.49 Tg N yr⁻¹, respectively. It is encouraging to see that, in general, the model captures the global spatial pattern in N₂O emissions well. Figure 3.1 (c) shows the absolute difference between N₂O_STAT and EDGAR. In comparison with EDGAR, N₂O_STAT gives relatively lower N₂O emission values in North America, South America, India, and China, while higher values in Eastern Europe and mid Africa.

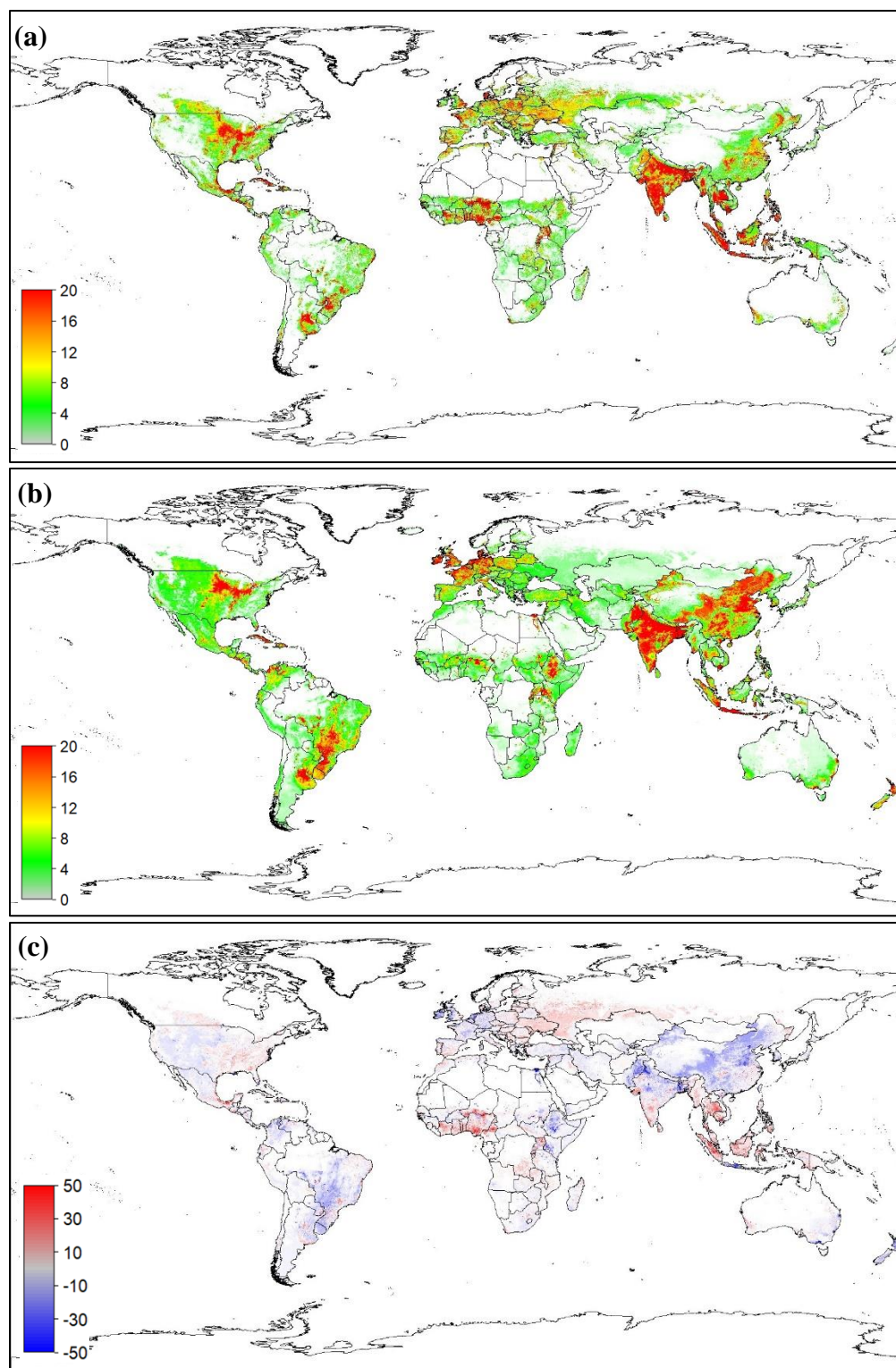


Figure 3.1. Comparison between the results from (a) N₂O_STAT, (b) EDGAR, and (c) absolute difference between the two models (in ton N₂O yr⁻¹ grid cell⁻¹).

3.2. Regional

3.2.1. *Continental U.S.*

Figure 3.2 provides the comparison between the results from N2O_STAT, EDGAR, and EPA/USGS over the continental U.S. (hereafter listed as U.S). Total N2O emissions from agricultural soils in the U.S. calculated from N2O_STAT, EDGAR, and EPA/USGS are 0.35 Tg N yr⁻¹, 0.43 Tg N yr⁻¹, and 0.46 Tg N yr⁻¹, respectively. Our model performs well in capturing the spatial pattern as well as the total annual emission. Figure 3.3 gives the absolute difference of N2O emissions between N2O_STAT model and EDGAR, and between N2O_STAT and EPA/USGS. Based on Figure 3.3(a), N2O_STAT underestimates N2O emission in the Midwest Region of the U.S., and overestimates emissions in the southeastern region, compared with EDGAR. Figure 3.3(b) shows that N2O_STAT gives lower N2O emission values in northern Texas, California, Florida, and states around Lake Michigan, while in other areas the values are comparable.

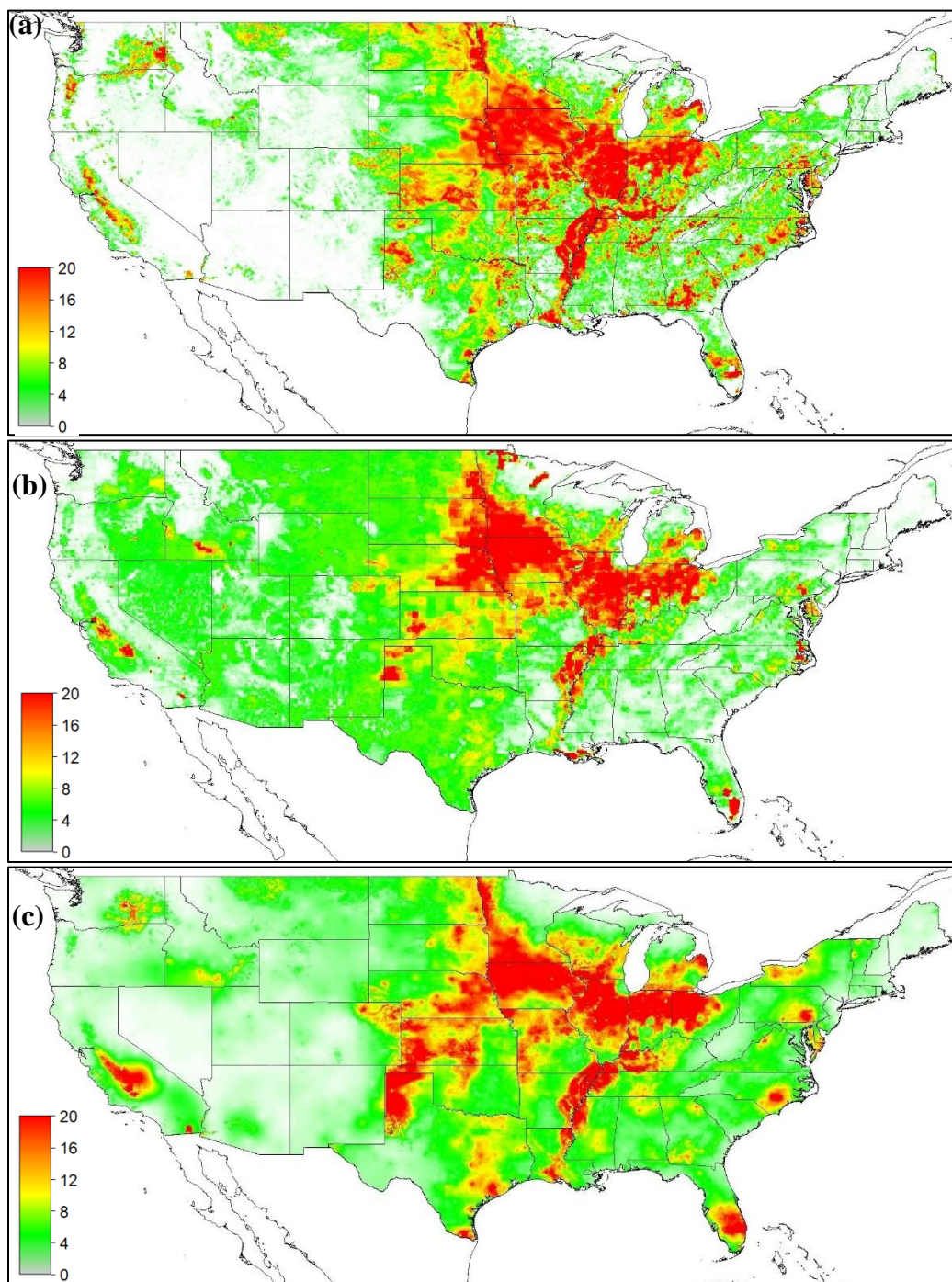


Figure 3.2. Comparison between the results from (a) N₂O_STAT, (b) EDGAR, and (c) EPA/USGS (in ton N₂O yr⁻¹ grid cell⁻¹).

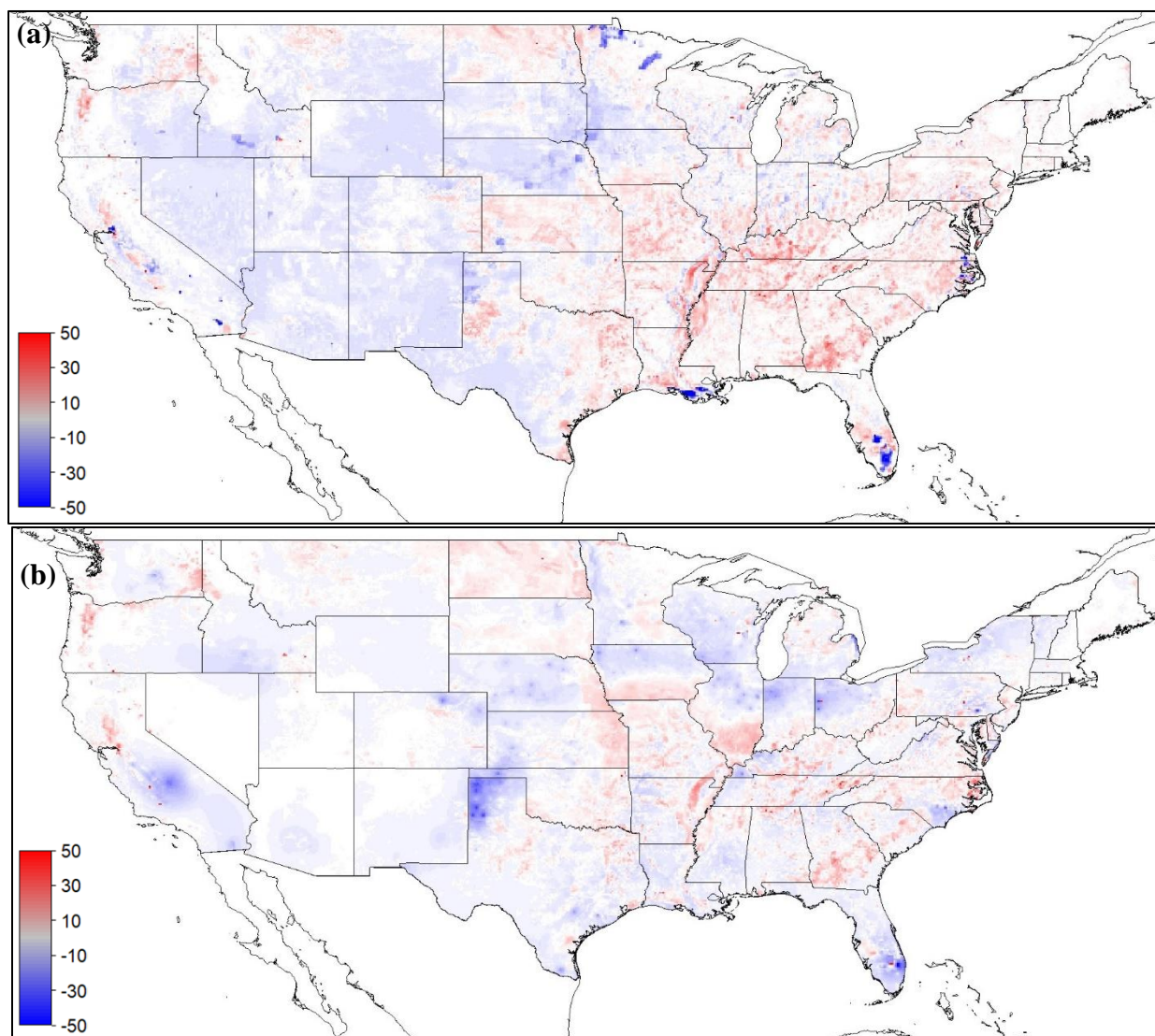


Figure 3.3. Absolute difference of N₂O emissions over CONUS between (a) N₂O_STAT and EDGAR, (b), between N₂O_STAT and EPA/USGS (in ton N₂O yr⁻¹ grid cell⁻¹).

3.2.2. India

Figure 3.4 and Table 3.1 show close agreement between the N₂O_STAT estimate of agricultural soil emissions for India when compared with EDGAR (~ -14%), the finely-resolved N₂O emission estimate by Aneja et al. (2012) (~17%); and FAOSTAT (~ -7%). Moreover, Table 3.1 compares the finely-resolved N₂O emission for India with other published studies. The Aneja et al. (2012) results are higher than the results of Garg et al. (2006) and Sharma et al.

(2011). The reason might be due to the selection of higher emission factors in Aneja et al. (2012) study. Overall, we believe that Aneja et al. (2012) is more appropriate in the sense that the researchers used the activity level data at district levels and chose the emission factors suitable for Asian context

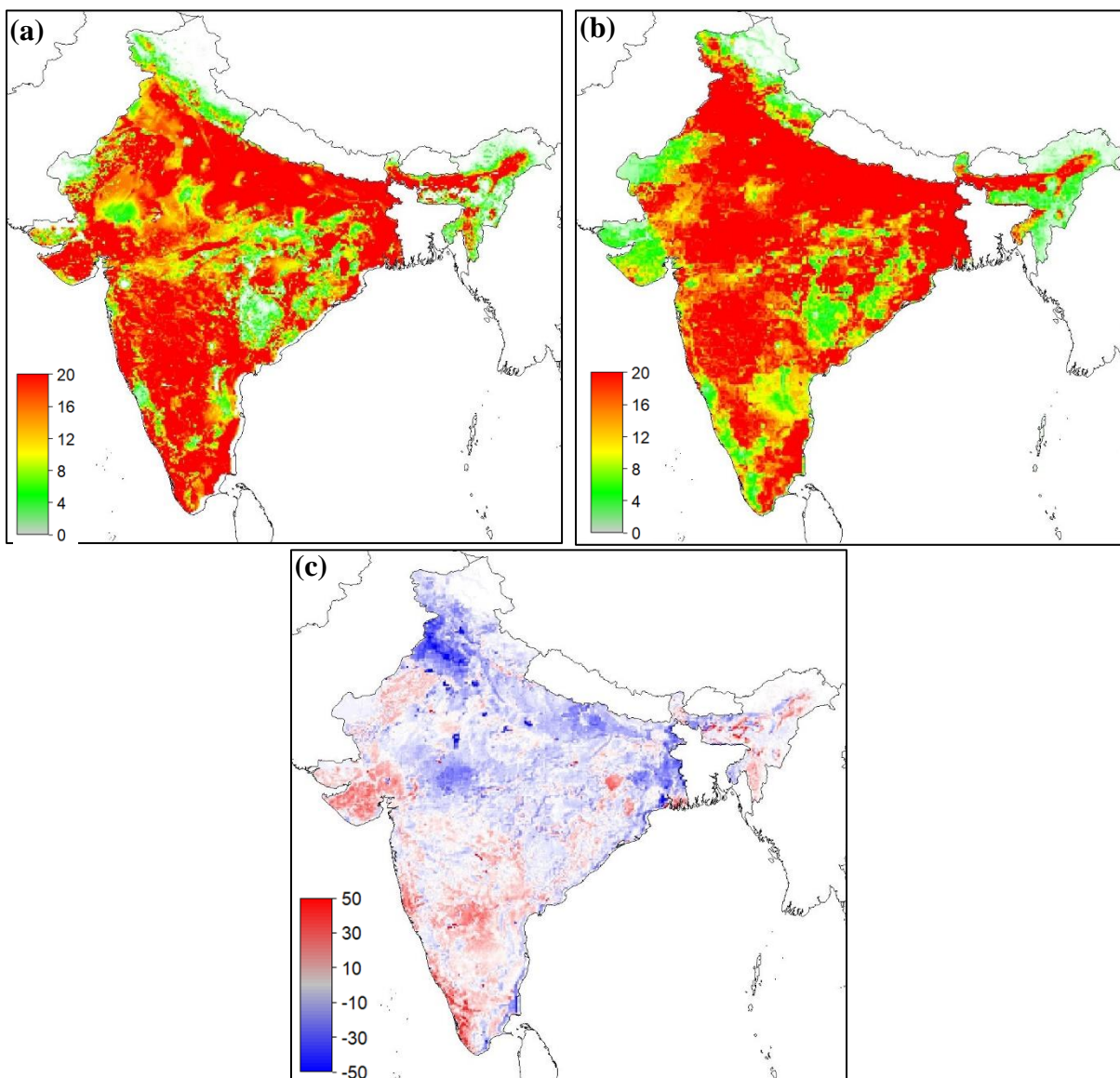


Figure 3.4. Comparison between the results from (a) N₂O_STAT, (b) EDGAR, and (c) the absolute difference between N₂O_STAT and EDGAR (in ton N₂O yr⁻¹ grid cell⁻¹).

3.2.3. China

For China, our estimate of N₂O emissions is 2.3-2.8 times lower than those estimated by EDGAR and FAO. Our N₂O emission estimate, however, is in better agreement with Gao et al. (2011) and Zhou et al. (2014), being 2% higher and 38% lower, respectively. These two studies employ different methods in estimating N₂O emission from Chinese agricultural soils. Gao et al. (2011) studied N₂O direct emissions from croplands by using localized emission factors; while Zhou et al. (2014), in addition to using regional emission factors, utilized high-resolution activity data and localized parameters.

Table 3.1. Summary of N₂O emission from different inventories.

Emission inventory	N ₂ O (Tg N yr ⁻¹)			
	Global	US	India	China
N ₂ O_STAT	3.75	0.400	0.412	0.300
EDGAR	4.49	0.432	0.468	0.832
FAOSTAT	4.07	0.350	0.440	0.686
EPA/USGS	-	0.457	-	-
EPA Report	-	0.529	-	-
Aneja et al. (2012)	-	-	0.344	-
Garg et al. (2006)	-	-	0.181	-
Sharma et al. (2011)	-	-	0.226	-
Gao et al. (2011)	-	-	-	0.294
Zhou et al. (2014)	-	-	-	0.414

3.3. Conclusions

Table 3.1 summarizes the data comparison between N₂O_STAT, EDGAR, EPA/USGS, and FAOSTAT for N₂O emission from agricultural soils for the U.S., India, China, and globally. The FAOSTAT, EDGAR and EPA estimates are slightly different, although statistically consistent given the large uncertainties in the IPCC default methodologies (Tubiello et al., 2013). Tubiello et al. (2013) compared N₂O emission from four databases. The results show that the

EPA 2006 and EPA 2011 provided the highest values while FAOSTAT gave the lowest, consistent with our study.

The region-based analysis (U.S., India, and China) indicates that our model gives lower estimates than other data sets. The global N₂O agricultural soil emission from N₂O_STAT estimate is 9-20% lower than EDGAR and FAOSTAT. The major differences, as observed in Figure 6(c), are most prominent in India and China. In India, negative differences are depicted in Figure 9(c) in the northern and central regions while the positive deviations are mainly to the west and south of the subcontinent. However, the finely-resolved agricultural emissions for N₂O (Aneja et al, 2012), as well as EDGAR and FAOSTAT, agrees well with N₂O_STAT (< 20% differences). It is to be noted that the estimate from Aneja et al. (2012) was calculated for 2003. Despite this difference, there are similarities found between these two data sets with respect to the areas from which high emissions are estimated. Such similarities are not observed when we compared our model estimate with EDGAR.

The underestimation of N₂O emissions in comparison to other data sets and the fact that N₂O_STAT may be still missing some key N inputs to its variables should be taken into account in the future work. Another component worth investigating is soil biological activity that would more fully represent the processes governing the N₂O emissions.

Supplementary information

Table 1S. List of publications from where N₂O emissions and physicochemical parameters were obtained.

Soil type	Air temperature (°C)	Soil temperature (°C)	Soil moisture (%)	Soil pH	Usage (kg N ha ⁻¹)	Fertilizer type (0:synthetic fertilizer; 1: manure)	Annual N ₂ O emission (kg N ₂ O ha ⁻¹)	Reference
grassland	20	21.6	28.9	6	63.9	1	1.6	Dittert et al., 2001
grassland	12.4	14.8	79	6	100	0	1.4	Ganesan et al., 2015
grassland	11.7	14.2	64	6	100	0	0.8	Ganesan et al., 2015
grassland	2.8	6.3	82	6	100	0	0.9	Ganesan et al., 2015
grassland	10.2	12.9	75	6	100	0	2	Ganesan et al., 2015
grassland	12.3	14.7	77	6	100	0	2.8	Ganesan et al., 2015
grassland	2.1	5.7	91	6	100	0	0.3	Ganesan et al., 2015
paddy	35	34.9	23.1	7.6	120	0	1	Ghosh et al., 2003
upland	28	28.7	30.4	4.6	90	0	6.9	Hadi et al., 2008
upland	27	15	50	5.7	90	0	1.9	Jumadi et al., 2008
grassland	14.8	15.6	35	4.9	1000	1	6.25	Kelly et al., 2008
grassland	18.3	20.2	40	4.9	1000	1	4.99	Kelly et al., 2008
paddy	35	34.9	24.1	7.8	140	0	0.16	Kumar et al., 2000
grassland	10.7	11.4	80	5.1	270	1	11.5	Merino et al., 2005
grassland	20.4	16.7	60	5.1	194	1	9.1	Merino et al., 2005
paddy	35	34.9	60	7.9	140	0	0.06	Majumdar et al., 2000
paddy	35	34.9	45	8.1	120	0	0.76	Malla et al., 2005
paddy	35	34.9	66.7	8	120	0	0.66	Malla et al., 2005
grassland	18.2	20	50	6.6	97	0	4.4	Menendes etl al., 2006
grassland	18.2	20	60	6.6	181	1	15.5	Menendes etl al., 2006
grassland	15.9	17.9	80	7	80	0	1.2	Merino et al., 2002
grassland	15.9	17.9	80	7	85	1	1.2	Merino et al., 2002
winter wheat	9	9.2	28.9	5.1	160	0	0.5	Misselbrook et al., 2014
silage	9.7	10.9	28.9	5.1	120	0	1.5	Misselbrook et al., 2014
silage	11.1	7.9	28.9	5.8	120	0	1.1	Misselbrook et al., 2014
winter wheat	9.1	9.9	28.9	5.8	200	0	3.1	Misselbrook et al., 2014
winter barley	8.3	8.6	28.9	5.1	160	0	1.7	Misselbrook et al., 2014
grassland	9.9	16	28.9	5.1	625	1	2.4	Misselbrook et al., 2014
grassland	9.9	14.2	28.9	5.1	488	1	1.9	Misselbrook et al., 2014
grassland	10.3	8.7	28.9	5.1	470	1	1.6	Misselbrook et al., 2014

Table 1S. (continued).

grassland	9.5	14.5	28.9	5.1	365	1	2.8	Misselbrook et al., 2014
grassland	9.6	14	28.9	5.1	181	1	2.7	Misselbrook et al., 2014
grassland	10.7	8.3	28.9	5.1	167	1	0.8	Misselbrook et al., 2014
grassland	9.8	16.3	28.9	5.1	106	1	0.8	Misselbrook et al., 2014
grassland	9.1	6.8	28.9	5.1	146	1	0.8	Misselbrook et al., 2014
maize	16.7	20	39	6.5	224	0	3.6	Adviento-Borbe et al., 2010
maize	16.7	20	39	6.5	225	1	5.5	Adviento-Borbe et al., 2010
vegetable	15	15	31.8	5.9	150	0	0.4	Baruah et al., 2010
vegetable	15	15	31.8	5.9	150	1	1.8	Baruah et al., 2010
vegetable	15	15	31.8	5.9	150	1	0.6	Baruah et al., 2010
rapeseed	13.8	15	27.6	7	120	0	2.48	Asgedom et al., 2014
rapeseed	13.8	15	27.6	7	129	1	1.19	Asgedom et al., 2014
maize	4.2	7.6	60	6.3	171	1	8.2	Chantigny et al., 2012
maize	4.2	7.6	60	6.3	146	1	4	Chantigny et al., 2012
maize	4.2	7.6	60	6.3	130	1	6.4	Chantigny et al., 2012
maize	4.2	7.6	60	6.3	109	1	2.4	Chantigny et al., 2012
maize	4.2	7.6	60	6.3	149	1	2.5	Chantigny et al., 2012
maize	4.2	7.6	60	6.3	137	1	3.8	Chantigny et al., 2012
maize	4.2	7.6	60	6.3	124	1	1.7	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	151	1	4	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	148	1	4.7	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	136	1	3.5	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	151	1	2.3	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	149	1	1.8	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	156	1	2.8	Chantigny et al., 2012
maize	4.2	7.6	55	6.7	156	1	1.3	Chantigny et al., 2012
maize	1.5	5.2	40	5.9	150	1	4.4	Chen et al, 2014
maize	1.5	5.2	40	5.9	150	1	3.14	Chen et al, 2014
maize	1.5	5.2	40	5.9	150	0	2.82	Chen et al, 2014
maize	19.5	21.1	14.4	6.7	224	0	0.7	Collin et al, 2011
maize	18.9	20.6	15.6	6.7	224	0	1.4	Collin et al, 2011
maize	19.5	21.1	14.4	6.7	224	1	1.1	Collin et al, 2011
maize	18.9	20.6	15.6	6.7	224	1	1.4	Collin et al, 2011

Table 1S. (continued).

wheat and maize	13.9	15.7	44.9	8	150	0	4.4	Ding et al, 2013
wheat and maize	13.9	15.7	44.9	8	150	0	3.46	Ding et al, 2013
wheat and maize	13.9	15.7	44.9	8.1	150	1	3.77	Ding et al, 2013
maize	19.7	21.3	51.1	6.2	230	0	3	Guo et al, 2013
maize	19.5	21.1	46.7	6.2	230	0	1.8	Guo et al, 2013
maize	20	21.1	37.2	5.9	120	0	1.7	Li et al., 2013
grassland	5.6	8.8	39	5.8	98	0	0.3	Shimizu et al., 2013
grassland	6.1	9.2	39	5.8	100	0	0.6	Shimizu et al., 2013
grassland	6.2	9.3	39	5.8	100	0	1	Shimizu et al., 2013
grassland	6.1	9.2	39	5.8	162	0	0.5	Shimizu et al., 2013
grassland	6.2	9.3	39	5.8	157	0	1	Shimizu et al., 2013
grassland	6.5	9.6	39	5.8	160	0	5.5	Shimizu et al., 2013
grassland	5.6	8.8	39	5.8	208	1	0.6	Shimizu et al., 2013
grassland	6.1	9.2	39	5.8	271	1	1.8	Shimizu et al., 2013
grassland	6.2	9.3	39	5.8	242	1	9.5	Shimizu et al., 2013
grassland	6.1	9.2	39	5.8	162	1	0.7	Shimizu et al., 2013
grassland	6.2	9.3	39	5.8	203	1	0.7	Shimizu et al., 2013
grassland	6.5	9.6	39	5.8	121	1	0.7	Shimizu et al., 2013
grassland	8.4	11.3	58	5.3	164	0	2.8	Shimizu et al., 2013
grassland	8.1	11	58	5.3	183	0	2.9	Shimizu et al., 2013
grassland	9.1	11.9	58	5.3	74	0	1.5	Shimizu et al., 2013
grassland	8.3	11.2	58	5.3	74	0	2.1	Shimizu et al., 2013
grassland	8.6	11.5	58	5.3	91	0	1.2	Shimizu et al., 2013
grassland	8.4	11.3	58	5.3	236	1	3.6	Shimizu et al., 2013
grassland	8.1	11	58	5.3	310	1	4.9	Shimizu et al., 2013
grassland	9.1	11.9	58	5.3	331	1	2.2	Shimizu et al., 2013
grassland	8.3	11.2	58	5.3	308	1	0.9	Shimizu et al., 2013
grassland	8.6	11.5	58	5.3	491	1	1.4	Shimizu et al., 2013
grassland	8.4	11.3	49	5.5	90	0	2.5	Shimizu et al., 2013
grassland	8.5	11.4	49	5.5	100	0	2.4	Shimizu et al., 2013
grassland	8.5	11.4	49	5.5	89	0	5.2	Shimizu et al., 2013
grassland	8.4	11.3	49	5.5	205	1	4.8	Shimizu et al., 2013
grassland	8.5	11.4	49	5.5	241	1	1.7	Shimizu et al., 2013
grassland	8.5	11.4	49	5.5	152	1	4.9	Shimizu et al., 2013
grassland	12.3	14.7	48	5.3	209	0	4.7	Shimizu et al., 2013
grassland	11.7	14.2	48	5.3	198	0	9.1	Shimizu et al., 2013
grassland	12.6	15	48	5.3	186	0	10.1	Shimizu et al., 2013
grassland	12	14.5	48	5.3	190	0	10	Shimizu et al., 2013

Table 1S. (continued).

grassland	12.2	14.7	48	5.3	190	0	3.8	Shimizu et al., 2013
grassland	12.4	14.8	48	5.3	184	0	5.7	Shimizu et al., 2013
grassland	12.3	14.7	48	5.3	76	1	7	Shimizu et al., 2013
grassland	11.7	14.2	48	5.3	195	1	11	Shimizu et al., 2013
grassland	12.6	15	48	5.3	123	1	11.9	Shimizu et al., 2013
grassland	12	14.5	48	5.3	253	1	5.4	Shimizu et al., 2013
grassland	12.2	14.7	48	5.3	219	1	1.9	Shimizu et al., 2013
grassland	12.4	14.8	48	5.3	173	1	6.6	Shimizu et al., 2013
grassland	16.4	18.4	39	6.1	164	0	1.8	Shimizu et al., 2013
grassland	16.1	18.1	39	6.1	233	0	3.1	Shimizu et al., 2013
grassland	16.8	18.7	39	6.1	218	0	5.1	Shimizu et al., 2013
grassland	16	18	39	6.1	194	0	15.2	Shimizu et al., 2013
grassland	16.4	18.4	39	6.1	160	0	4.9	Shimizu et al., 2013
grassland	16.4	18.4	39	6.1	119	1	11.2	Shimizu et al., 2013
grassland	16.1	18.1	39	6.1	284	1	5.3	Shimizu et al., 2013
grassland	16.8	18.7	39	6.1	242	1	6.1	Shimizu et al., 2013
grassland	16	18	39	6.1	272	1	10.7	Shimizu et al., 2013
grassland	16.4	18.4	39	6.1	310	1	7.8	Shimizu et al., 2013
rapeseed	18.6	20.3	37.5	5.2	32.5	1	5.4	Nyamadzawo et al, 2014
rapeseed	18.6	20.3	37.5	5.2	60	0	12.6	Nyamadzawo et al, 2014
rapeseed	18.6	20.3	37.5	5.2	65	1	6.8	Nyamadzawo et al, 2014
rapeseed	18.6	20.3	37.5	5.2	120	0	12	Nyamadzawo et al, 2014
rapeseed	18.6	20.3	37.5	5.2	97.5	1	19.3	Nyamadzawo et al, 2014
rapeseed	18.6	20.3	37.5	5.2	240	0	112	Nyamadzawo et al, 2014
maize	4.4	7.7	50	4.6	90	0	7.4	Pelster et al, 2012
maize	4.4	7.7	50	4.6	90	1	6.7	Pelster et al, 2012
maize	4.4	7.7	50	4.6	92.2	1	6.4	Pelster et al, 2012
maize	4.4	7.7	50	4.6	88.2	1	6.9	Pelster et al, 2012
Rye	18	20	25	4.3	90	0	0.6	Sosulski et al, 2014
Rye	18	20	25	4.3	100	1	0.7	Sosulski et al, 2014
rice	23	24.2	25.2	7.9	180	0	0.2	Wang et al, 2013
rice	23	24.2	25.2	7.9	180	1	0.2	Wang et al, 2013
vegetable	17.7	22.8	15.2	4.6	144	0	4.4	Watanabe et al, 2014
wheat	12.9	10.9	17.1	5.7	90	1	2.51	Zhai et al, 2011
wheat	12.9	10.9	17.1	5.7	90	0	1	Zhai et al, 2011
wheat	12.9	10.9	17.1	5.7	90	0	1	Zhai et al, 2011
wheat	12.9	10.9	17.1	5.7	90	0	0.53	Zhai et al, 2011
maize	21.1	23.1	12.4	5.7	90	1	4.46	Zhai et al, 2011
maize	21.1	23.1	12.4	5.7	210	0	1.16	Zhai et al, 2011

Table 1S. (continued).

maize	21.1	23.1	12.4	5.7	210	0	0.84	Zhai et al, 2011
maize	21.1	23.1	12.4	5.7	210	0	0.56	Zhai et al, 2011
maize and wheat	17.3	15	60	8.3	280	0	3.14	Zhou et al, 2014
maize and wheat	17.3	15	60	8.3	280	1	5.97	Zhou et al, 2014

CHAPTER 4 Characteristic of NH₃ Emission

4.1. Global

After calculating the coefficients of all variables, we applied the NH₃_STAT model for each grid cell (5 arc-minute x 5 arc-minute) and generated a global map for NH₃ emissions. Figure 4.1(a) gives the spatial distribution of global NH₃ emission from agricultural soils calculated using NH₃_STAT in kg NH₃ yr⁻¹ grid cell⁻¹. The resolution of this map is 5 arc-minute, which is equivalent to 8464 ha. Figure 4.1(b) is spatial distribution of global NH₃ emission from agricultural soil based on EDGAR. Total annual global NH₃ emissions from NH₃_STAT and EDGAR are 14.2 Tg N/yr and 33.3 Tg N/yr, respectively. There is a large difference between our estimate and EDGAR. This difference can be attributed to sources that are excluded in our model. Nevertheless, it is encouraging to see that, in general, the model captures the global spatial pattern in NH₃ emissions well. This is understandable since NH₃_STAT is exclusive to emissions from fertilizer and manure applied as fertilizer, whereas EDGAR has additional sources in their estimate. Based on our literature survey, NH₃ emission from fertilizers ranges from 9 to 12 Tg N/yr, a range that is similar to our estimate.

Figure 4.1 (c) shows the absolute difference between NH₃_STAT and EDGAR. In comparison with EDGAR, NH₃_STAT gives relatively lower NH₃ emission values in North America, South America, India, and China, while higher values in Eastern Europe.

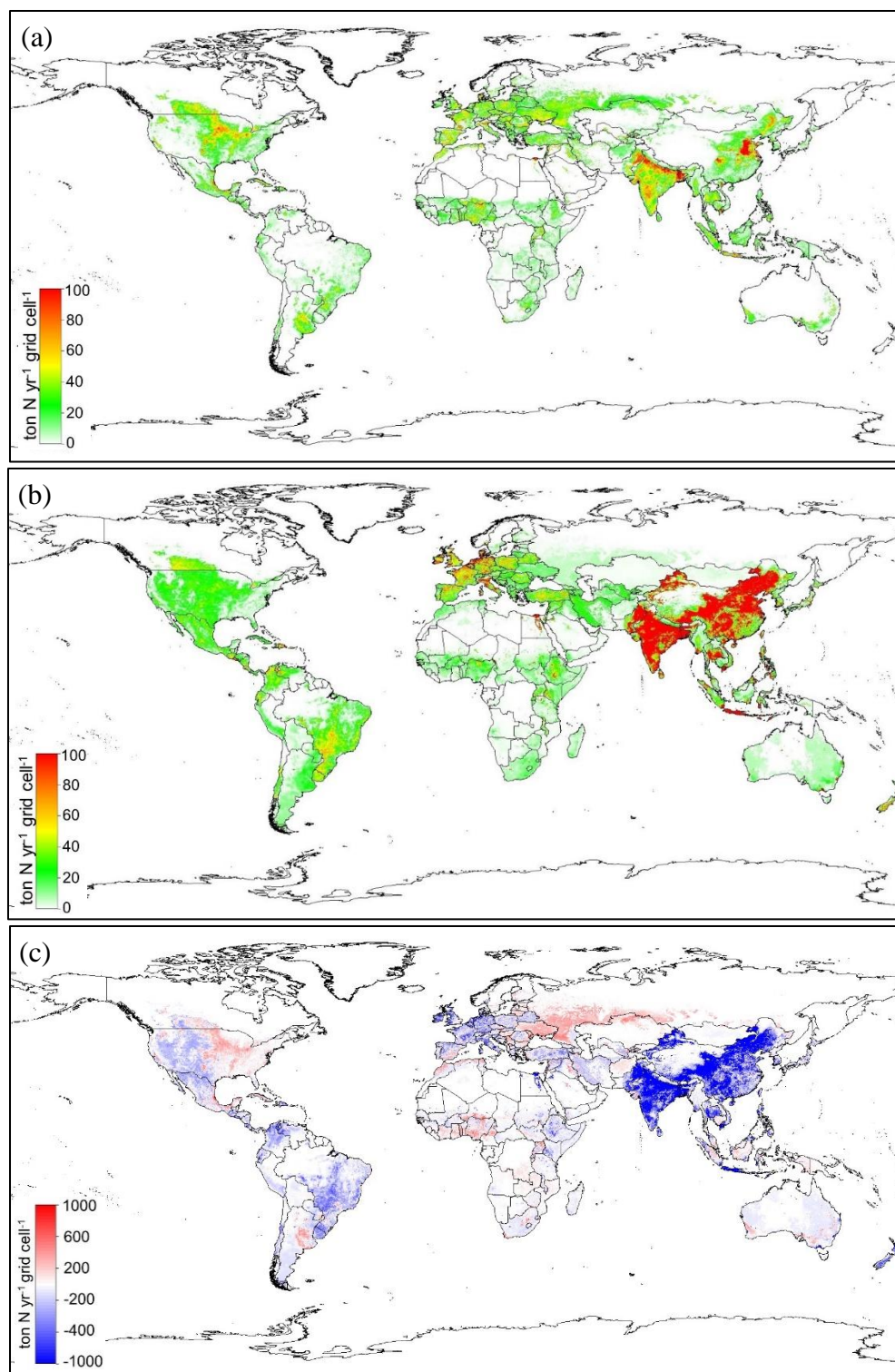


Figure 4.1. Comparison between the results from (a) NH3_STAT, (b) EDGAR, and (c) absolute difference between the two models (in $\text{ton N yr}^{-1} \text{ grid cell}^{-1}$).

4.2. Regional

4.2.1. Continental U.S.

Figure 4.2 provides the comparison between the results from NH3_STAT over U.S. Total U.S. NH3 emission from agricultural soil calculated from NH3_STAT and EPA are 1.7 Tg N/yr and 0.7 Tg N/yr, respectively. Our model produces an estimate that is 143% higher in comparison to EPA. Our model performs well in capturing the spatial trend as well as the total annual emission. Figure 8(c) gives the absolute difference of NH3 emissions between NH3_STAT model and EDGAR, and between NH3_STAT and EPA/USGS. Based on Figure 8(a), NH3_STAT underestimates N2O emission in California and some parts in Arizona, New Mexico, and Texas, while shows overestimations for most of the U.S., compared with EPA.

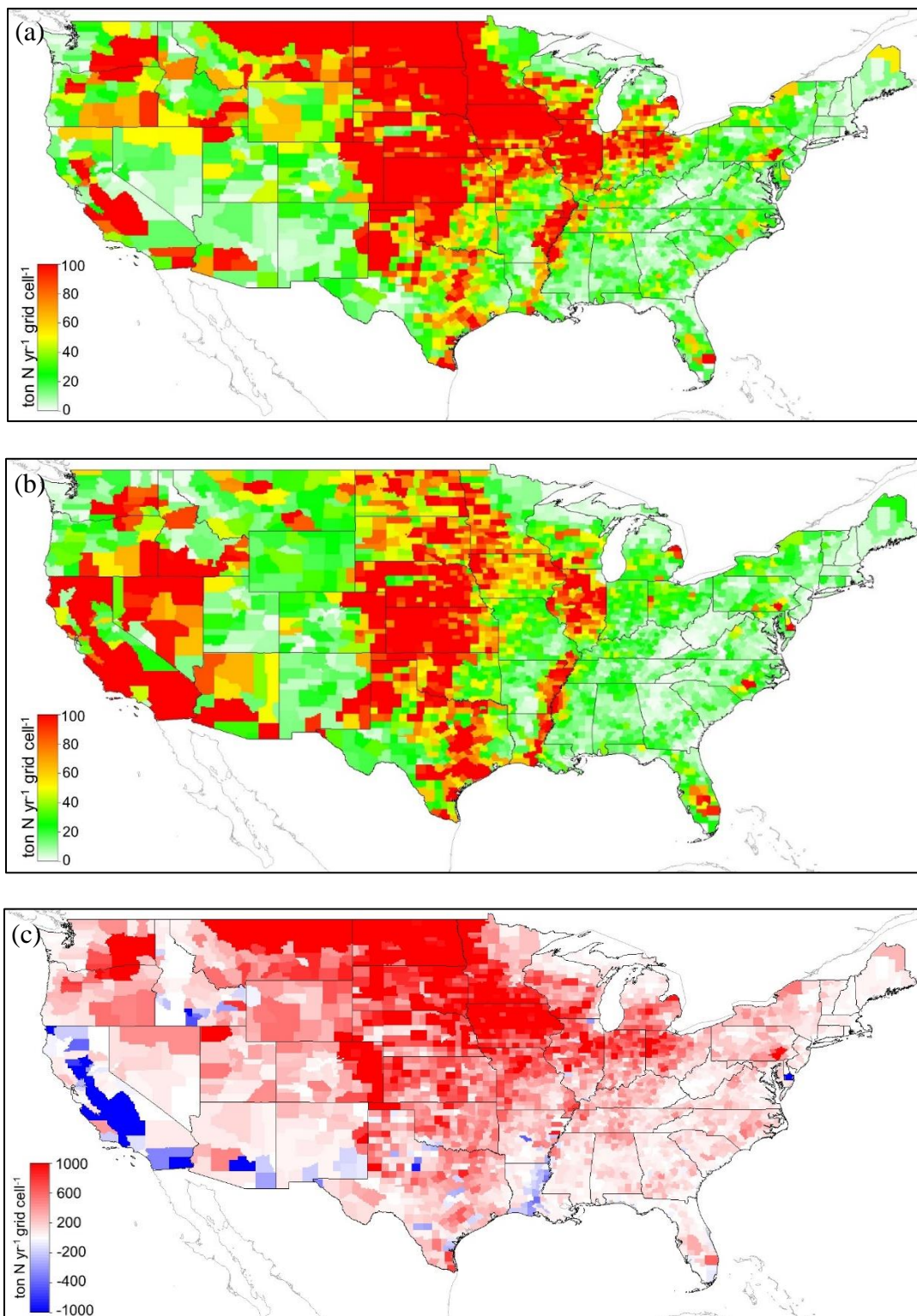


Figure 4.2. Comparison between the results from (a) NH3_STAT, (b) EDGAR, and (c) absolute difference between the two models (in $\text{ton N yr}^{-1} \text{ grid cell}^{-1}$).

4.2.2. Indian and China

We also performed a region-based analysis in India, and China) using the NH₃_STAT model. The NH₃_STAT model estimate for India shows NH₃ emissions between -141% and -77% when compared to other data sets. A lower estimate is also seen for China, where the model estimates NH₃ emissions 36-482% lower than other data sets.

4.3. Conclusions

The statistical model captures the spatial distribution of global NH₃ emissions by utilizing a more simplified approach than those used by other readily available data sets.

From global scale, the results indicate that, in comparison to other data sets, the model generates a lower global NH₃ estimate by 57%, (NH₃_STAT: 14.2 Tg N yr⁻¹; EDGAR: 33.0 Tg N yr⁻¹).

The region-based analysis (U.S., India, and China) using the NH₃_STAT model indicates that there are variations in developed country and developing country. For the U.S., our model produces an estimate that is 143% higher in comparison to EPA. Meanwhile, the NH₃_STAT model estimate for India shows NH₃ emissions between -141% and -77% when compared to other data sets. A lower estimate is also seen for China, where the model estimates NH₃ emissions 36-482% lower than other data sets. The difference in the global estimates is attributed to the lower estimates in major agricultural countries like China and India.

The underestimation of NH₃ emissions in comparison to EDGAR can be attributed to other sources that are excluded in our model. This is understandable that EDGAR has additional sources in their estimate, e.g. enteric fermentation, manure management, rice cultivation, whereas NH₃_STAT is exclusive to emissions from fertilizer and manure applied as fertilizer.

Supplementary Information

Table 2S. List of publications from where NH₃ emissions and physicochemical parameters were obtained.

Location	soil_ph	soil_moisture	soil_temperature	total_N	fertilizer_type	Nh3 emission	Reference
CA	5.3	17.0	19.4	872.1	1	268.9	ALFAM2
CA	5.3	17.0	19.4	872.1	1	264.2	ALFAM2
CA	5.3	17.0	19.4	703.8	1	237.5	ALFAM2
CA	5.3	17.0	19.4	872.1	1	224.7	ALFAM2
CA	5.3	17.0	19.4	703.8	1	194.6	ALFAM2
CA	5.3	17.0	19.4	703.8	1	193.1	ALFAM2
DK	6.3	57.0	15.1	139.7	1	166.2	ALFAM2
DK	5.5	47.0	6.7	181.9	1	151.3	ALFAM2
DK	5.5	47.0	6.7	167.8	1	149.0	ALFAM2
DK	6.3	57.0	15.2	139.7	1	135.3	ALFAM2
DK	5.5	47.0	6.7	181.9	1	112.6	ALFAM2
DK	5.5	47.0	6.7	181.9	1	104.1	ALFAM2
UK	7.9	32.4	15.4	190.4	1	103.3	ALFAM2
UK	7.9	32.4	15.4	190.4	1	74.5	ALFAM2
UK	7.9	32.4	15.4	190.4	1	63.4	ALFAM2
UK	7.9	32.4	15.4	190.4	1	58.7	ALFAM2
DK	5.6	41.0	14.0	160.4	1	54.6	ALFAM2
UK	8.0	25.9	5.8	221.4	1	54.1	ALFAM2
UK	7.1	24.7	14.4	150.0	1	54.0	Webb et al.,2004
UK	7.1	24.7	14.4	180.0	1	51.0	Webb et al.,2004
CA	6.6	30.0	13.4	159.1	1	50.3	ALFAM2
CA	6.4	24.0	9.7	150.2	1	47.5	ALFAM2
CA	6.6	30.0	13.4	101.2	1	46.2	ALFAM2
CA	6.4	24.0	9.7	139.7	1	43.4	ALFAM2
CA	6.4	24.0	9.7	139.7	1	42.3	ALFAM2
CA	6.4	24.0	9.7	150.2	1	41.6	ALFAM2
CA	6.6	30.0	13.4	159.1	1	41.3	ALFAM2
CN	8.3	40.0	18.6	119.3	0	40.6	Yang et al., 2013
CA	6.4	24.0	9.7	111.7	1	39.5	ALFAM2
CA	6.4	24.0	9.7	139.7	1	39.3	ALFAM2
UK	7.9	32.4	15.4	190.4	1	39.1	ALFAM2
CA	6.6	30.0	13.4	159.1	1	38.1	ALFAM2
CA	6.6	30.0	13.4	115.5	1	37.2	ALFAM2
CN	8.3	22.0	23.2	174.0	0	36.8	Yang et al., 2017
CA	6.6	30.0	13.4	115.5	1	36.7	ALFAM2
CA	6.4	24.0	9.7	150.2	1	36.1	ALFAM2

Table 2S. (continued).

IT	8.2	18.9	10.2	107.0	1	35.9	ALFAM2
UK	7.9	32.4	15.4	190.4	1	35.4	ALFAM2
DK	5.6	41.0	14.0	160.4	1	34.8	ALFAM2
DK	5.6	41.0	14.0	160.4	1	34.5	ALFAM2
FR	7.6	17.9	24.5	76.0	1	34.5	ALFAM2
CA	6.4	24.0	9.7	111.7	1	33.9	ALFAM2
DE	6.5	31.0	5.8	120.9	1	33.7	ALFAM2
CA	6.6	30.0	13.4	95.7	1	33.7	ALFAM2
CA	6.6	30.0	13.4	85.9	1	33.1	ALFAM2
CA	6.6	30.0	13.4	101.2	1	33.1	ALFAM2
UK	8.0	25.1	17.4	164.8	1	33.1	ALFAM2
CN	8.3	22.0	23.2	174.0	0	32.8	Yang et al., 2017
CA	6.4	24.0	9.7	142.8	1	32.7	ALFAM2
CA	6.4	24.0	9.7	139.4	1	32.3	ALFAM2
DK	5.5	47.0	6.7	167.8	1	31.7	ALFAM2
CA	6.5	24.0	16.4	152.4	1	31.7	ALFAM2
IT	6.4	14.5	12.6	223.4	1	31.6	ALFAM2
DK	5.5	47.0	6.7	204.8	1	31.4	ALFAM2
IT	6.4	14.5	12.9	223.4	1	31.2	ALFAM2
IT	6.4	21.0	18.5	109.0	1	31.1	Carozzi et al., 2013
UK	8.0	25.1	17.4	164.8	1	31.1	ALFAM2
CA	6.6	30.0	13.4	95.7	1	30.9	ALFAM2
DK	8.1	100.0	13.4	131.4	1	30.4	ALFAM2
IT	8.2	22.0	12.3	68.0	1	30.2	Carozzi et al., 2013
DE	6.5	31.0	5.8	120.9	1	30.0	ALFAM2
UK	7.1	24.7	8.4	110.0	1	30.0	Webb et al.,2004
CA	6.6	30.0	13.4	85.9	1	29.6	ALFAM2
CA	6.6	30.0	13.4	101.2	1	29.1	ALFAM2
DE	6.5	19.0	18.6	170.0	1	28.7	ALFAM2
DE	5.8	31.0	15.2	208.1	1	28.4	ALFAM2
CA	6.4	24.0	9.7	111.7	1	28.2	ALFAM2
DK	7.9	100.0	18.0	125.1	1	28.1	ALFAM2
CA	6.5	24.0	16.4	142.6	1	27.9	ALFAM2
CA	6.5	24.0	16.4	142.6	1	27.6	ALFAM2
CA	6.6	30.0	13.4	95.7	1	27.6	ALFAM2
CA	6.4	24.0	9.7	142.8	1	27.3	ALFAM2
CL	5.4	56.2	17.3	100.0	0	26.7	Salazar et al., 2012
CA	6.5	24.0	16.4	142.6	1	26.7	ALFAM2
CA	6.5	24.0	16.4	151.7	1	26.6	ALFAM2

Table 2S. (continued).

IT	7.1	13.8	12.8	188.3	1	26.4	ALFAM2
DE	6.5	19.0	18.6	170.0	1	26.4	ALFAM2
IT	7.1	17.0	11.2	95.0	1	26.4	Carozzi et al., 2013
DE	5.8	31.0	16.2	199.9	1	26.2	ALFAM2
CA	6.6	30.0	13.4	85.9	1	26.1	ALFAM2
CA	6.5	24.0	16.4	145.2	1	26.1	ALFAM2
UK	7.1	24.7	11.3	145.0	1	26.0	Webb et al.,2004
UK	7.1	24.7	11.3	105.0	1	26.0	Webb et al.,2004
DE	6.5	31.0	5.9	120.9	1	25.8	ALFAM2
CA	6.6	30.0	13.4	115.5	1	25.8	ALFAM2
DE	5.8	31.0	16.2	153.2	1	25.8	ALFAM2
CL	6.1	50.0	10.6	100.0	0	25.7	Salazar et al., 2014
CA	6.4	24.0	9.7	139.4	1	25.7	ALFAM2
CL	6.1	50.0	14.3	100.0	0	25.3	Salazar et al., 2014
CN	8.3	22.0	23.2	174.0	0	24.3	Yang et al., 2017
UK	8.0	25.9	5.8	221.4	1	24.2	ALFAM2
CA	6.1	20.0	23.3	150.0	1	24.2	ALFAM2
CA	6.4	24.0	9.7	139.4	1	24.0	ALFAM2
DE	6.6	45.0	9.1	180.0	1	24.0	Wolf et al., 2014
CA	6.5	24.0	16.4	152.4	1	24.0	ALFAM2
UK	8.0	25.1	17.4	164.8	1	23.7	ALFAM2
CN	8.3	40.0	18.6	90.5	0	23.5	Yang et al., 2013
UK	7.9	32.4	15.4	190.4	1	23.4	ALFAM2
CA	6.5	24.0	16.4	152.4	1	23.2	ALFAM2
DE	5.8	31.0	15.8	167.5	1	22.9	ALFAM2
SW	7.1	30.0	25.0	88.4	1	22.7	Hani et al., 2016
DK	7.0	100.0	12.9	108.4	1	22.6	ALFAM2
CA	6.4	24.0	9.7	142.8	1	22.5	ALFAM2
IT	7.8	10.6	25.8	240.0	0	22.4	Ferrara et al., 2014
UK	7.1	32.5	7.7	115.6	1	22.0	ALFAM2
DE	6.5	19.0	18.6	170.0	1	21.3	ALFAM2
DE	6.5	19.0	18.4	218.9	1	21.2	ALFAM2
DE	6.5	19.0	18.5	218.9	1	21.2	ALFAM2
DK	6.8	14.5	11.9	129.0	1	20.9	ALFAM2
UK	8.0	25.1	17.3	92.3	1	20.9	ALFAM2
CL	6.1	50.0	16.3	100.0	1	20.5	Salazar et al., 2014
DK	5.6	41.0	14.0	160.4	1	20.5	ALFAM2

Table 2S. (continued).

SW	7.1	30.0	11.0	90.0	1	20.4	Hani et al., 2016
CL	5.6	50.0	13.9	100.0	1	20.1	Martinez-Lagos et al., 2013
UK	7.9	32.4	15.4	190.4	1	19.9	ALFAM2
CA	6.5	23.0	12.5	144.7	1	19.9	ALFAM2
CA	6.5	24.0	16.4	151.7	1	19.3	ALFAM2
DK	5.5	47.0	6.7	167.8	1	19.2	ALFAM2
DE	5.8	31.0	16.1	134.0	1	19.2	ALFAM2
DE	6.5	31.0	6.2	143.8	1	19.1	ALFAM2
DE	6.5	19.0	18.2	182.8	1	19.0	ALFAM2
DK	6.4	16.2	11.9	110.9	1	18.9	ALFAM2
SW	7.1	30.0	10.0	93.0	1	18.9	Hani et al., 2016
CA	6.5	24.0	16.4	145.2	1	18.6	ALFAM2
UK	7.9	32.4	15.4	190.4	1	18.6	ALFAM2
SW	6.9	30.0	20.0	78.0	1	18.6	Hani et al., 2016
IT	7.1	13.8	12.8	188.3	1	18.4	ALFAM2
DE	6.5	19.0	18.2	182.8	1	18.3	ALFAM2
DK	6.6	17.0	11.9	105.3	1	18.0	ALFAM2
UK	8.0	25.9	5.8	221.4	1	17.8	ALFAM2
DE	5.8	10.0	17.5	193.0	1	17.7	ALFAM2
DE	6.5	31.0	6.2	143.8	1	17.4	ALFAM2
UK	8.0	29.6	6.2	160.2	1	17.3	ALFAM2
DE	5.8	31.0	16.1	134.0	1	17.0	ALFAM2
DE	6.5	19.0	18.2	182.8	1	16.8	ALFAM2
DE	5.8	10.0	17.1	105.9	1	16.6	ALFAM2
UK	7.9	32.4	15.4	190.4	1	16.6	ALFAM2
UK	7.2	32.5	7.8	115.6	1	16.5	ALFAM2
DE	6.5	19.0	18.5	218.9	1	16.4	ALFAM2
DK	5.5	47.0	6.7	204.8	1	16.3	ALFAM2
UK	7.2	32.5	7.9	115.6	1	16.1	ALFAM2
DE	5.8	31.0	16.1	152.8	1	16.0	ALFAM2
IT	8.1	60.0	15.5	120.0	0	16.0	Badagliacca et al., 2018
CL	6.1	50.0	16.3	100.0	0	15.9	Salazar et al., 2014
CN	8.2	23.0	13.0	140.0	0	15.8	Huo et al., 2015
CL	5.7	50.0	9.7	100.0	1	15.8	Martinez-Lagos et al., 2013
DE	5.8	31.0	15.8	192.0	1	15.8	ALFAM2
UK	8.0	28.5	9.8	126.1	1	15.7	ALFAM2
UK	8.0	29.6	6.2	160.2	1	15.7	ALFAM2

Table 2S. (continued).

DE	5.8	10.0	17.4	172.1	1	15.5	ALFAM2
CA	6.5	23.0	12.5	143.6	1	15.5	ALFAM2
DE	5.8	31.0	16.1	142.8	1	15.4	ALFAM2
UK	8.0	29.6	6.2	160.2	1	15.3	ALFAM2
DE	5.8	10.0	17.5	114.7	1	15.3	ALFAM2
DE	5.8	31.0	16.1	197.5	1	15.2	ALFAM2
SW	6.9	30.0	20.0	58.9	1	15.2	Hani et al., 2016
UK	8.0	25.1	17.3	92.3	1	15.2	ALFAM2
DE	5.8	23.0	9.7	161.4	1	15.1	ALFAM2
DE	5.8	10.0	17.5	114.7	1	15.1	ALFAM2
DE	5.8	31.0	16.2	134.0	1	15.0	ALFAM2
UK	8.0	25.9	5.8	94.7	1	14.9	ALFAM2
CA	6.1	20.0	23.3	144.7	1	14.8	ALFAM2
SW	7.2	30.0	10.0	93.0	1	14.8	Hani et al., 2016
DE	6.5	31.0	6.2	132.0	1	14.7	ALFAM2
DE	5.8	10.0	12.1	124.6	1	14.6	ALFAM2
DE	5.8	31.0	16.2	184.6	1	14.4	ALFAM2
UK	8.0	25.9	5.8	94.7	1	14.4	ALFAM2
CA	6.1	20.0	23.3	143.6	1	14.2	ALFAM2
DE	5.8	10.0	17.4	172.1	1	14.2	ALFAM2
DE	5.8	10.0	17.5	114.7	1	14.1	ALFAM2
DK	8.0	100.0	17.9	126.4	1	14.1	ALFAM2
DE	6.5	31.0	6.1	132.0	1	14.1	ALFAM2
DE	6.5	19.0	18.6	170.0	1	14.1	ALFAM2
DE	6.1	40.0	16.0	140.0	1	14.0	Gericke et al., 2011
UK	8.0	28.5	9.8	126.1	1	13.9	ALFAM2
UK	7.2	20.3	17.5	130.6	1	13.9	ALFAM2
DE	5.8	10.0	17.1	105.9	1	13.8	ALFAM2
CA	6.1	20.0	23.3	140.4	1	13.8	ALFAM2
DE	5.8	31.0	15.2	238.6	1	13.7	ALFAM2
DE	6.5	31.0	6.2	143.8	1	13.3	ALFAM2
DE	6.1	40.0	16.0	140.0	1	13.3	Gericke et al., 2011
UK	8.0	29.6	6.2	66.3	1	13.3	ALFAM2
UK	7.2	32.5	7.8	115.6	1	13.2	ALFAM2
DE	5.8	10.0	17.5	172.1	1	13.2	ALFAM2
CA	6.1	30.0	19.2	142.0	1	13.2	ALFAM2
DK	5.6	41.0	14.0	160.4	1	13.1	ALFAM2
DE	5.8	10.0	17.0	158.9	1	13.1	ALFAM2
DE	5.8	31.0	16.1	174.9	1	13.0	ALFAM2
DE	5.8	10.0	17.2	158.9	1	12.9	ALFAM2

Table 2S. (continued).

SE	7.1	24.7	15.3	67.5	1	12.9	Rodhe et al., 2006
SW	6.6	30.0	19.0	65.0	1	12.9	Hani et al., 2016
DE	5.8	31.0	16.1	197.5	1	12.9	ALFAM2
DE	5.8	31.0	15.8	167.5	1	12.7	ALFAM2
DE	6.5	40.0	15.0	140.0	1	12.6	Ni et al., 2015
DE	5.8	10.0	17.0	52.9	1	12.6	ALFAM2
DE	6.5	17.0	21.0	191.8	1	12.6	ALFAM2
CA	6.1	20.0	23.3	143.4	1	12.6	ALFAM2
DE	5.8	10.0	17.5	114.7	1	12.6	ALFAM2
DE	6.5	31.0	6.1	132.0	1	12.5	ALFAM2
DE	5.8	31.0	16.2	142.8	1	12.4	ALFAM2
DE	6.5	17.0	21.0	191.8	1	12.4	ALFAM2
DE	5.8	31.0	16.2	142.8	1	12.4	ALFAM2
DE	5.8	23.0	9.7	184.5	1	12.3	ALFAM2
CL	5.6	63.3	16.8	100.0	0	12.2	Salazar et al., 2012
DE	5.8	10.0	16.9	105.9	1	12.1	ALFAM2
UK	7.1	24.7	8.4	115.0	1	12.0	Webb et al.,2004
DE	5.8	10.0	17.5	172.1	1	11.9	ALFAM2
DK	6.1	24.9	10.7	116.0	1	11.8	ALFAM2
DE	6.5	17.0	20.7	149.5	1	11.8	ALFAM2
UK	8.0	25.2	12.0	136.6	1	11.7	ALFAM2
DK	5.6	41.0	14.0	160.4	1	11.6	ALFAM2
DE	5.8	23.0	19.8	109.6	1	11.6	ALFAM2
DE	5.8	10.0	12.1	127.8	1	11.6	ALFAM2
CA	6.5	21.0	23.0	150.0	1	11.5	ALFAM2
DE	5.8	10.0	12.1	73.0	1	11.5	ALFAM2
CA	6.5	23.0	12.5	143.4	1	11.4	ALFAM2
UK	8.0	20.2	16.9	164.0	1	11.4	ALFAM2
CA	6.5	24.0	16.4	97.4	1	11.4	ALFAM2
DE	5.8	31.0	15.3	143.3	1	11.4	ALFAM2
UK	7.1	28.3	16.1	102.0	1	11.3	ALFAM2
UK	7.2	30.6	10.5	110.1	1	11.3	ALFAM2
CA	6.5	21.0	23.0	144.7	1	11.3	ALFAM2
DE	6.5	17.0	20.8	149.5	1	11.3	ALFAM2
DE	5.8	10.0	17.4	193.0	1	11.2	ALFAM2
FR	7.6	30.7	9.2	141.4	1	11.2	ALFAM2
CN	8.2	23.0	13.0	117.0	0	11.1	Huo et al., 2015
DE	5.8	10.0	12.2	157.4	1	11.1	ALFAM2
DE	5.8	23.0	9.7	184.5	1	11.0	ALFAM2

Table 2S. (continued).

DE	5.8	10.0	12.1	127.8	1	11.0	ALFAM2
DE	6.5	17.0	20.9	191.8	1	10.9	ALFAM2
UK	7.1	32.5	7.7	115.6	1	10.9	ALFAM2
DE	5.8	31.0	16.1	184.6	1	10.9	ALFAM2
DE	5.8	10.0	12.2	157.4	1	10.8	ALFAM2
DE	6.5	17.0	20.9	149.5	1	10.7	ALFAM2
UK	8.0	25.9	5.8	94.7	1	10.7	ALFAM2
DK	6.1	22.8	10.7	109.2	1	10.7	ALFAM2
CA	6.5	21.0	23.0	143.6	1	10.7	ALFAM2
UK	7.2	20.3	17.5	130.6	1	10.7	ALFAM2
UK	8.0	20.2	16.8	89.4	1	10.6	ALFAM2
DE	5.8	31.0	15.9	192.0	1	10.6	ALFAM2
DE	5.8	23.0	9.7	161.4	1	10.6	ALFAM2
DE	5.8	23.0	9.7	184.5	1	10.5	ALFAM2
CA	6.5	23.0	12.5	150.0	1	10.5	ALFAM2
UK	7.1	32.5	7.7	115.6	1	10.4	ALFAM2
DE	5.8	23.0	9.7	184.5	1	10.4	ALFAM2
DE	5.8	10.0	12.1	124.6	1	10.3	ALFAM2
DE	5.8	23.0	19.5	174.3	1	10.3	ALFAM2
UK	7.2	26.5	16.1	102.0	1	10.3	ALFAM2
UK	7.2	20.4	21.6	130.3	1	10.3	ALFAM2
DE	6.5	17.0	20.8	149.5	1	10.2	ALFAM2
DE	5.8	23.0	19.9	109.6	1	10.2	ALFAM2
DK	6.1	24.9	10.7	116.0	1	10.2	ALFAM2
CA	6.5	21.0	23.0	140.4	1	10.2	ALFAM2
DE	5.8	10.0	17.5	193.0	1	10.1	ALFAM2
DK	7.9	100.0	10.5	135.9	1	10.1	ALFAM2
UK	7.1	26.6	12.5	130.7	1	10.1	ALFAM2
GE	6.5	40.0	15.0	140.0	1	10.1	Ni et al., 2015
GE	6.5	40.0	8.0	140.0	1	10.1	Ni et al., 2015
IT	8.1	60.0	15.5	80.0	0	10.0	Badagliacca et al., 2018
GE	6.5	40.0	15.0	140.0	1	10.0	Ni et al., 2015
DK	6.1	26.9	9.9	110.9	1	9.9	ALFAM2
DE	5.8	10.0	12.1	54.8	1	9.9	ALFAM2
DE	5.8	10.0	12.1	124.6	1	9.9	ALFAM2
UK	7.2	20.3	17.5	130.6	1	9.8	ALFAM2
CA	6.5	24.0	16.4	151.7	1	9.8	ALFAM2
UK	8.0	28.5	10.0	149.1	1	9.7	ALFAM2
DE	5.8	23.0	9.7	161.4	1	9.6	ALFAM2
UK	8.0	20.2	16.9	164.0	1	9.5	ALFAM2

Table 2S. (continued).

DE	5.8	10.0	12.1	54.8	1	9.5	ALFAM2
UK	8.0	25.1	17.4	92.3	1	9.5	ALFAM2
UK	7.1	26.6	12.5	130.7	1	9.5	ALFAM2
DE	5.8	10.0	12.1	127.8	1	9.4	ALFAM2
DE	5.8	23.0	19.9	109.6	1	9.4	ALFAM2
DE	6.5	28.0	2.2	124.2	1	9.4	ALFAM2
UK	7.2	20.4	21.6	130.3	1	9.4	ALFAM2
UK	8.0	25.2	12.0	136.6	1	9.4	ALFAM2
DE	5.8	23.0	19.5	174.3	1	9.4	ALFAM2
UK	8.0	20.2	16.8	89.4	1	9.3	ALFAM2
DE	5.8	23.0	19.5	174.3	1	9.3	ALFAM2
DE	5.8	10.0	12.3	157.4	1	9.3	ALFAM2
DK	8.2	100.0	16.7	108.9	1	9.3	ALFAM2
DK	6.1	23.0	10.7	157.6	1	9.3	ALFAM2
DE	6.5	28.0	2.2	124.2	1	9.2	ALFAM2
CA	6.5	23.0	12.5	140.4	1	9.2	ALFAM2
SW	7.0	30.0	20.0	70.2	1	9.2	Hani et al., 2016
DE	6.5	28.0	2.2	124.3	1	9.1	ALFAM2
UK	7.1	28.8	12.1	110.1	1	9.1	ALFAM2
DE	5.8	23.0	9.8	161.4	1	9.1	ALFAM2
SW	6.8	30.0	20.0	69.6	1	9.0	Hani et al., 2016
DK	8.2	56.6	5.1	115.1	1	9.0	ALFAM2
DK	5.6	41.0	14.0	160.4	1	9.0	ALFAM2
DE	6.5	30.0	5.1	111.2	1	9.0	ALFAM2
UK	8.0	20.2	16.9	164.0	1	8.9	ALFAM2
DE	5.8	23.0	19.8	109.6	1	8.9	ALFAM2
DE	6.1	40.0	2.7	120.0	1	8.9	Gericke et al., 2011
DK	6.1	26.9	9.9	110.9	1	8.9	ALFAM2
DE	6.5	17.0	20.8	115.3	1	8.8	ALFAM2
UK	8.0	29.6	6.2	66.3	1	8.8	ALFAM2
DE	6.1	40.0	2.7	120.0	1	8.8	Gericke et al., 2011
UK	8.0	20.2	16.8	89.4	1	8.7	ALFAM2
DE	5.8	10.0	12.1	127.8	1	8.7	ALFAM2
CL	6.1	50.0	14.3	100.0	1	8.7	Salazar et al., 2014
DE	5.8	31.0	5.8	167.4	1	8.6	ALFAM2
UK	7.2	20.4	21.6	130.3	1	8.6	ALFAM2
DE	5.8	23.0	8.8	188.4	1	8.6	ALFAM2
DE	6.5	30.0	5.1	111.2	1	8.6	ALFAM2
UK	7.1	28.3	16.1	102.0	1	8.6	ALFAM2
GE	6.5	40.0	8.0	140.0	1	8.5	Ni et al., 2015

Table 2S. (continued).

UK	8.0	28.5	9.8	126.1	1	8.5	ALFAM2
SW	7.1	30.0	15.0	44.0	1	8.5	Hani et al., 2016
DE	5.8	10.0	12.2	157.4	1	8.4	ALFAM2
CA	6.5	21.0	23.0	143.4	1	8.4	ALFAM2
DK	6.1	23.0	10.7	157.6	1	8.4	ALFAM2
DE	6.5	17.0	20.4	95.5	1	8.4	ALFAM2
DE	5.8	23.0	19.9	73.0	1	8.4	ALFAM2
DK	6.1	28.3	9.9	121.8	1	8.4	ALFAM2
SW	7.1	30.0	16.0	54.0	1	8.3	Hani et al., 2016
UK	7.1	26.6	12.5	130.7	1	8.3	ALFAM2
DE	5.8	10.0	12.1	54.8	1	8.2	ALFAM2
DK	6.1	24.2	10.7	116.3	1	8.2	ALFAM2
DE	5.8	28.0	14.6	177.5	1	8.1	ALFAM2
CA	6.1	30.0	19.2	140.1	1	8.1	ALFAM2
SW	6.9	30.0	19.0	59.8	1	8.1	Hani et al., 2016
DE	6.5	17.0	20.8	115.3	1	8.1	ALFAM2
UK	8.0	25.2	12.0	87.3	1	8.0	ALFAM2
DK	7.0	10.8	12.8	76.8	1	8.0	ALFAM2
DK	6.1	24.2	10.7	116.3	1	8.0	ALFAM2
UK	7.2	30.6	10.5	110.1	1	7.9	ALFAM2
GE	6.5	40.0	17.0	110.0	0	7.8	Ni et al., 2015
DE	6.5	17.0	20.8	191.8	1	7.8	ALFAM2
DE	5.8	31.0	15.3	178.3	1	7.8	ALFAM2
CA	6.1	30.0	19.2	140.5	1	7.8	ALFAM2
SW	7.3	30.0	14.0	64.0	1	7.7	Hani et al., 2016
CL	6.4	42.6	10.0	100.0	0	7.7	Salazar et al., 2012
DE	5.8	10.0	12.1	73.0	1	7.7	ALFAM2
DE	6.5	30.0	5.1	111.2	1	7.7	ALFAM2
DE	5.8	23.0	19.9	91.3	1	7.7	ALFAM2
DE	5.8	23.0	19.9	91.3	1	7.7	ALFAM2
GE	6.5	40.0	8.0	140.0	1	7.7	Ni et al., 2015
DE	5.8	10.0	12.1	124.6	1	7.6	ALFAM2
DE	5.8	10.0	16.9	52.9	1	7.6	ALFAM2
DE	5.8	31.0	5.3	170.7	1	7.6	ALFAM2
DE	6.1	40.0	5.3	120.0	1	7.6	Gericke et al., 2011
CL	5.3	50.0	9.4	100.0	1	7.6	Martinez-Lagos et al., 2013
DE	6.5	28.0	2.2	124.2	1	7.6	ALFAM2
DE	5.8	10.0	12.3	74.7	1	7.6	ALFAM2
DE	6.5	30.0	4.4	130.0	1	7.6	ALFAM2

Table 2S. (continued).

DE	5.8	23.0	8.9	188.4	1	7.6	ALFAM2
UK	7.1	28.8	12.1	110.1	1	7.5	ALFAM2
DE	6.1	40.0	18.5	80.0	1	7.5	Gericke et al., 2011
DK	7.0	12.0	12.8	83.6	1	7.5	ALFAM2
DK	7.8	100.0	0.3	109.1	1	7.5	ALFAM2
DE	6.5	30.0	4.6	89.0	1	7.4	ALFAM2
DE	5.8	10.0	12.3	74.7	1	7.4	ALFAM2
SW	7.3	30.0	2.0	67.5	1	7.3	Hani et al., 2016
DE	6.5	30.0	5.0	111.2	1	7.3	ALFAM2
GE	6.1	40.0	5.3	120.0	1	7.3	Gericke et al., 2011
CL	6.1	50.0	10.6	100.0	1	7.3	Salazar et al., 2014
DE	6.5	30.0	4.7	89.0	1	7.3	ALFAM2
DE	5.8	23.0	19.8	73.0	1	7.3	ALFAM2
GE	6.5	40.0	17.0	110.0	0	7.3	Ni et al., 2015
DE	6.5	30.0	4.5	130.0	1	7.2	ALFAM2
DE	5.8	10.0	12.1	73.0	1	7.2	ALFAM2
DK	5.6	41.0	14.0	160.4	1	7.2	ALFAM2
DE	6.1	40.0	18.5	80.0	1	7.1	Gericke et al., 2011
CA	6.5	24.0	16.4	145.2	1	7.0	ALFAM2
DE	5.8	31.0	5.8	167.4	1	7.0	ALFAM2
UK	8.0	28.5	9.8	149.1	1	7.0	ALFAM2
SW	7.3	30.0	21.0	55.1	1	7.0	Hani et al., 2016
DK	8.0	100.0	17.3	112.7	1	6.9	ALFAM2
UK	8.0	28.5	9.8	149.1	1	6.9	ALFAM2
IE	7.0	15.0	13.1	119.6	1	6.9	Meade et al., 2011
UK	8.0	29.6	6.2	66.3	1	6.9	ALFAM2
DE	6.5	17.0	20.6	95.5	1	6.8	ALFAM2
DK	7.4	100.0	10.2	106.5	1	6.8	ALFAM2
DE	5.8	31.0	5.3	170.7	1	6.8	ALFAM2
DK	6.1	27.4	9.9	118.4	1	6.8	ALFAM2
DE	5.8	23.0	19.9	36.5	1	6.8	ALFAM2
CA	6.1	30.0	19.2	139.2	1	6.8	ALFAM2
DK	5.5	47.0	6.7	204.8	1	6.8	ALFAM2
GE	6.5	40.0	17.0	110.0	0	6.7	Ni et al., 2015
DK	6.1	27.8	9.9	111.4	1	6.7	ALFAM2
DE	5.8	28.0	14.6	165.4	1	6.5	ALFAM2
DK	6.1	22.8	10.7	109.2	1	6.5	ALFAM2

Table 2S. (continued).

DE	6.5	28.0	1.6	116.7	1	6.5	ALFAM2
DK	6.1	29.5	9.9	115.9	1	6.5	ALFAM2
CA	6.5	24.0	16.4	97.4	1	6.3	ALFAM2
DK	5.6	41.0	14.0	160.4	1	6.3	ALFAM2
DE	5.8	23.0	19.8	73.0	1	6.3	ALFAM2
UK	7.2	30.6	10.5	110.1	1	6.2	ALFAM2
SW	7.7	30.0	2.0	49.4	1	6.2	Hani et al., 2016
DE	6.5	17.0	20.5	95.5	1	6.2	ALFAM2
CA	6.1	30.0	19.2	139.9	1	6.2	ALFAM2
DE	6.5	30.0	4.5	130.0	1	6.2	ALFAM2
DK	6.1	27.4	9.9	118.4	1	6.2	ALFAM2
DK	6.1	24.5	10.7	118.3	1	6.1	ALFAM2
DE	5.8	28.0	14.6	165.4	1	6.1	ALFAM2
DE	5.8	31.0	15.3	178.3	1	6.1	ALFAM2
DE	6.5	30.0	4.5	130.0	1	6.1	ALFAM2
IE	7.0	15.0	11.7	103.2	1	6.0	Meade et al., 2011
CL	5.5	50.0	8.7	100.0	1	6.0	Martinez-Lagos et al., 2013
DE	6.5	30.0	4.8	89.0	1	6.0	ALFAM2
UK	7.2	26.5	16.1	102.0	1	5.9	ALFAM2
IT	5.5	25.0	12.2	92.0	1	5.9	Carozzi et al., 2013
SW	7.7	30.0	21.0	35.1	1	5.9	Hani et al., 2016
GE	6.5	40.0	17.0	110.0	0	5.8	Ni et al., 2015
CN	8.2	23.0	13.0	122.0	0	5.8	Huo et al., 2015
SW	7.3	30.0	23.0	46.4	1	5.8	Hani et al., 2016
DE	5.8	23.0	19.7	99.0	1	5.7	ALFAM2
DK	6.1	27.8	9.9	111.4	1	5.7	ALFAM2
SW	6.8	30.0	16.0	69.6	1	5.7	Hani et al., 2016
DE	5.8	10.0	12.2	74.7	1	5.6	ALFAM2
DE	6.5	40.0	15.0	140.0	1	5.6	Ni et al., 2015
DE	5.8	23.0	8.9	188.4	1	5.5	ALFAM2
IT	5.9	15.8	7.8	204.1	1	5.5	ALFAM2
DE	5.8	23.0	19.9	73.0	1	5.4	ALFAM2
DE	5.8	23.0	19.8	36.5	1	5.4	ALFAM2
DE	6.5	17.0	20.8	115.3	1	5.3	ALFAM2
SW	7.2	30.0	20.0	70.2	1	5.3	Hani et al., 2016
DE	5.8	23.0	19.7	99.0	1	5.3	ALFAM2
SW	7.3	30.0	13.0	56.0	1	5.2	Hani et al., 2016
SW	7.2	30.0	9.0	55.1	1	5.2	Hani et al., 2016
DK	6.1	24.5	10.7	118.3	1	5.2	ALFAM2

Table 2S. (continued).

DE	5.8	28.0	14.6	177.5	1	5.1	ALFAM2
DE	5.8	10.0	12.1	73.0	1	5.0	ALFAM2
DE	5.8	31.0	5.8	167.4	1	5.0	ALFAM2
DE	6.5	21.0	13.3	121.3	1	4.9	ALFAM2
DE	5.8	28.0	14.6	177.5	1	4.9	ALFAM2
DK	6.1	29.5	9.9	115.9	1	4.9	ALFAM2
DK	7.0	12.9	12.8	82.9	1	4.9	ALFAM2
SW	7.1	30.0	16.0	64.0	1	4.8	Hani et al., 2016
DE	6.1	40.0	16.7	120.0	1	4.8	Gericke et al., 2011
SW	7.3	30.0	17.0	49.6	1	4.7	Hani et al., 2016
CA	6.5	24.0	16.4	97.4	1	4.7	ALFAM2
SW	7.3	30.0	13.0	56.0	1	4.6	Hani et al., 2016
DK	6.4	14.7	11.9	94.4	1	4.6	ALFAM2
IT	8.0	14.9	10.2	103.9	1	4.5	ALFAM2
UK	7.2	26.5	16.1	102.0	1	4.5	ALFAM2
DE	5.8	28.0	14.6	75.4	1	4.5	ALFAM2
DE	6.1	40.0	16.7	120.0	1	4.5	Gericke et al., 2011
DE	5.8	28.0	14.6	165.4	1	4.5	ALFAM2
DE	5.8	28.0	14.6	37.7	1	4.5	ALFAM2
UK	8.0	25.2	12.0	87.3	1	4.3	ALFAM2
DE	5.8	23.0	19.9	36.5	1	4.3	ALFAM2
DE	6.5	21.0	13.3	121.3	1	4.3	ALFAM2
DK	6.1	28.3	9.9	121.8	1	4.3	ALFAM2
SW	7.6	30.0	2.0	55.0	1	4.3	Hani et al., 2016
DE	6.5	28.0	1.7	116.7	1	4.2	ALFAM2
SW	7.3	30.0	9.0	48.6	1	4.2	Hani et al., 2016
SW	7.3	30.0	16.0	46.4	1	4.2	Hani et al., 2016
CN	7.7	18.5	10.6	200.0	0	4.1	Roelcke et al., 2002
DE	5.8	23.0	19.9	36.5	1	4.0	ALFAM2
DE	5.8	10.0	12.1	54.8	1	3.9	ALFAM2
DE	6.5	21.0	13.4	121.3	1	3.9	ALFAM2
SW	7.2	30.0	15.0	59.4	1	3.9	Hani et al., 2016
SW	7.7	30.0	23.0	24.7	1	3.9	Hani et al., 2016
DE	6.5	28.0	2.0	94.8	1	3.9	ALFAM2
DE	5.8	23.0	19.8	99.0	1	3.8	ALFAM2
IT	8.0	22.0	12.3	66.0	1	3.7	Carozzi et al., 2013
DE	5.8	28.0	14.6	111.9	1	3.6	ALFAM2
DE	6.5	28.0	2.0	94.7	1	3.6	ALFAM2
IT	7.5	25.5	19.6	191.8	1	3.5	ALFAM2

Table 2S. (continued).

DE	5.8	28.0	14.6	111.9	1	3.5	ALFAM2
CN	7.8	14.7	30.2	200.0	0	3.5	Roelcke et al., 2002
DE	5.8	10.0	16.9	52.9	1	3.4	ALFAM2
DE	6.5	28.0	2.0	94.7	1	3.4	ALFAM2
IT	7.5	36.0	19.2	139.0	1	3.4	Carozzi et al., 2013
SW	7.0	30.0	15.0	44.8	1	3.4	Hani et al., 2016
CN	8.3	20.0	13.4	180.0	0	3.3	Yang et al., 2015
SW	7.1	30.0	15.0	75.4	1	3.3	Hani et al., 2016
DE	5.8	28.0	14.6	177.5	1	3.3	ALFAM2
DE	5.8	28.0	14.6	75.4	1	3.2	ALFAM2
DK	7.0	9.0	12.8	65.5	1	3.2	ALFAM2
UK	7.1	28.8	12.1	110.1	1	3.2	ALFAM2
DE	6.5	30.0	4.7	86.6	1	3.2	ALFAM2
DE	6.5	30.0	4.6	59.3	1	3.1	ALFAM2
CL	6.1	50.0	8.5	100.0	1	3.1	Salazar et al., 2014
DE	5.8	31.0	5.6	170.7	1	3.0	ALFAM2
UK	7.1	28.3	16.1	102.0	1	2.9	ALFAM2
DE	5.8	28.0	14.6	37.7	1	2.8	ALFAM2
DE	5.8	28.0	14.6	81.1	1	2.8	ALFAM2
SW	7.7	30.0	7.0	47.6	1	2.7	Hani et al., 2016
DE	6.5	30.0	4.6	43.3	1	2.7	ALFAM2
DE	6.5	28.0	2.0	94.8	1	2.7	ALFAM2
DE	5.8	28.0	14.6	81.1	1	2.7	ALFAM2
CN	8.3	16.0	22.5	180.0	0	2.7	Yang et al., 2015
DE	5.8	28.0	14.6	37.7	1	2.5	ALFAM2
SW	7.7	30.0	23.0	29.9	1	2.4	Hani et al., 2016
DE	5.8	28.0	14.6	111.9	1	2.4	ALFAM2
SW	7.7	30.0	22.0	36.4	1	2.4	Hani et al., 2016
DE	5.8	28.0	14.6	81.1	1	2.3	ALFAM2
GE	6.5	40.0	8.0	140.0	1	2.3	Ni et al., 2015
DE	6.5	21.0	13.4	119.2	1	2.3	ALFAM2
CN	7.8	14.7	30.2	100.0	0	2.3	Roelcke et al., 2002
SW	6.8	30.0	10.0	33.6	1	2.2	Hani et al., 2016
UK	8.0	25.2	12.0	87.3	1	2.1	ALFAM2
DE	5.8	28.0	14.6	75.4	1	2.1	ALFAM2
DE	5.8	28.0	14.6	37.7	1	2.0	ALFAM2
DE	6.5	30.0	4.7	29.7	1	2.0	ALFAM2
DE	5.8	28.0	14.6	111.9	1	2.0	ALFAM2
DE	6.5	21.0	13.4	119.2	1	1.9	ALFAM2

Table 2S. (continued).

SW	7.8	30.0	7.0	28.6	1	1.8	Hani et al., 2016
CL	6.1	50.0	8.5	100.0	0	1.8	Salazar et al., 2014
DE	6.5	21.0	13.4	119.2	1	1.8	ALFAM2
SW	7.2	30.0	14.0	20.9	1	1.7	Hani et al., 2016
SW	7.7	30.0	20.0	32.2	1	1.6	Hani et al., 2016
SW	7.7	30.0	20.0	24.7	1	1.6	Hani et al., 2016
UK	8.0	25.2	12.0	136.6	1	1.5	ALFAM2
SW	6.9	30.0	10.0	26.6	1	1.5	Hani et al., 2016
SW	7.2	30.0	14.0	27.0	1	1.5	Hani et al., 2016
CL	6.4	42.6	9.2	100.0	0	1.4	Salazar et al., 2012
DE	6.5	21.0	13.4	121.3	1	1.4	ALFAM2
SW	7.7	30.0	7.0	54.4	1	1.3	Hani et al., 2016
SW	7.8	30.0	24.0	31.2	1	1.2	Hani et al., 2016
CN	7.8	18.5	10.6	100.0	0	1.1	Roelcke et al., 2002
SW	6.9	30.0	20.0	33.6	1	1.0	Hani et al., 2016
SW	6.9	30.0	21.0	32.4	1	0.8	Hani et al., 2016
SW	7.3	30.0	14.0	22.0	1	0.7	Hani et al., 2016
SW	6.8	30.0	10.0	32.5	1	0.7	Hani et al., 2016
GE	6.6	40.0	9.1	180.0	0	0.6	Wolf et al., 2014

CHAPTER 5 Characteristic of NO_x Emission

5.1. Global

The NO₂ emission from agricultural soil is negligible, so this chapter only focus on the emission of NO. Similar to previous procedures, after calculating the coefficients of all variables, we applied the NO_STAT model for each grid cell (5 arc-minute x 5 arc-minute) and generated a global map for NO emissions. Figure 5.1(a) gives the spatial distribution of global NO emission from agricultural soils calculated using NO_STAT in kg N/yr/grid cell. The resolution of this map is 5 arc-minute, which is equivalent to 8464 ha. Figure 5.1(b) is spatial distribution of global NO emission from agricultural soil based on EDGAR. Total annual global NO emissions from NO_STAT and EDGAR are 0.208 Tg N/yr and 1.623 Tg N/yr, respectively. The NO_STAT model gives a lower global NO_x estimates by 87%. Similar to NH₃, there is a large difference between our estimate and EDGAR. As discussed previously, this difference can be attributed to other sources that are excluded in our model. EDGAR has additional sources in their estimate, e.g. Enteric fermentation, Manure management, Rice cultivation, whereas NO_STAT is exclusive to emissions from fertilizer and manure applied as fertilizer. Nevertheless, it is encouraging to see that, in general, the model captures the global spatial pattern in NO emissions well. The hot spots show most NO emission comes from northern China, northern India, and Middle Eastern U.S.

Our model also underestimated NO emission compare with literature survey. Based on our literature survey, Hudman et al. (2012) used a mechanistic model of global soil NO emissions to estimate that total global NO emission from soil is 10.7 Tg N yr⁻¹ and from

fertilizer N input (1.5% of applied N) is 1.8Tg N yr⁻¹. This might be due to that most of the experiments conducted on fields are short-term and non-continuous.

Figure 5.1 (c) shows the absolute difference between NO₂ and EDGAR. In comparison with EDGAR, NO₂ gives relatively lower NH₃ emission values in all the places.

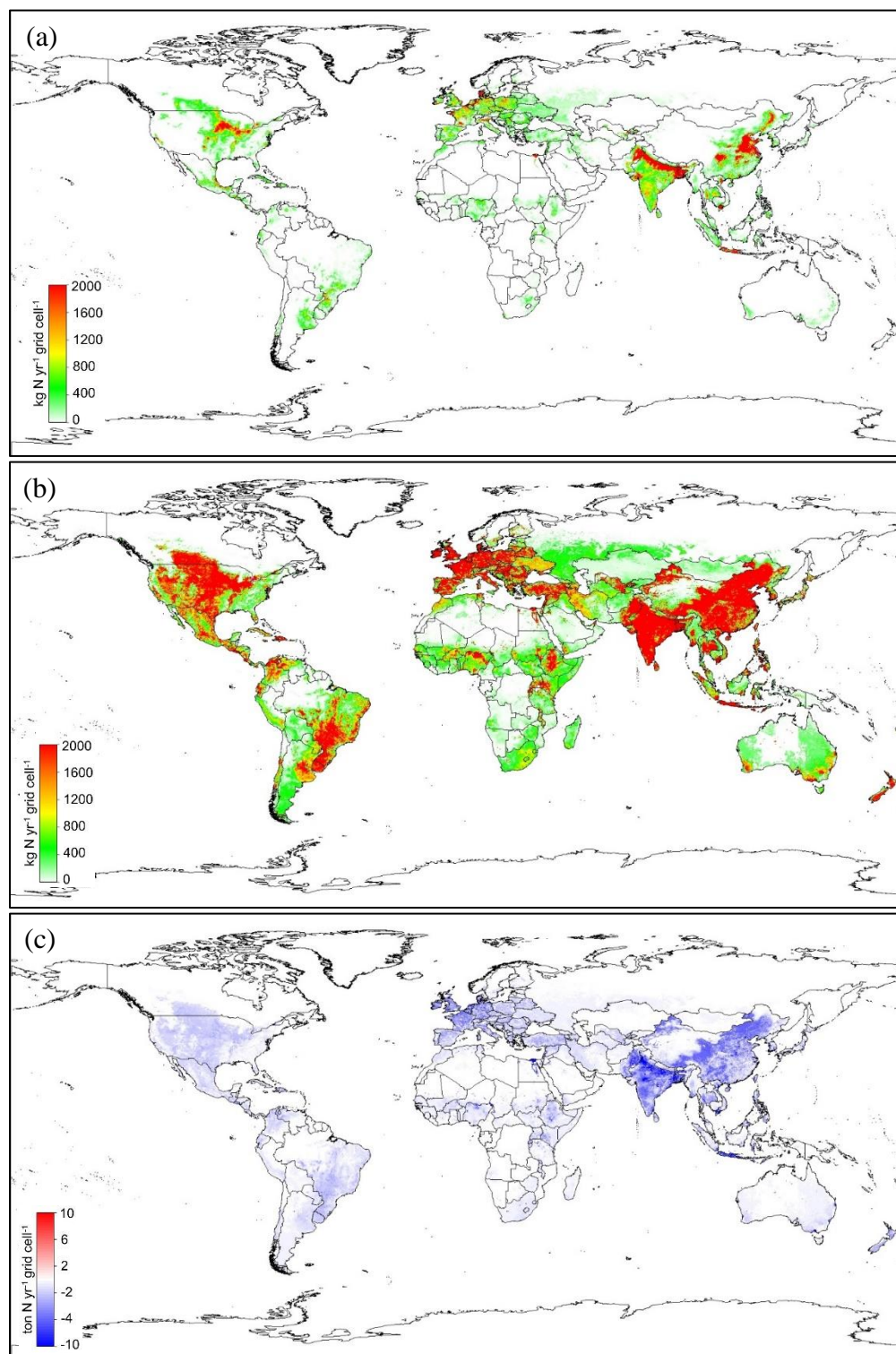


Figure 5.1 Comparison between the results from (a) NO_STAT, (b) EDGAR, and (c) absolute difference between the two models (in $\text{ton N yr}^{-1} \text{ grid cell}^{-1}$).

5.2. Conclusions

Overall, it is encouraging to see that the statistical model captures the spatial distribution of global NO_x emissions but the model estimates is far below other model estimate and literature survey. From global scale, the results indicate that, in comparison to other data sets, the model generates a lower global NO estimate by 87%, (NO_STAT: 0.208 Tg N/yr and EDGAR:1.623 Tg N/yr.). Two reasons can contribute to this underestimation. One reason is that the underestimation of NO emissions in comparison to EDGAR can be attributed to EDGAR has additional sources in their estimate, e.g. Enteric fermentation, Manure management, Rice cultivation, whereas NO_STAT is exclusive to emissions from fertilizer and manure applied as fertilizer. The other reason is that most NO field measurement campaign are short-term and non-continuous.

Supplementary Information

Table 3S. List of publications from where NO emissions and physicochemical parameters were obtained.

Location	pH	Soil moisture (%)	Temperature	N_type (0: fer 1:man)	N_rate (kg N/ha)	NO emission	Length of Experiment	Reference
Japan	5.9	39	23	0	200	1.5166656	131	Akiyama et al (2000)
Japan	5.9	39	23	0	200	2.3089536	131	Akiyama et al (2000)
Japan	5.9	39	23	0	200	2.376864	131	Akiyama et al (2000)
Colorado, USA	7.5	14.3	28	0	63	0.1714176	64	Anderson et al (1987)
Virginia, USA	6	25	15	0	40	1.33056	350	Anderson et al (1987)
Virginia, USA	6	25	15	0	40	1.48176	350	Anderson et al (1987)
Virginia, USA	6	25	15	0	100	1.897344	360	Anderson et al (1987)
Virginia, USA	6	25	15	0	100	4.292352	360	Anderson et al (1987)
NC, USA	6.2	28.33	14.9	0	68	0.536	365	Aneja_et_al._(1996)
NC, USA	6	35.75	13.7	0	168	2.208	365	Aneja_et_al._(1996)
Puerto Rico	4	87	22.6	0	150	0.637632	90	
Australia	8.2	22	20.3	0	80	0.002	8	Galbally_et_al._(1987)
France	5.2	30	12.5	0	280	32	365	Jambert_et_al._(1997)
France	4.8	30	6.5	0	280	0.001	1	Jambert_et_al._(1994)
France	4.5	30	6.5	0	150	0.002	1	Jambert_et_al._(1994)
France	6.6	30	18.5	0	150	0.473	1	Jambert_et_al._(1994)
Sweden	6.3	30	15.4	0	120	0.85	120	
Sweden	6.3	30	13.65	0	200	0.89	120	
Maui	6.8	40.5	20.3	0	34	0.004	8	Matson_et_al._(1996)
Maui	7.6	68.3	17.8	0	45	0.012	8	Matson_et_al._(1996)
Maui	4.7	49.5	19.7	0	84	0.012	8	Matson_et_al._(1996)
Maui	7	50.4	20.3	0	35	0.001	8	Matson_et_al._(1996)
Maui	6	68.3	17.8	0	22	0.008	8	Matson_et_al._(1996)

Table 3S. (continued).

Mauna Loa, Hawaii	5.4	68.8	5.6	0	94	0.71	30	Matson_et_al._(1996)
Maui	7	40.5	20.3	0	39	0.013	8	Matson_et_al._(1996)
Mauna Loa, Hawaii	4.3	57.7	5.8	0	95	1.95	15	Matson_et_al._(1996)
Brazil	5.84	57.75	32.25	0	33	1.58112	183	
Brazil	5.94	39.5	32.75	0	42	2.134512	183	
NC, USA	5.8	22.7	14.3	0	150	0.231	4	Roelle_et_al._(2001)
NC, USA	5.8	22.7	14.3	0	150	0.231	4	Roelle_et_al._(2001)
NC, USA	5.8	6.9	15.4	0	140	0.009	4	Roelle_et_al._(2001)
NC, USA	5.8	9.4	17.1	0	150	0.033	4	Roelle_et_al._(2001)
NC, USA	5.8	11.3	23	0	45	0.029	4	Roelle_et_al._(2001)
NC, USA	5.8	12.4	25.6	0	190	0.033	6	Roelle_et_al._(2001)
NC, USA	5.8	5.6	27.4	0	70	0.025	7	Roelle_et_al._(2001)
NC, USA	5.8	21.7	19.1	0	197	0.339	7	Roelle_et_al._(2001)
NC, USA	5.8	21.1	24.1	0	175	1.055	24	Roelle_et_al._(2001)
Venezuela	5.6	30	31	0	200	0.3	8	Rondon_et_al._(1993)
Venezuela	5.6	30	31	0	200	0.69	8	Rondon_et_al._(1993)
Venezuela	5.6	30	31	0	200	1.69	8	Rondon_et_al._(1993)
Guárice state	3.7	2.7	30	0	600	1.349	19	Sanhueza_et_al._(1994)
Venezuela	4.6	20	27	0	200	0.006	5	Sanhueza_et_al._(1990)
Venezuela	4.6	20	27	0	200	0.258	5	Sanhueza_et_al._(1990)
NC, USA	5.7	17.5	25.6	0	21	0.02	13	Aneja_et_al._(1995)
NC, USA	6.2	13.15	27.5	0	84	0.046	13	Aneja_et_al._(1995)
NC, USA	6	5.5	35	0	173	0.097	13	Aneja_et_al._(1995)
Tennessee, USA	5.8	25	11.5	1	100	0.04752	20	
Tennessee, USA	5.5	25	27.3	0	111	0.327	9	Valente_and_Thornton_ (1993)
Tennessee, USA	5.7	25	26.2	0	111	0.311	15	Valente_and_Thornton_ (1993)

Table 3S. (continued).

Costa Rica	5.1	70	25.8	0	360	10.7	365	Veldkamp_and_Keller_(1997)
Costa Rica	5.1	72.9	25.8	0	300	4.643	365	Veldkamp_et_al._(1998)
Costa Rica	5.1	72.9	25.8	0	300	5.413	365	Veldkamp_et_al._(1998)
Costa Rica	5.1	72.9	25.8	0	300	6.973	365	Veldkamp_et_al._(1998)
Costa Rica	5.1	72.9	25.8	0	300	7.483	365	Veldkamp_et_al._(1998)
Costa Rica	5.1	72.9	25.8	0	300	8.383	365	Veldkamp_et_al._(1998)
NC, USA	6	5.6	27.3	0	70	0.025	7	Aneja_et_al._(1998)
NC, USA	6	12.6	24.8	0	190	0.029	6	Aneja_et_al._(1998)
NC, USA	6	11.3	22.6	0	197	0.066	9	Aneja_et_al._(1998)
Colorado, USA	5.9	4.8	27.2	1	100	0.14688	17	Williams et al (1991)
England	7	22	12	0	200	0.79	365	Yamulki et al (1995)
England	7	22	10	0	150	0.64	365	Yamulki et al (1995)
Jiangsu, China	6.5	41	9	1	66	0.53	240	Yamulki et al (1995)
Jiangsu, China	6.5	41	9	0	66	0.485	240	Zheng et al (2003)
Jiangsu, China	6.5	34	5	0	29	0.03	240	Zheng et al (2003)
Jiangsu, China	6.5	34	17	0	96	4.22	240	Zheng et al (2003)

CHAPTER 6 Conclusions

6.1. Conclusions

Nr includes N₂O, NH₃, and NO_x. They are biologically active, chemically reactive, and radiatively active in the Earth's atmosphere and biosphere, which are different from non-reactive nitrogen gas. In this thesis work, three statistical model (N₂O_STAT, NH₃_STAT, and NO_STAT) are developed for characterizing atmospheric Nr emissions from agricultural soils. The results indicate that, in comparison to other data sets, the model generates a lower global N₂O estimate by 9-20% (N₂O_STAT: 3.75 Tg N yr⁻¹; EDGAR: 4.49 Tg N yr⁻¹; FAO: 4.07 Tg N yr⁻¹), a lower global NH₃ estimates by 57%, (NH₃_STAT: 14.2 Tg N yr⁻¹; EDGAR: 33.0 Tg N yr⁻¹), and a lower global NO_x estimates by 87%, (NO_STAT: 0.2 Tg N yr⁻¹; EDGAR: 1.6 Tg N yr⁻¹). We also performed a region-based analysis (U.S., India, and China) using the N₂O_STAT and NH₃_STAT models. Based on results, statistical models can capture the spatial distribution of global Nr emissions by utilizing a more simplified approach than those used by other readily available data sets, but all three models generated lower estimates compared to other inventories.

For N₂O_STAT model, we collected the measured N₂O emissions reported from previous studies that correspond to the use of fertilizer and manure on particular soil properties, which does not include emissions from leaching and surface runoff as such information is not often available. Therefore, our estimate is only driven by the direct N₂O emissions from agricultural soils. However, EDGAR and FAOSTAT incorporate both direct and indirect N₂O emissions in estimating the global emissions. Moreover, EDGAR includes rice cultivation and crop residue in its estimate (IPCC, 1996). Another key difference between N₂O_STAT and other

data sets lies in the methodology of collecting the model inputs. The N2O_STAT model uses the measured N2O emissions reported from field experiments. This is different from other data sets that derive their emissions by using intermediate data sets (e.g., fertilizer production, livestock counts) and apply measured emission factors to the corresponding data sets.

For NH₃_STAT, the model generated lower estimates compared to EDGAR, but the difference varies in difference areas. For the U.S., our model produces an estimate that is 143% higher in comparison to EPA, while 77% and 141% lower in India and 36-482% lower in China compared to other datasets. The difference in the global estimates is attributed to the lower estimates in major agricultural countries like China and India. Also, the underestimation of NH₃ emissions in comparison to EDGAR can be due to other sources that are excluded in our model.

For NO_STAT, statistical model captures the spatial distribution of global NO_x emissions but the model estimate is far below other model estimate and literature survey. Two reasons can contribute to this underestimation. One reason is that the underestimation of NO emissions in comparison to EDGAR can be attributed to EDGAR has additional sources in their estimate, e.g. Enteric fermentation, Manure management, Rice cultivation, whereas NO_STAT is exclusive to emissions from fertilizer and manure applied as fertilizer. The other reason is that most NO field measurement campaign are short-term and non-continuous.

6.2. Limitation and Future Work

Literature survey demonstrates that there is difficulty of using an emission factor approach to compute Nr emissions. In fact, this work provides an opportunity to estimate Nr emission factor from agricultural soils. Our model diagnostic tests indicated that Nr emissions

are a nonlinear function of N input. However, our model can be used to estimate a median value of Nr emission factor by calculating the ratio between Nr emission and applied N fertilizer in every grid cell of the map.

While statistical model provide an innovative and relatively simple way to estimate global Nr emission from agricultural sources, there are some limitations . These statistical models only considers physicochemical variables of the emissions, excluding the soil management practices that might contribute to the emissions. In addition, differences between animal types and fertilizer types, and crop types are not taken into account. The underestimation of Nr emissions in comparison to other data sets and the fact that statistical models may be still missing some key N inputs to its variables should be taken into account in the future work.

Another component worth investigating is soil biological activity that would more fully represent the processes governing the Nr emissions. Therefore, future work can be focused on including other important physicochemical variables of the emissions, e.g. nitrogen content, organic carbon content, taking into account the differences due to different measure method and process, and including other key N input.

Notwithstanding the above arguments, the N₂O_STAT, NH₃_STAT, and NO_STAT use measured values for Nr emissions including N content in the fertilizer to develop the model. In addition, the correlations between emissions and most physicochemical variables are at a high significance level (90%), suggesting that these variables are likely affecting the emissions despite excluding other variables. These models provide an opportunity to predict the impacts of climactic Nr emissions.

REFERENCES

- Adviento-Borbe, M. A. A., J. P. Kaye, M. A. Bruns, M. D. McDaniel, M. McCoy, and S. Harkcom (2010), Soil Greenhouse Gas and Ammonia Emissions in Long-Term Maize-Based Cropping Systems, *Soil Sci. Soc. Am. J.*, 74, 1623–1634, doi:10.2136/sssaj2009.0446.
- Akiyama, H., H. Tsuruta, and T. Watanabe (2000), N₂O and NO Emissions from Soils after the Application of Different Chemical Fertilizers. *Chemosphere Global Change Science*, 2, 313-320. [http://dx.doi.org/10.1016/S1465-9972\(00\)00010-6](http://dx.doi.org/10.1016/S1465-9972(00)00010-6).
- ALFAM2, <https://biotransformers.shinyapps.io/ALFAM2/>
- Anderson, R. C., and J. S. Levine (1987), Simultaneous field measurements of biogenic emissions of nitric oxide and nitrous oxide, *J. Geophys. Res.* 92(D1), 965-C976.
- Aneja, V.P., W.P. Robarge, B.D (1995). Holbrook Measurements of nitric oxide flux from an upper coastal plain, North Carolina agricultural soil *Atmospheric Environment*, 29, pp:3037-3042
- Aneja, V. P., W. P. Robarge, L. Sullivan, T. Moore, T Pierce, C D. Geron, AND B. Gay (1996). SEASONAL VARIATIONS OF NITRIC OXIDE FLUX FROM AGRICULTURAL SOILS IN THE SOUTHEAST UNITED STATES. *TELLUS* 48B:626-640.
- Aneja, V. P., Paul A. Roelle and Wayne P. Robarge (1998), Characterization of biogenic nitric oxide source strength in the southeast United States *Environmental Pollution* 102, S1211-218
- Aneja, V. P., W. H. Schlesinger, and J. W. Erisman, *Effects of Agriculture upon the Air Quality and Climate: Research, Policy, and Regulations* (2009), *Environ. Sci. Technol.*, 10.1021/es8024403
- Aneja, V. P., W. H. Schlesinger, J. W. Erisman, S. N. Behera, M. Sharma, and W. Battye (2012), Reactive Nitrogen Emissions from Crop and Livestock Farming in India, *Atmospheric Environment*, 47, 92-103.
- Asgedom, H., M. Tenuta, D. N. Flaten, X. Gao, and E. Kebreab (2014), Nitrous Oxide Emissions from a Clay Soil Receiving Granular Urea Formulations and Dairy Manure, *Agron. J.*, 106, 732–744, doi:10.2134/agronj2013.0096.

- Badagliacca, G., E. Benítez, G. Amato, L. Badalucco, D. Giambalvo, V. A. Laudicina, P. Ruisi, Long-term effects of contrasting tillage on soil organic carbon, nitrous oxide and ammonia emissions in a Mediterranean Vertisol under different crop sequences (2018), *Science of the Total Environment* 619–620, 18–27
- Baggs E. M. (2008), A review of stable isotope techniques for N₂O source partitioning in soils: Recent progress, remaining challenges and future considerations. *Rapid Communications in Mass Spectrometry*, 22, 1664-1672.
- Baruah, A. and Baruah, K. K. (2015), Organic Manures and Crop Residues as Fertilizer Substitutes: Impact on Nitrous Oxide Emission, Plant Growth and Grain Yield in Pre-Monsoon Rice Cropping System, *Journal of Environmental Protection*, 6, 755-770. <http://dx.doi.org/10.4236/jep.2015.67069>.
- Battye, W., V. P. Aneja, and P. A. Roelle, Evaluation and improvement of ammonia emissions inventories (2003), *Atmospheric Environment* 37 (2003) 3873–3883
- Battye, W., V. P. Aneja, and W. H. Schlesinger, Is nitrogen the next carbon? (2017), *Earth's Future*, Volume 5, Issue 9, 894-904, <https://doi.org/10.1002/2017EF000592>
- Behera, S. N. V. P. Aneja, R. Balasubramanian, Ammonia in the atmosphere: a review on emission sources, atmospheric chemistry and deposition on terrestrial bodies (2013), *Environmental Science and Pollution Research*, Volume 20, Issue 11, pp 8092–8131
- Bobbink, R., Hicks K, Galloway J, Spranger T, Alkemade R, Ashmore M, Bustamante M, Cinderby S, Davidson E, Dentener F, Emmett B, Erisman JW, Fenn M, Gilliam F, Nordin A, Pardo L, De Vries W., Global assessment of nitrogen deposition effects on terrestrial plant diversity: a synthesis (2010), *Ecol Appl.*, 20(1):30-59.
- Bouwman AF, Lee DS, Asman WAH, Dentener FJ, Van der Hoeck KW, Olivier JGJ (1997) A global high-resolution emission inventory for ammonia. *Global Biogeochem Cy* 11:561–587
- Bouwman, A. F., K. W. Van der Hoek, and J. G. J. Olivier (1995), Uncertainties in the global source distribution of nitrous oxide. *J. Geophys. Res.*, 100, 2785-2800.
- Bouwman, A. F., Pawłowski, M., Liu, C., Beusen, A. H. W., Shumway, S. E., Glibert, P. M. and Overbeek, C. C. 2011. Global hindcasts and future projections of coastal nitrogen and phosphorus loads due to shellfish and seaweed aquaculture. *Rev. Fisheries Sci.*, 19: 331–357.

- Bouwman, A.F., A.H.W. Beusen, J. Griffioen, J.W. Van Groenigen, M.M. Hefting, O. Oenema, P.J.T.M. Van Puijenbroek, S. Seitzinger, C.P. Slomp, E. Stehfest (2013), Uncertainties in the global source distribution of nitrous oxide, *Philosophical Transactions of the Royal Society B: Biological Sciences*, 268, 1621.
- Bouwman, A.F., Lee, D.S., Asman, W.A.H., Dentener, F.J., Van der Hock, K.W., Olivier, J.G.J., (1997). A global high-resolution emission inventory for ammonia. *Global Biogeochemical Cycles*, 11, 561-587.
- Butterbach-Bahl, K., Elizabeth M. B., Michael D., Ralf K., and Sophie Z-B., Nitrous oxide emissions from soils: how well do we understand the processes and their controls? (2013), *Philos Trans R Soc Lond B Biol Sci.*, 368(1621), 20130122., doi: 10.1098/rstb.2013.0122.
- Carozzi, M., R.M. Ferrara, G. Rana, M. Acutis, Evaluation of mitigation strategies to reduce ammonia losses from slurry fertilization on arable lands (2013), *Science of the Total Environment* 449,126–133
- Chantigny, M. H., D. E. Pelster, M.-H. Perron, P. Rochette, D. A. Angers, L.-É. Parent, D. Massé, and N. Ziadi (2012), Nitrous Oxide Emissions from Clayey Soils Amended with Paper Sludges and Biosolids of Separated Pig Slurry, *J. Environ. Qual.*, 42, 30–39.
- Chen, Z., W. Ding, Y. Luo, H. Yu, Y. Xu, and C. Müller X. Xu, and T. Zhu (2014), Nitrous oxide emissions from cultivated black soil: A case study in Northeast China and global estimates using empirical model, *Global Biogeochem. Cycles*, 27, 1311–1326, doi:10.1002/2014GB004871.
- Ciais, P., C. Sabine, G. Bala, L. Bopp, V. Brovkin, J. Canadell, A. Chhabra, R. DeFries, J. Galloway, M. Heimann, C. Jones, C. Le Quéré, R.B. Myneni, S. Piao and P. Thornton, (2013), Carbon and Other Biogeochemical Cycles. In: *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* [Stocker, T.F., D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bex and P.M. Midgley (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Collins, H. P., A. K. Alvaa, J. D. Streubelb, S. F. Fransenb, C. Frearc, S. Chenc, C. Krugerd and D. Granatsteind (2011), Greenhouse Gas Emissions from an Irrigated Silt Loam Soil Amended with Anaerobically Digested Dairy Manure, *Soil Sci. Soc. Am. J.*, 75, 2206–2216, doi:10.2136/sssaj2010.0360.

- Davidson, E. A. , The contribution of manure and fertilizer nitrogen to atmospheric nitrous oxide since 1860 (2009), *Nature Geoscience* volume 2, pages 659–662
- De Klein, C., Novoa, R.S.A., Ogle, S., Smith, K.A., Rochette, P., Wirth, T.C., McConkey, B.G., Mosier, A., Rypdal, K., Walsh, M., Williams, S.A., 2006. N₂O emissions from managed soils, and CO₂ emissions from lime and urea application (Chapter 11). In: 2006 IPCC Guidelines For National Greenhouse Gas Inventories.
- Ding, W., J. Luo, J. Li, H. Yu, J. Fan, and D. Liu (2013), Effect of long-term compost and inorganic fertilizer application on background N₂O and fertilizer-induced N₂O emissions from an intensively cultivated soil, *Science of the Total Environment*, 465, 115–124.
- Dittert, K., R. Bol, R. King, D. Chadwick, and D. Hatch (2001), Use of a novel nitrification inhibitor to reduce nitrous oxide emission from ¹⁵N-labelled dairy slurry injected into soil, *Rapid Communications in Mass Spectrometry*, 15(15), 1291-1296, doi: <https://doi.org/10.1002/rcm.335>
- EPA (2018), Inventory of U.S. Greenhouse Gas Emissions and Sinks, 1990-2016. U.S. Environmental Protection Agency, Washington D.C., pp. 655.
- Erickson, H., M. Keller, and E. A. Davidson (2001), Nitrogen oxide fluxes and nitrogen cycling during post-agricultural succession and forest fertilization in the humid tropics, *Ecosystems*, 4, 67-C84.
- Ferrara R.M., B. Loubet, C. Decuq, A.D. Palumbo, P. Di Tommasi, V. Magliulo, S. Masson, E. Personne, P. Cellier, G. Rana, Ammonia volatilisation following urea fertilisation in an irrigated sorghum crop in Italy (2014), *Agricultural and Forest Meteorology*, 195–196, 179–191
- Firestone, M. K. and Davidson, E. A. (1989). Microbial basis of NO and N₂O production and consumption in soil. Pages 7–22 in M. O. Andreae and D. S. Schimel, eds. Exchange of trace gases between terrestrial ecosystems and the atmosphere. John Wiley & Sons, Ltd., Toronto, ON.
- Forster, P., V. Ramaswamy, P. Artaxo, T. Berntsen, R. Betts, D.W. Fahey, J. Haywood, J. Lean, D.C. Lowe, G. Myhre, J. Nganga, R. Prinn, G. Raga, M. Schulz and R. Van Dorland (2007), Changes in Atmospheric Constituents and in Radiative Forcing. In: *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* [Solomon, S., D. Qin, M. Manning, Z. Chen, M. Marquis, K.B. Averyt, M. Tignor and H.L. Miller

- (eds.)). Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Fowler D, M. Coyle, U. Skiba, M. A. Sutton, J. N. Cape, S. Reis, L. J. Sheppard, A. Jenkins, B. Grizzetti, J. N. Galloway, P. Vitousek, A. Leach, A. F. Bouwman, K. Butterbach-Bahl, F. Confidential manuscript submitted to GEOPHYSICAL RESEARCH LETTERS
- Dentener, D. Stevenson, M. Amann, and M. Voss (429 2013), The global nitrogen cycle in the twenty-first century. *Phil Trans R Soc B* 368, 20130164, doi:10.1098/rstb.2013.0164
- Fowler, D., C. E. Steadman, D. Stevenson, M. Coyle, R. M. Rees, U. M. Skiba, M. A. Sutton, J. N. Cape, A. J. Dore, M. Vieno, D. Simpson, S. Zaehle, B. D. Stocker, M. Rinaldi, M. C. Facchini, C. R. Flechard, E. Nemitz, M. Twigg, J. W. Erisman, K. Butterbach-Bahl, and J. N. Galloway (2015), Effects of global change during the 21st century on the nitrogen cycle, *Atmos. Chem. Phys.*, 15, 13849–13893, doi:10.5194/acp-15-13849-2015
- Galbally et al., (1987) I.E. Galbally, J.R. Freney, W.A. Muirhead, J.R. Simpson, A.C.F. Trevitt, P.M. Chalk, Emission of nitrogen oxides (NO_x) from a flooded soil fertilized with urea: relation to other nitrogen loss processes, *J. atmos. Chem.*, 5, pp. 343-365
- Ganesan, A. L., A. Chatterjee, R. G. Prinn, C. M. Harth, P. K. Salameh, A. J. Manning, B. D. Hall, J. Mühle, L. K. Meredith, R. F. Weiss, S. O'Doherty, and D. Young (2013), The variability of methane, nitrous oxide and sulfur hexafluoride in Northeast India, *Atmos. Chem. Phys.*, 13, 10633–10644, doi:10.5194/acp-13-10633-2013
- Ganesan, A. L., A. J. Manning, A. Grant, D. Young, D. E. Oram, W. T. Sturges, J. B. Moncrieff, and S. O'Doherty (2015), Quantifying methane and nitrous oxide emissions from the UK and Ireland using a national-scale monitoring network, *Atmos. Chem. Phys.*, 15, 6393-6406, doi:10.5194/acp-15-6393-2015
- Gao, B.; Ju, X. T.; Zhang, Q.; Christie, P.; Zhang, F. S. New estimates of direct N₂O emissions from Chinese croplands from 1980 to 2007 using localized emission factors. *Biogeosciences* 2011, 8(10), 3011-3024.
- Garg, A., Shukla, P.R., Kapshe, M., 2006. The sectoral trends of multigas emissions inventory of India. *Atmospheric Environment*, 40, 4608-4620.
- Gericke, D., A. Pacholski, H. Kage, Measurement of ammonia emissions in multi-plot field experiments (2011), *biosystems engineering*, 108, 164-173
- Ghosh, S., D. Majumdar, and M. C. Jain (2003), Methane and nitrous oxide emissions from an irrigated rice of North India, *Chemosphere* 51 181-195.

- Goldberg SD, Gebauer G. (2009). N₂O and NO fluxes between a Norway spruce forest soil and atmosphere as affected by prolonged summer drought. *Soil Biol. Biochem.* 41, 1986–1995. DOI: 10.1016/j.soilbio.2009.07.001
- Granli, T. and Bockman, O. C. (1994), Nitrous oxide from agriculture. *Nor. J. Agric. Sci.* (Suppl. 12), pp. 128.
- Gronberg, J. M. and N. E. Spahr (2012), County-level estimates of nitrogen and phosphorus from commercial fertilizer for the conterminous United States, 1987-2006, U.S. Geological Survey Report.
- Gronberg, J. M. and T. L. Arnold (2017), County-level estimates of nitrogen and phosphorus from animal manure for the conterminous United States, 2007 and 2012, U.S. Geological Survey Report.
- Guo, Y., L. Luo, G. Chen, Y. Kou, and H. Xu (2013), Mitigating nitrous oxide emissions from a maize-cropping black soil in northeast China by a combination of reducing chemical N fertilizer application and applying manure in autumn, *Soil, Science and Plant Nutrition*, 59(3), 392-402, DOI: 10.1080/00380768.2013.775006.
- Hadi, A., O. Jumadi, K. Inubushi and K. Yagi (2008), Mitigation options for N₂O emission from a corn field in Kalimantan, Indonesia, *Soil Science and Plant Nutrition*, 54, 644–649 doi: 10.1111/j.1747-0765.2008.00280.x.
- Häni, C., J. Sintermann, T. Kuppera, M. Jocherb, A. Neftelb, Ammonia emission after slurry application to grassland in Switzerland (2016), *Atmospheric Environment*, 125, 92–99
- Hudman, R. C. and Moore, N. E. and Mebust, A. K. and Martin, R. V. and Russell, A. R. and Valin, L. C. and Cohen, R. C. (2012), Steps towards a mechanistic model of global soil nitric oxide emissions: implementation and space based-constraints, *Atmospheric Chemistry and Physics*, 12, 7779—779.
- Hutchinson, G. L. (1995), Biosphere-atmosphere exchange of gaseous N oxides in soul and global change, edited by R. Lal et al., chap 18, pp. 219-236, CRC Press, Boca Raton. Fla., 1995.
- Huo, Q., X. Cai, L. Kang, H. Zhang, Y. Song, T. Zhu, Estimating ammonia emissions from a winter wheat cropland in North China Plain with field experiments and inverse dispersion modeling (2015), *Atmospheric Environment*, 104, 1-10

- IPCC (1996), Revised 1996 IPCC guideline for national greenhouse gas inventories, Common reporting framework: Agriculture, IPCC, 1.11-1.12.
- IPCC (2006), Agriculture, forestry and other land uses. In: 2006 IPCC Guidelines for National Greenhouse Gas Inventories, Prepared by the National Greenhouse Gas Inventories Programme (eds Eggleston HS, Buendia L, Miwa K, Ngara T, Tanabe K), pp. 1.1–1.21. IGES, Hayama, Japan.
- Jambert C., R. A. Delmas, L. Labroue, and P. Chassin (1994), Nitrogen compound emissions from fertilized soils in a maize field-pine tree forest system in the Southwest of France, *J. Geophys. Res.*, 99: 16,523–16,530.
- Jambert C Serca D Delmas R . (1997). Quantification of N-losses as NH₃, NO, and N₂O and N₂ from fertilized maize fields in southwestern France. *Nutrient Cycling in Agroecosystems* . 48: 91-104.
- Janssens-Maenhout, G., Monica C., Diego G., Marilena M., Edwin S., Frank D., Peter B., Valerio P., Jos G. J. O., Jeroen A. H. W. P., John A. van A., Suvi M., Ulrike D., and A. M. Roxana P. (2017), EDGAR v4.3.2 Global Atlas of the three major Greenhouse Gas Emissions for the period 1970–2012, *Earth Syst. Sci. Data Discuss.*, <https://doi.org/10.5194/essd-2017-79>.
- Johansson, C., and L. Granat (1984), Emissions of nitric oxide from arable land, *Tellus*, 36B, 25-C37.
- Jumadi, O., Y. Hala, A. Muis, A. Ali, M. Palennari, K. Yagi, K. Inubushi(2008), Influences of Chemical Fertilizers and a Nitrification Inhibitor on Greenhouse Gas Fluxes in a Corn (*Zea mays* L.) Field in Indonesia, *Microbes Environ.*, 23(1), 29-34.
- Keller, M., and W. A. Reiners (1994), Soil-atmosphere exchange of nitrous oxide, nitric oxide, and methane under secondary succession of pasture to forest in the Atlantic lowlands of Costa Rica, *Global Biogeochem. Cycles*, 8(4), 399–409, doi:10.1029/94GB01660.
- Kelly, K. B., F. A. Phillips and R. Baigent (2008). Impact of dicyandiamide application on nitrous oxide emissions from urine patches in northern Victoria, Australia, *Australian Journal of Experimental Agriculture*, 48, 156-159.
- Kim, D-G., D. Giltrap, G. Hernandez-Ramirez (2013), Background nitrous oxide emissions in agricultural and natural lands: a meta-analysis, *Plant Soil*, 373, 17-30.

- Kumar, U., M.C. Jain, H. Pathak, S. Kumar, D. Majumdar (2000). Nitrous oxide emission from different fertilizers and its mitigation by nitrification inhibitors in irrigated rice, *Biol Fertil Soils*, 32, 474–478.
- Kwok, H. F., S. L. Napelenok, and K. R. Baker, Implementation and evaluation of PM 2.5 source contribution analysis in a photochemical model (2013), *Atmospheric Environment*. 80. 10.1016/j.atmosenv.2013.08.017.
- Lee, D.S., Kohler I., Grobler, E., Rohrer, F., Sausen, R., Gallardo-Klenner, L., Olivier, J.G.J., Dentener, F.J., Bouwman, A.F., (1997). Estimations of global NO_x emissions and their uncertainties, *Atmospheric Environment*, 31, 1735-1749.
- Li, L., X Han, M. You, and W. R. Horwath (2013), Nitrous oxide emissions from Mollisols as affected by long-term applications of organic amendments and chemical fertilizers, *Science of the Total Environment*, 452, 302–308.
- Linzmeier, W., R. Gutser, and U. Schmidhalter (2001), Nitrous oxide emission from soil and from a nitrogen-15-labelled fertilizer with the new nitrification inhibitor 3,4-dimethylpyrazole phosphate (DMPP), *Biol Fertil Soils*, 34, 103–108, doi: 10.1007/s003740100383
- Macadam, X. M. B., A. Del Prado, P. Merino, J. M. Estavillo, M. Pinto, and C. Gonza'lez-Murua (2003), Dicyandiamide and 3,4-dimethyl pyrazole phosphate decrease N₂O emissions from grassland but dicyandiamide produces deleterious effects in clover, *Journal of Plant Physiology*, 160, 1517-1523.
- Majumdar, D., H. Pathak, S. Kumar, and M. C. Jain (2002), Nitrous oxide emission from a sandy loam Inceptisol under irrigated wheat in India as influenced by different nitrification inhibitors, *Agriculture, Ecosystems and Environment*, 91, 283–293.
- Majumdar, D., S. Kumar, H. Pathak, M.C. Jain, and U. Kumar (2000), Reducing nitrous oxide emission from an irrigated rice field of North India with nitrification inhibitors, *Agriculture, Ecosystems and Environment*, 81, 163-169.
- Malla, G., A. Bhatia, H. Pathak, S. Prasad, N. Jain, and J. Singh (2005), Mitigating nitrous oxide and methane emissions from soil in rice–wheat system of the Indo-Gangetic plain with nitrification and urease inhibitors, *Chemosphere*, 58, 141–147.
- Matson, P. A., C. Billow, S. Hall, J. Zachariassen, (1996), Fertilization practices and soil variations control nitrogen oxide emissions from tropical sugar cane, Volume101, IssueD13, Pages 18533-18542

- Martínez-Lagos, J., F. Salazar, M. Alfaro, T. Misselbrook, Ammonia volatilization following dairy slurry application to a permanent grassland on a volcanic soil (2013), *Atmospheric Environment* 80, 226-231
- Meadea, G., K. Piercea, J.V. O'Dohertya, C. Muellerb, G. Laniganc, T. Mc Cabea, Ammonia and nitrous oxide emissions following land application of high and low nitrogen pig manures to winter wheat at three growth stages (2011), *Agriculture, Ecosystems and Environment*, 140, 208–217
- Menendez, S., P. Merino, M. Pinto, C. Gonzalez-Murua, and J. M. Estavillo (2006), 3,4-Dimethylpyrazol Phosphate Effect on Nitrous Oxide, Nitric Oxide, Ammonia, and Carbon Dioxide Emissions from Grasslands, *J. Environ. Qual.*, 35, 973– 981, doi:10.2134/jeq2005.0320.
- Merino, P., J.M. Estavillo, L. A. Graciolli, M. Pinto, M. Lacuesta, A. MunÄoz-Rueda, and C. Gonzalez-Murua (2002), Mitigation of N₂O emissions from grassland by nitrification inhibitor and Actilith F2 applied with fertilizer and cattle slurry, *Soil Use and Management*, 18, 135±141 DOI: 10.1079/SUM2002120.
- Merino, P., S. Mene´ndez, M. Pinto, C. Gonza´lez-Murua, and J.M. Estavillo (2005), 3, 4-Dimethylpyrazole phosphate reduces nitrous oxide emissions from grassland after slurry application, *Soil Use and Management*, 21, 53–57 DOI: 10.1079/SUM2005292.
- Misselbrook, T. H., L. M. Cardenas, V. Camp, R. E. Thorman, J. R. Williams, A. J. Rollett, and B. J. Chambers (2014), An assessment of nitrification inhibitors to reduce nitrous oxide emissions from UK agriculture, *Environ. Res. Lett.*, 9, 115006, doi: 10.1088/1748-9326/9/11/115006.
- Miyazaki K., H. J. Eskes, and K. Sudo (2012) Global NO_x emission estimates derived from an assimilation of OMI tropospheric NO₂ columns, *Atmos. Chem. Phys.*, 12, 2263-2288.
- Mosier, A., Kroeze, C., Nevison, C., Oenema, O., Seitzinger, S., Cleemput, O. van (1998), Closing the global N₂O budget: nitrous oxide emissions through the agricultural nitrogen cycle. *Nutr. Cycl. Agroecosyst.*, 52, 225–248. <https://doi.org/10.1023/A:1009740530221>.
- Myhre, G., D. Shindell, F.-M. Bréon, W. Collins, J. Fuglestedt, J. Huang, D. Koch, J.-F. Lamarque, D. Lee, B. Mendoza, T. Nakajima, A. Robock, G. Stephens, T. Takemura, and H. Zhang (2013), Anthropogenic and natural radiative forcing. In *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. T.F. Stocker, D. Qin, G.-K.

- Plattner, M. Tignor, S.K. Allen, J. Doschung, A. Nauels, Y. Xia, V. Bex, and P.M. Midgley, Eds. Cambridge University Press, pp. 659-740.
- Ni, K., J. R. Köster, A. Seidelc, A. Pacholski, Field measurement of ammonia emissions after nitrogenfertilization—A comparison between micrometeorological andchamber methods (2015), *Europ. J. Agronomy*, 71, 115–122
- NOAA (2018), Halocarbon and other atmospheric trace gases: Nitrous oxide (N₂O) – data set. <https://www.esrl.noaa.gov/gmd/hats/combined/N2O.html>.
- Nyamadzawo, G., M. Wuta, J. Nyamangara, J. L. Smith, and R. M. Rees (2014), Nitrous oxide and methane emissions from cultivated seasonal wetland (dambo) soils with inorganic, organic and integrated nutrient management, *Nutr Cycl Agroecosyst*, 100:161–175, DOI 10.1007/s10705-014-9634-9.
- Oertel, C., M. Jörg, Z. Kamal , Z. Frank, E. Stefan, (2016), Greenhouse gas emissions from soils: A review, *Chemie der Erde - Geochemistry*, 76(3), 327-352.
- Olivier, J. G. J. et al. (1998), Global air emission inventories for anthropogenic sources of NO_x, NH₃ and N₂O in 1990, *Environmental Pollution* 102, 135-148.
- Passianoto, C. C., T. Ahrens, B. J. Feigl, P. A. Steudler, J. B. do Carmo, and J. M. Melillo (2003), Emissions of CO₂, N₂O, and NO in conventional and no-till management practices in Rondonia, Brazil, *Biol. Fertil. Soils*, 38, 200-C208.
- Pathak, H., A. Bhatia, S. Prasad, S. Singh, S. Kumar, M. C. Jain, and U. Kumar (2002), Emission of Nitrous Oxide from Rice-Wheat Systems of Indo-Gangetic Plains of India, *Environmental Monitoring and Assessment*, 77, 163–178.
- Paulot F., D. J. Jacob, R. W. Pinder, J. O. Bash, K. Travis, D. K. Henze (2014), Ammonia emissions in the United States, European Union, and China derived by high - resolution inversion of ammonium wet deposition data: Interpretation with a new agricultural emissions inventory (MASAGE_NH₃), *Journal of Geophysical Research: Atmospheres*, 119, 4343-4364.
- Pelster, D. E., M. H. Chantigny, P. Rochette, D. A. Angers, C. Rieux, and A. Vanasse (2012), Nitrous Oxide Emissions Respond Differently to Mineral and Organic Nitrogen Sources in Contrasting Soil Types, *J. Environ. Qual.*, 41, 427–435, doi:10.2134/jeq2011.0261.

- Pilegaard K. (2013). Processes regulating nitric oxide emissions from soils. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences*, 368(1621), 20130126. doi:10.1098/rstb.2013.0126
- Poth, Mark; Anderson, Iris Cofman; Miranda, Heloisa Sinatora; Miranda, Antonia Carlos; Riggan, Philip J. 1995. The magnitude and persistence of soil NO, N₂O, CH₄, and CO₂ fluxes from burned tropical savanna in Brazil. *Global Biogeochemical Cycles*. 9(4): 503-513.
- Rashti, M. R., W. Wang, P. Moody, C. Chen, and H. Ghadiri (2015), Fertilizer-induced nitrous oxide emissions from vegetable production in the world and the regulating factors: A review, *Atmospheric Environment*, 112, 225-233.
- Ravishankara, A. R., J. S., Daniel, R. W. Portmann (2009), Nitrous oxide (N₂O): The dominant ozone-depleting substance emitted in the 21st century, *Science*, 326, 123-125.
- Reis, S., R. W. Pinder, M. Zhang, G. Lijie, and M. A. Sutton, Reactive nitrogen in atmospheric emission inventories (2009), *Atmos. Chem. Phys.*, 9, 7657–7677, 2009
- Riddick, S. N., et al. “Measurement of ammonia emissions from temperate and polar seabird colonies.”. *Atmospheric Environment* 134 (2016): , 134, 40-50. Print.,
- Rodhe, L., M. Pell, S. Yamulki, Nitrous oxide, methane and ammonia emissions following slurry spreading on grassland (2006), *Soil Use and Management*, 22, 229–237 doi: 10.1111/j.1475-2743.2006.00043.x
- Roelcke, M., S.X. Li, X.H. Tian, Y.J. Gao, J. Richter, In situ comparisons of ammonia volatilization from N fertilizers in Chinese loess soils (2002), *Nutrient Cycling in Agroecosystems*, 62: 73–88.
- Roelle, P, A., Viney P. Aneja, B. Gay, C. Geron, T (2001). Biogenic nitric oxide emissions from cropland soils, *Atmospheric Environment*, Volume 35, Issue 1, Pages 115-124.
- Rondón, A., C. Johansson, and L. Granat (1993), Dry deposition of nitrogen dioxide and ozone to coniferous forests, *J. Geophys. Res.*, 98(D3), 5159–5172, doi:10.1029/92JD02335.
- Salazar, F., J. Martínez-Lagos, M. Alfaro, T. Misselbrook, Ammonia emissions from urea application to permanent pasture on a volcanic soil (2012), *Atmospheric Environment* 61, 395-399

- Salazar, F., J. Martínez-Lagos, M. Alfaro, T. Misselbrook, Ammonia emission from a permanent grassland on volcanic soil after the treatment with dairy slurry and urea (2014), *Atmospheric Environment* 95, 591-597
- Sanhueza, E. and Santana, M. (1994), Atmospheric Wet Depositions in Tropical America. *Isr. J. Chem.*, 34: 327-334. doi:10.1002/ijch.199400036
- Schlesinger, W. H. and E. S. Bernhardt (2013), *Biogeochemistry: An analysis of global change*. 3rd edition. Academic Press, London.
- Schindlbacher A, Zechmeister-Boltenstern S, Butterbach-Bahl K. (2004). Effects of soil moisture and temperature on NO, NO₂, and N₂O emissions from European forest soils. *J. Geophys. Res.* 109, d1730210.1029/2004JD004590
- Sharma, S.K., Choudhury, A., Sarkar, P., Biswas S., et al., 2011. Greenhouse gas inventory estimates for India. *Current Science*, 101(3), 1-11.
- Shcherbak, I, N. Millar, and G. P. Robertson (2014), Global metaanalysis of the nonlinear response of soil nitrous oxide (N₂O) emissions to fertilizer nitrogen, *PNAS* 111(25), 9199-9204.
- Shimizu M., R. Hatano, T. Arita, Y. Kouda, A. Mori, S. Matsuura, M. Niimi, T. Jin, A. R. Desyatkin, O. Kawamura, M. Hojito, and A. Miyata (2013) The effect of fertilizer and manure application on CH₄ and N₂O emissions from managed grasslands in Japan, *Soil Science and Plant Nutrition*, 59(1), 69-86, DOI: 10.1080/00380768.2012.733926
- Smith P., M. Bustamante, H. Ahammad, H. Clark, H. Dong, E.A. Elsiddig, H. Haberl, R. Harper, J. House, M. Jafari, O. Masera, C. Mbow, N.H. Ravindranath, C.W. Rice, C. Robledo Abad, A. Romanovskaya, F. Sperling, and F. Tubiello, 2014: Agriculture, Forestry and Other Land Use (AFOLU). In: *Climate Change 2014: Mitigation of Climate Change. Contribution of Working Group III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* [Edenhofer, O., R. Pichs-Madruga, Y. Sokona, E. Farahani, S. Kadner, K. Seyboth, A. Adler, I. Baum, S. Brunner, P. Eickemeier, B. Kriemann, J. Savolainen, S. Schlömer, C. von Stechow, T. Zwickel and J.C. Minx (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Sosulski, T., E. Szara, W. Stępień, and M. Szymańska (2014), Nitrous oxide emissions from the soil under different fertilization systems on a long-term experiment, *Plant Soil Environ.*, 60(11), 481-488.

- Stehfest, E., and L. Bouwman (2006), N₂O and NO emission from agricultural fields and soils under natural vegetation: summarizing available measurement data and modeling of global annual emissions, *Nutrient Cycling in Agroecosystems*, 74, 207-228, DOI:10.1007/s10705-006-9000-7
- Tubiello, F. N., M. Salvatore, S. Rossi, A. Ferrara, N. Fitton, and P. Smith (2013), The FAOSTAT database of greenhouse gas emissions from agriculture. *Environmental Research Letters*, 8, 1-11. doi: 10.1088/1748-9326/8/1/015009, ISSN: 1748-9326.
- Vallejo, A., L. Garcia-Torres, J. A. Diez, A. Arce, and S. Lopez-Fernandez (2005), Comparison of N losses (NO₃⁻, N₂O, NO) from surface applied, injected or amended (DCD) pig slurry of an irrigated soil in a Mediterranean climate, *Plant Soil*, 272, 313–25.
- Valente, R. J., and F. C. Thornton (1993), Emissions of NO from soil at a rural site in central Tennessee, *J. Geophys. Res.*, 98, 16745-C16753.15, Guenzi, W. D., G. L. Hutchinson, and W. E. Beard (1994), Nitric and nitrous oxide emissions and soil nitrate distribution in a center-pivot-irrigated cornfield, *J. Environ. Qual.* 23, 483-C487.
- Veldkamp, E., M. Keller, and M. Nuñez (1998), Effects of pasture management on N₂O and NO emissions from soils in the humid tropics of Costa Rica, *Global Biogeochem. Cycles*, 12(1), 71–79, doi:10.1029/97GB02730.
- Wang, J., Z. Chen, Y. Ma, L. Sun, Z. Xiong, Q. Huang, and Q. Sheng (2013), Methane and nitrous oxide emissions as affected by organic–inorganic mixed fertilizer from a rice paddy in southeast China, *J Soils Sediments*, 13, 1408-1417, DOI 10.1007/s11368-013-0731-1.
- Wang, Y. J. Guo, R. D. Vogt, J. Mulder, J. Wang, X. Zhang (2017), Soil pH as the chief modifier for regional nitrous oxide emissions: New evidence and implications for global estimates and mitigation, *Global Change Biology*, 24, 617-626.
- Warneck, P., *Chemistry of the Natural Atmosphere* Academic Press, San Diego (1988)
- Warner, J. X., Dickerson, R. R., Wei, Z., Strow, L. L., Wang, Y., & Liang, Q. Increased atmospheric ammonia over the world's major agricultural areas detected from space. (2017). *Geophysical Research Letters*, 44, 2875–2884. <https://doi.org/10.1002/2016GL072305>
- Watanabe, A., K. Ikeya, N. Kanazaki, S. Makabe, Y. Sugiura, A. Shibata (2014), Five crop seasons' records of greenhouse gas fluxes from upland fields with repetitive applications of biochar and cattle manure, *Journal of Environmental Management*, 144, 168-175.

- Webb, J., D. Chadwick, and S. Ellis, Emissions of ammonia and nitrous oxide following incorporation into the soil of farmyard manures stored at different densities (2004), *Nutrient Cycling in Agroecosystems* 70: 67–76
- Williams, E. J., and F. C. Fehsenfeld (1991), Measurement of soil nitrogen oxide emissions at three North American ecosystems, *J. Geophys. Res.*, 96, 1033-C1042
- Wolf, U., R. Fu, F. Hoppner, H. Flessa, Contribution of N₂O and NH₃ to total greenhouse gas emission from fertilization: results from a sandy soil fertilized with nitrate and biogas digestate with and without nitrification inhibitor (2014), *Nutr Cycl Agroecosyst*, 100:121–134, DOI 10.1007/s10705-014-9631-z
- Yamulki, S., K. W. T. Goulding, C. P. Webster, and R. M. Harrison (1995), Studies on NO and N₂O fluxes from a wheat field, *Atmos. Environ.*, 29, 1627-C1635
- Yang W., A. Zhu, J. Zhang, X. Xin, X. Zhang, Evaluation of a backward Lagrangian stochastic model for determining surface ammonia emissions (2017), *Agricultural and Forest Meteorology* 234–235, 196–202
- Yang W., A. Zhu, J. Zhang, Y. Zhang, X. Chen, Y. He, L. Wang, An inverse dispersion technique for the determination of ammonia emissions from urea-applied farmland (2013), *Atmospheric Environment*, 217-224
- Yang, Y., C. Zhou, N. Li, K. Han, Y. Meng, X. Tian, L. Wang, Effects of conservation tillage practices on ammonia emissions from Loess Plateau rain-fed winter wheat fields (2015), *Atmospheric Environment* 104, 59-68
- Zheng, X., Y. Huang, Y. Wang, and M. Wang (2003), Seasonal characteristics of nitric oxide from a typical Chinese rice-wheat rotation during the non-waterlogged period, *Global Change Biol.*, 9, 219-C229
- Zhai, L., H. Liu, J. Zhang, J. Huang, and B. Wang (2011), Long-Term Application of Organic Manure and Mineral Fertilizer on N₂O and CO₂ Emissions in a Red Soil from Cultivated Maize-Wheat Rotation in China, *Agricultural Sciences in China*, 10(11), 1748-1757.
- Zhou, F., Z. Shang, P. Ciais, S. Tao, S. Piao, P. Raymond, C. He, B. Li, R. Wang, X. Wang, S. Peng, Z. Zeng, H. Chen, N. Ying, X. Hou, and P. Xu (2014). A new high-resolution N₂O emission inventory for China in 2008, *Environmental Science and Technology*, 48, 8538-8547.

Zhou, M., B. Zhu, N. Brügemann, J. Bergmann, Y. Wang, and K. Butterbach-Bahl (2014), N₂O and CH₄ Emissions, and NO₃⁻ Leaching on a Crop-Yield Basis from a Subtropical Rain-fed Wheat–Maize Rotation in Response to Different Types of Nitrogen Fertilizer, *Ecosystems*, 17, 286–301, DOI: 10.1007/s10021-013-9723-7

Zhuang, X. L., Y. Wang, H. He, J. Liu, X. Wang, T. Zhu, M. Ge, J. Zhou, G. Tang, J. Ma, Haze insights and mitigation in China: An overview (2014), *Journal of Environmental Sciences*, Volume 26, Issue 1, 2-12